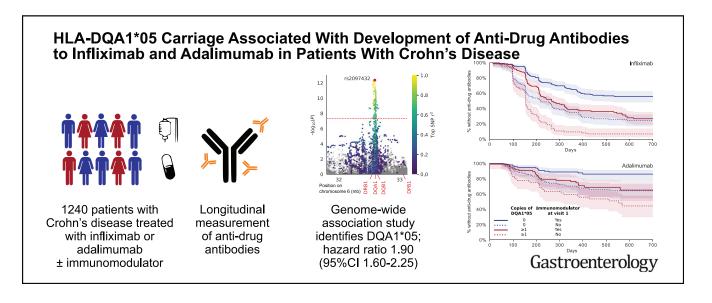
# HLA-DQA1\*05 Carriage Associated With Development of Anti-Drug Antibodies to Infliximab and Adalimumab in Patients With Crohn's Disease



Aleksejs Sazonovs,<sup>1</sup> Nicholas A. Kennedy,<sup>2,3</sup> Loukas Moutsianas,<sup>1</sup> Graham A. Heap,<sup>2,4</sup> Daniel L. Rice,<sup>1</sup> Mark Reppell,<sup>4</sup> Claire M. Bewshea,<sup>3</sup> Neil Chanchlani,<sup>2,3</sup> Gareth J. Walker,<sup>2,3</sup> Mandy H. Perry,<sup>5</sup> Timothy J. McDonald,<sup>5</sup> Charlie W. Lees,<sup>6</sup> J. R. Fraser Cummings,<sup>7,8</sup> Miles Parkes,<sup>9</sup> John C. Mansfield,<sup>10</sup> Peter M. Irving,<sup>11</sup> Jeffrey C. Barrett,<sup>1</sup> Dermot McGovern,<sup>12</sup> James R. Goodhand,<sup>2,3</sup> Carl A. Anderson,<sup>1</sup> Tariq Ahmad,<sup>2,3</sup> and PANTS Consortium

<sup>1</sup>Wellcome Sanger Institute, Wellcome Genome Campus, Hinxton, UK; <sup>2</sup>Department of Gastroenterology, Royal Devon and Exeter Hospital National Health Service Foundation Trust, Exeter, UK; <sup>3</sup>Exeter Inflammatory Bowel Disease and Pharmacogenetics Research Group, University of Exeter, Exeter, UK; <sup>4</sup>AbbVie Inc, North Chicago, Illinois; <sup>5</sup>Department of Blood Science, Royal Devon and Exeter Hospital National Health Service Foundation Trust, Exeter, UK; <sup>6</sup>Department of Gastroenterology, Western General Hospital, National Health Service Lothian, Edinburgh, UK; <sup>7</sup>Department of Gastroenterology, University Hospital Southampton National Health Service Foundation Trust, Southampton, UK; <sup>8</sup>Faculty of Experimental Medicine, University of Southampton, Southampton, UK; <sup>9</sup>Department of Gastroenterology, Addenbrooke's Hospital, Cambridge University Hospitals National Health Service Foundation Trust, Cambridge, UK; <sup>10</sup>Department of Gastroenterology, Newcastle Upon Tyne Hospitals National Health Service Foundation Trust, Newcastle upon Tyne, UK; <sup>11</sup>Department of Gastroenterology, Guy's and St Thomas' NHS Foundation, Trust, London, UK; and <sup>12</sup>F. Widjaja Foundation Inflammatory Bowel and Immunobiology Research Institute, Cedars-Sinai Medical Center, Los Angeles, California



BACKGROUND & AIMS: Anti-tumor necrosis factor (anti-TNF) therapies are the most widely used biologic drugs for treating immune-mediated diseases, but repeated administration can induce the formation of anti-drug antibodies. The ability to identify patients at increased risk for development of anti-drug antibodies would facilitate selection of therapy and use of preventative strategies. METHODS: We performed a genome-wide association study to identify variants associated with time to development of anti-drug antibodies in a discovery cohort of 1240 biologic-naïve patients with Crohn's disease starting infliximab or adalimumab therapy. Immunogenicity was defined as an anti-drug antibody titer ≥10 AU/mL using a drug-tolerant enzyme-linked immunosorbent assay. Significant association signals were confirmed in a replication cohort of

178 patients with inflammatory bowel disease. **RESULTS:** The HLA-DQA1\*05 allele, carried by approximately 40% of Europeans, significantly increased the rate of immunogenicity (hazard ratio [HR], 1.90; 95% confidence interval [CI], 1.60–2.25;  $P=5.88\times10^{-13}$ ). The highest rates of immunogenicity, 92% at 1 year, were observed in patients treated with infliximab monotherapy who carried HLA-DQA1\*05; conversely the lowest rates of immunogenicity, 10% at 1 year, were observed in patients treated with adalimumab combination therapy who did not carry HLA-DQA1\*05. We confirmed this finding in the replication cohort (HR, 2.00; 95% CI, 1.35–2.98;  $P=6.60\times10^{-4}$ ). This association was consistent for patients treated with adalimumab (HR, 1.89; 95% CI, 1.32–2.70) or infliximab (HR, 1.92; 95% CI, 1.57–2.33), and for patients treated with anti-TNF therapy alone

(HR, 1.75; 95% CI, 1.37–2.22) or in combination with an immunomodulator (HR, 2.01; 95% CI, 1.57–2.58). **CONCLUSIONS:** In an observational study, we found a genomewide significant association between HLA-DQA1\*05 and the development of antibodies against anti-TNF agents. A randomized controlled biomarker trial is required to determine whether pretreatment testing for HLA-DQA1\*05 improves patient outcomes by helping physicians select anti-TNF and combination therapies. ClinicalTrials.gov ID: NCT03088449.

Keywords: PANTS; GWAS; Loss Of Response; Drug Persistence.

Anti-tumor necrosis factor (anti-TNF) therapies are the most widely used biologics for treating immune-mediated diseases and in 2018 accounted for a global expenditure in excess of \$23.5 billion. Repeated administration often induces the formation of anti-drug antibodies (immunogenicity), leading to treatment failure.

Immunogenicity is more common in patients treated with infliximab (a murine-human chimeric monoclonal antibody) than adalimumab (a fully human monoclonal antibody) and is a major cause of low anti-TNF drug level, infusion reactions, and non-remission in patients with Crohn's disease. Combination immunomodulator therapy reduces the risk of immunogenicity to both adalimumab and infliximab, and for infliximab, improves treatment outcomes. Despite these benefits, many patients are still treated with anti-TNF monotherapy because of concerns about the increased risk of adverse drug reactions, opportunistic infections, and malignancies associated with combination therapy with immunomodulators.

The ability to identify patients at increased risk of immunogenicity may influence the choice of anti-TNF treatment and the use of preventative strategies, including combination therapy with immunomodulators. However, our understanding of the cellular and molecular mechanisms underpinning immunogenicity to biologics is limited. Retrospective candidate gene studies have suggested variants in *FCGR3A*<sup>10</sup> and *HLA-DRB1*<sup>11,12</sup> increase susceptibility to immunogenicity to anti-TNF therapy. These associations did not achieve genome-wide significance and are yet to be independently replicated. Here, we report the first genetic locus robustly associated with immunogenicity to anti-TNF therapies.

### **Methods**

The PANTS (Personalising Anti-TNF Therapy in Crohn's Disease) study is a United Kingdom-wide, multicenter, prospective observational cohort reporting the treatment failure rates of the anti-TNF drugs infliximab (originator, Remicade [Merck Sharp & Dohme, Hertfordshire, UK] and biosimilar, CT-P13 [Celltrion, Incheon, South Korea]), and adalimumab (Humira [AbbVie, Cambridge, MA]) in 1610 anti-TNF-naïve patients with Crohn's disease.<sup>2</sup>

At inclusion, subjects were aged 6 years or older and had active luminal Crohn's disease involving the colon and/or small intestine. Choice of anti-TNF drug and use of concomitant

### WHAT YOU NEED TO KNOW

### BACKGROUND AND CONTEXT

Anti-tumor necrosis factor (anti-TNF) therapies are the most widely used biologic drugs for treating immune-mediated diseases, but repeated administration can induce the formation of anti-drug antibodies, increasing the risk of treatment failure.

### **NEW FINDINGS**

In a genome-wide association study, we identified a variant (HLA-DQA1\*05) that increases risk of development of antibodies against infliximab and adalimumab 2-fold in patients with Crohn's disease, regardless of concomitant immunomodulator use.

### LIMITATIONS

The study was limited to patients of European descent, 40% of whom carry the risk allele.

### IMPACT

Testing patients for HLA-DQA1\*05 might help physicians decide whether they should receive anti-TNF and combination immunomodulator therapy.

immunomodulator therapy was at the discretion of the treating physician as part of usual care. Patients were initially studied for 12 months or until drug withdrawal. In the first year, study visits were scheduled at first dose, post-induction (weeks 12–14), weeks 30 and 54, and at treatment failure. For infliximabtreated patients, additional visits occurred at each infusion. After 12 months, patients were invited to continue follow-up for an additional 2 years. Drug persistence was defined as the duration of time from initiation of anti-TNF therapy to exit from the study due to treatment failure. Patients who exited the study for other reasons, declined to participate in the 2-year extension, or were lost to follow-up were censored at the time of last drug dose or study visit.

At each visit, serum infliximab or adalimumab drug and anti-drug antibody levels were analyzed using total antibody enzyme linked immunosorbent assays. See the Supplementary Material for a detailed description of drug and anti-drug antibody testing. The total antibody, unlike the more commonly reported free antibody assay, includes a drugantibody disassociation step that allows the assessment of anti-drug antibodies in the presence of drug. We defined immunogenicity as an anti-drug antibody concentration of  $\geq 10$  AU/mL, irrespective of drug level, at 1 or more time points.

DNA was extracted from pretreatment blood samples from 1524 individuals in the PANTS cohort and genotyping undertaken using the Illumina CoreExome microarray (Illumina, San Diego, CA). See the Supplementary Material for a detailed description of the genetic analyses. We excluded individuals of

Abbreviations used in this paper: AIC, Akaike information criterion; CI, confidence interval; HR, hazard ratio; IBD, inflammatory bowel disease; TNF, tumor necrosis factor; UC, ulcerative colitis.



© 2020 by the AGA Institute. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

non-European ancestry (using principal component analysis; see Supplementary Figure 1), 1 individual from each related pair (defined as a pi-hat >.187), and those with an outlying number of missing or heterozygous genotypes. After quality control, 1323 individuals remained in the study, of which 1240 had drug and anti-drug antibody level data available (Supplementary Figure 2). With its case–control design, our study had >80% power to detect genome-wide significant evidence of association ( $\alpha=5\times10^{-8}$ ) to variants with a minor allele frequency >22% and a relative risk >1.4 (Supplementary Figure 3). Baseline demographics for these patients are shown in Supplementary Table 1.

Variants with a genotype call rate of <95% or with significant evidence of deviation from Hardy-Weinberg equilibrium ( $P < 1 \times 10^{-10}$ ) were excluded. Single-nucleotide polymorphisms were imputed via the Sanger Imputation Service using the Haplotype Reference Consortium panel, and 7,578,947 variants with an information content metric score  $\ge 0.4$  were subsequently taken forward for analysis. HLA types were imputed at 2- and 4-digit resolution for the following loci: HLA-A, HLA-C, HLA-B, HLA-DRB1, HLA-DQA1, HLA-DQB1, and HLA-DPB1. Long-read sequencing of these HLA alleles was undertaken to assess the accuracy of our imputation.

We assembled an independent cohort to replicate significant findings from the discovery cohort. This comprised 107 Crohn's disease, 64 ulcerative colitis, and 7 inflammatory bowel disease (IBD)–type unclassified patients with cross-sectional drug and anti-drug antibody levels measured as part of routine clinical practice. The samples were genotyped using either the Illumina CoreExome array (n = 164)<sup>16</sup> or the Affymetrix 500k array (n = 14; ThermoFisher Scientific, Waltham, MA).<sup>17</sup> Quality control and imputation methods were the same as in the discovery cohort.

# Statistical Analysis

Rates of immunogenicity were estimated using the Kaplan-Meier method. Clinical outcomes and genetic association tests with time to anti-drug antibody development were performed using multivariable Cox proportional hazards regression: sex, drug type (infliximab or adalimumab), immunomodulator use, and the first within-sample principal component were included as covariates (Supplementary Table 2). Patients who did not develop immunogenicity during the study were censored at the point of last observation. Post-hoc sensitivity analyses were undertaken to test our genetic findings with immunogenicity, firstly, at progressively higher antibody thresholds; secondly, to simulate a free-antibody assay; and thirdly, excluding patients with a single anti-drug antibody level  $\geq 10~{\rm AU/mL}$  and subsequent negative anti-drug antibodies  $< 10~{\rm AU/mL}$ .

The Akaike information criterion (AIC) was used to compare non-nested models to assess if the mode of inheritance was dominant or additive, and to determine whether HLA allele group, specific HLA alleles, or amino-acid sequence best explained the association. The fixed-effects Q statistic was used to perform tests of heterogeneity of effect; this test is an extension of Cochran's Q-test and examines whether the observed effect size variability is larger than expected by chance. Interaction tests of the differential effects of drug type (infliximab vs adalimumab and Remicade [Janssen

Biotech, Inc, Horsham, PA] vs CT-P13) and combination therapy (immunomodulator vs no immunomodulator) conditional on the genotype were performed. Mann-Whitney U tests were used to compare serum levels of anti-drug antibodies at week 54 stratified by anti-TNF drug and immunomodulator use.

### **Ethics**

The South West Research Ethics committee approved the study (Research Ethics Committee reference: 12/SW/0323) in January 2013. Patients were included after providing informed, written consent. The protocol is available online (www.ibdresearch.co.uk).

# Results

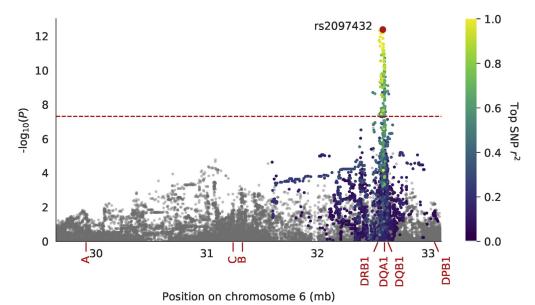
Within the first 12 months, 44% of patients developed anti-drug antibodies (95% confidence interval [CI], 0.41–0.48), and 62% of patients did so within 36 months (95% CI, 0.57–0.67). After correcting for immunomodulator use, the rate of immunogenicity was greater in patients treated with infliximab (n = 742) than adalimumab (n = 498) (hazard ratio [HR], 3.21; 95% CI, 2.61–3.95;  $P=1.18\times 10^{-28}$ ). In a model including drug type as a covariate, rates of immunogenicity were greater in patients treated with anti-TNF monotherapy (n = 544) compared to combination therapy with immunomodulators (n = 696) (HR, 2.30; 95% CI, 1.94–2.75;  $P<6.10\times 10^{-21}$ ).

# A Locus Within the HLA Region Is Associated With Time to Immunogenicity

We identified a genome-wide significant association on chromosome 6 with time to development of immunogenicity, with the most associated single-nucleotide polymorphism, rs2097432 (b38\_pos: 6:32622994; HR, 1.70; 95% CI, 1.48–1.94;  $P=4.24\times10^{-13}$ ), falling within the major histocompatibility complex region (Figure 1 and Supplementary Figures 4 and 5). We replicated this association in our independent cohort of 178 patients with IBD (HR, 1.69; 95% CI, 1.26–2.28;  $P=8.80\times10^{-4}$ ). A variant on chromosome 11, rs12721026 (b38\_pos: 11:116835452; HR, 0.46; 95% CI, 0.33–0.63;  $P=4.76\times10^{-8}$ ) also reached genome-wide significance in our discovery analysis, although the association was not replicated in our independent cohort (HR, 0.85; 95% CI, 0.49–1.44; P=.51).

### Fine-Mapping of the Signal in the HLA Region

At the HLA allele group level (2-digit resolution), only HLA-DQA1\*05 achieved genome-wide significance (HR, 1.90; 95% CI, 1.60–2.25;  $P=5.88\times 10^{-13}$ ) (Figure 2). At the specific allele level (4-digit resolution), no single allele reached genome-wide significance. The 2 most common HLA-DQA1\*05 subtype alleles, HLA-DQA1\*05:01 (HR, 1.57; 95% CI, 1.33–1.85;  $P=4.24\times 10^{-7}$ ) and HLA-DQA1\*05:05 (HR, 1.48; 95% CI, 1.24–1.78;  $P=5.54\times 10^{-5}$ ), had similar effects on time to immunogenicity and a model containing these two 4-digit alleles was virtually indistinguishable from a model including HLA-DQA1\*05 only (AIC<sub>05</sub> = 6659.07 vs AIC<sub>05:01&05:05</sub> = 6659.50). We did not identify any amino acids that better fit the data than HLA-DQA1\*05. We

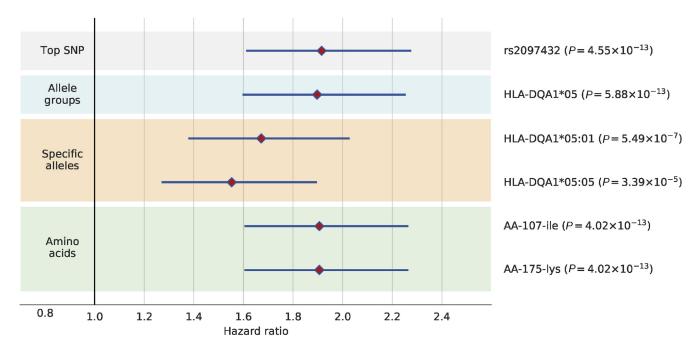


**Figure 1.** A regional plot of the association results with the major histocompatibility complex (MHC) region on chromosome 6. Midpoint positions of the HLA genes across the MHC region are shown in *red* on the *x-axis*. Single-nucleotide polymorphisms (SNPs) that passed the genome-wide significance threshold ( $P = 5 \times 10^{-8}$ ) are shown above the *red horizontal dashed line* with the most significant SNP in *red*. SNPs correlated with the lead SNP ( $r^2 > 0.05$ ) are color-coded from *purple* to *yellow*. Pairwise genotype correlation ( $r^2$ ) between SNPs was calculated using genotype data from the non-Finnish European population of the 1000 Genomes Project.

observed >99% concordance between imputed and sequenced HLA genotypes at *HLA-DQA1* (Supplementary Table 3).

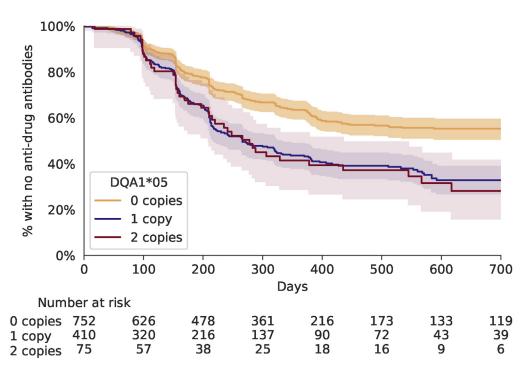
To formally assess the inheritance pattern of HLA-DQA1\*05 mediated immunogenicity, we compared the fit of additive and dominant models and found that the dominant model gave a better fit (AIC<sub>DOM</sub> = 6652.12 vs AIC<sub>ADD</sub> = 6659.07), and stronger association signal for HLA-DQA1\*05 (HR, 1.90; 95% CI, 1.60–2.25;  $P = 5.88 \times 10^{-13}$ ) (Figure 3

and Supplementary Table 4). We also looked for nonadditive effects across all other HLA alleles, but the model assuming a dominant effect for HLA-DQA1\*05 remained the best fit to the data. The HLA-DQA1\*05 association was confirmed in our replication cohort (entire cohort (HR, 2.00; 95% CI, 1.35–2.98;  $P = 6.60 \times 10^{-4}$ ), Crohn's disease-only subset (HR, 2.26; 95% CI, 1.33–3.84; P = .003), and ulcerative colitis-only subset of the replication cohort (HR, 2.02; 95% CI, 1.08–3.79; P = .03), again with a better fit for the



**Figure 2.** Effect sizes of the most strongly associated single-nucleotide polymorphisms (SNPs), HLA alleles, and amino acids on time to immunogenicity. *Blue lines* represent 95% Cls. Association test *P* values are shown in parentheses.

3. Kaplan-Meier **Figure** estimator showing the rate anti-drug antibody development, stratified by the number of HLA-DQA1\*05 alleles carried. Orange, blue, and red indicate 0, 1, and 2 copies DQA1\*05 allele, respectively. Carriers of 1 or 2 copies of the allele have a similar rate of immunogenicity development, and a dominant model is a better fit for the data than an additive model (AIC $_{DOM} = 6652.12$ vs  $AIC_{ADD} = 6659.07$ ). Xaxis truncated at 700 days, due to the low number of observations for longer time periods.



dominant model (AIC<sub>DOM</sub> = 942.51 vs AIC<sub>ADD</sub> = 944.81). After conditioning on HLA-DQA1\*05, we did not identify any secondary signals of association with time to immunogenicity within the major histocompatibility complex region (Supplementary Figure 6).

Sensitivity analyses showed that the effect size of HLA-DQA1\*05 carriage on immunogenicity was similar across subgroups (Figure 4A, 4B): firstly when the threshold for defining immunogenicity was increased from 10 AU/mL to  $\geq$ 150 AU/mL. Secondly when we simulated a drug-sensitive instead of a drug-tolerant assay, where immunogenicity was defined as an anti-drug antibody titer  $\geq$ 10 AU/mL without detectable drug (HR, 1.57; 95% CI, 1.23-2.01;  $P=3.66\times10^{-4}$ ). Thirdly when we removed patients with a one-off transient anti-drug antibody level  $\geq$ 10 AU/mL (HR, 1.94; 95% CI, 1.62-2.32;  $P=8.46\times10^{-13}$ ).

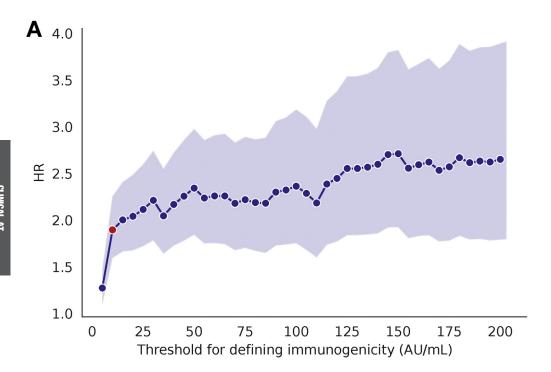
# The Effect of HLA-DQA1\*05 Across Drug and Treatment Regimes

While immunogenicity rates were lower with adalimumab-treated compared to infliximab-treated patients, we did not detect a significant difference in the effect of HLA-DQA1\*05 on the immunogenicity rate for these 2 drugs (HR, 1.89; 95% CI, 1.32–2.70 in adalimumab-, HR, 1.92; 95% CI, 1.57–2.33 in infliximab-treated patients;  $P_{\rm het}=.91$ ) (Figure 5). We also found no significant evidence for heterogeneity of effect of HLA-DQA1\*05 on immunogenicity between patients treated with the infliximab originator, Remicade, and its biosimilar CT-P13 ( $P_{\rm het}=.23$ ) (Supplementary Figure 7). Likewise, we did not detect any significant heterogeneity of effect of HLA-DQA1\*05 carriage on immunogenicity for individuals on monotherapy (HR, 1.75; 95% CI,

1.37–2.22) vs combination therapy (HR, 2.01; 95% CI, 1.57–2.58) with immunomodulators ( $P_{\text{het}} = .14$ ). In addition, we did not identify any significant interactions between HLA-DQA1\*05 and the clinical covariates (drug type: P = .83; mono- vs combination therapy: P = .71; Remicade vs CT-P13: P = .59).

The highest rates of immunogenicity, 92% at 1 year, were observed in patients treated with infliximab monotherapy who carried HLA-DQA1\*05 (Figure 6A). Conversely, the lowest rates of immunogenicity, 10% at 1 year, were observed in patients treated with adalimumab combination therapy who did not carry HLA-DQA1\*05 (Figure 6B). Our final model, which includes HLA-DQA1\*05 status, sex, drug, and immunomodulator usage, explained 18% of the variance in immunogenicity to anti-TNF in our cohort.

Having demonstrated that HLA-DQA1\*05 was associated with time to immunogenicity, we sought associations with anti-drug antibody titers after 1 year of treatment and subsequent non-persistence on drug. Carriage of HLA-DQA1\*05 was associated with higher maximal anti-drug antibody titers ( $P_{\text{infliximab}} = 8 \times 10^{-10}$ ;  $P_{\text{adalimumab}} = .002$ ). We observed lower drug persistence rates to year 3 in patients treated with an anti-TNF drug without an immunomodulator. HLA-DQA1\*05 carriage was associated with lower drug persistence only in patients treated with adalimumab without an immunomodulator (Figure 7A, 7B); the optimal model here used the interaction between immunomodulator use and HLA-DQA1\*05 (DQA1\*05: HR, 1.40; 95% CI, 1.08–1.80; P = .011, immunomodulator use: HR, 0.74; 95% CI, 0.58-0.94; P = .014, interaction between DQA1\*05 and immunomodulator use: HR, 0.65; 95% CI, 0.45-0.95; P = .026).



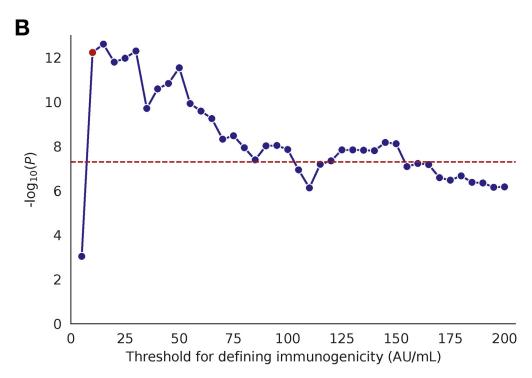
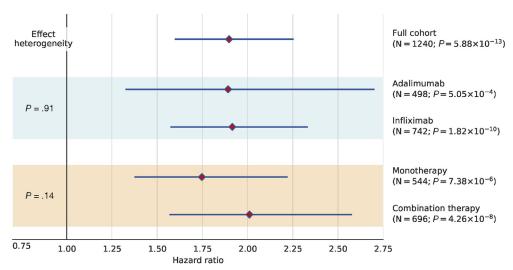


Figure 4. (A) Sensitivity analysis of the effect size of DQA1\*05 association and time to immunogenicity. (B) Sensitivity analysis of the significance of DQA1\*05 association and time to immunogenicity. In primary analyses, immunogenicity defined as an anti-drug antibody concentration >10 AU/mL, irrespective of drug concentration (red dot). We repeated the time to immunogenicity analysis varying this definition from  $\geq$ 5 to  $\geq$ 200 AU/mL.

# **Discussion**

Immunogenicity to biologic therapies is a major concern for patients, regulatory authorities, and the pharmaceutical industry. We report the first genomewide significant association with immunogenicity to anti-TNF therapy using the largest prospective cohort study of infliximab and adalimumab in Crohn's disease. We have demonstrated that carriage of 1 or more HLA-DQA1\*05 alleles confers an almost 2-fold risk of immunogenicity to anti-TNF therapy, irrespective of

concomitant immunomodulator use or drug type (infliximab [Remicade or CT-P13] or adalimumab). Finemapping and confirmatory sequencing of the HLA identified that the specific alleles HLA-DQA1\*05:01 and HLA-DQA1\*05:05 mediated most of this risk. Carriage of HLA-DQA1\*05 was associated with higher anti-drug antibody levels and lower drug persistence rates, although further studies are needed to more accurately quantify the relationship between HLA-DQA1\*05 and the risk of anti-TNF treatment failure.



**Figure 5.** HLA-DQA1\*05 has a consistent effect on immunogenicity in different patient subgroups. We repeated the proportional hazard association analysis, separating the full cohort into subgroups by drug and therapy type. Estimated HRs and SEs between the pairings were compared using a heterogeneity of effects test (P > .05), suggesting that the effect of DQA1\*05 on immunogenicity is not affected by these clinical covariates. *Blue lines* represent 95% Cls. Association test P values are shown in parentheses.

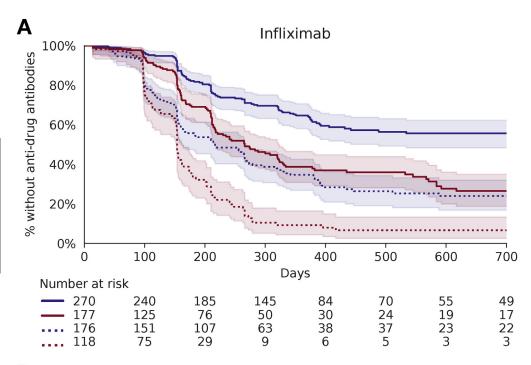
Based on our data in patients with luminal Crohn's disease, all patients treated with an anti-TNF should be prescribed an immunomodulator to lower the risk of immunogenicity. We hypothesize that for patients who carry HLA-DQA1\*05 in whom immunomodulators are contraindicated or not tolerated, clinicians might advise against the use of anti-TNF drugs, particularly infliximab. This is because 90% of patients who carry HLA-DQA1\*05 who are treated with infliximab monotherapy have evidence of immunogenicity by week 54. In contrast, patients who do not carry HLA-DQA1\*05 might be given the choice between adalimumab or infliximab combination therapy. Patients without the risk allele and a history of adverse drug reactions to thiopurines and/or methotrexate, or who are at high risk of opportunistic infections, might be spared the additional risks of combination therapy and treated with adalimumab monotherapy. A randomized controlled biomarker trial is required to explore these hypotheses and confirm whether HLA-DQA1\*05 testing may help direct treatment choices in order to improve clinical outcomes.

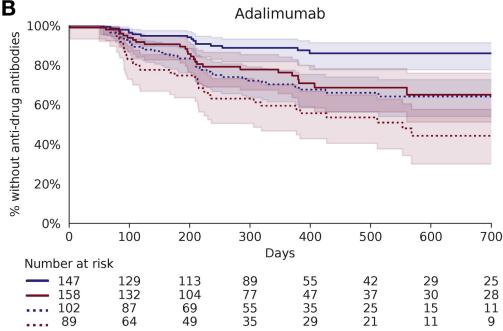
The shared genetic association between HLA-DQA1\*05 and immunogenicity to infliximab and adalimumab may explain the widely reported diminishing returns of switching between anti-TNF therapies at the time of loss of response. <sup>18,19</sup> If the immunogenic effect of HLA-DQA1\*05 extends to other therapeutic antibodies, then subjects who carry the variant may be candidates for non-antibody modality therapies, such as small molecule drugs. Allelic variation in the HLA-DQA1 gene has been linked to aberrant adaptive immune responses. The HLA class II gene HLA-DQA1 is expressed by antigen presenting cells and encodes the  $\alpha$ -chain of the HLA-DQ heterodimer that forms part of the antigen-binding site where epitopes are presented to T-helper cells. Relevant to immunogenicity, carriage of HLA-DQA1\*05 has been associated with celiac

disease, type 1 diabetes, and protection against rheumatoid arthritis and pulmonary tuberculosis. <sup>20–24</sup> Several hypotheses have been proposed, but exactly how specific HLA alleles contribute to disease pathogenesis or, in this case, increased immunogenicity, remains unknown.

HLA-DQ1A\*05 may serve as a useful biomarker of immunogenicity risk and may impact how the next-generation of anti-TNF drugs are designed to minimize HLA-DQA1\*05 mediated immunogenicity. Previous studies have shown that it is possible to map and eliminate potential immunogenic T-cell epitopes with the aim of producing safer and more durable biologic drugs. <sup>25,26</sup> However, caution needs to be exercised to ensure protein sequence modifications designed to reduce the risk of immunogenicity to patients carrying HLA-DQA1\*05 do not put a different group of patients at risk.

Multiple assays are available to detect anti-drug antibodies and there is no universally accepted, validated threshold to diagnose immunogenicity. We deliberately chose a total or drug-tolerant assay that permits the measurement of anti-drug antibodies in the presence of drug in order to minimize the number of false-negative patients assigned to the control group. We then validated the manufacturer's positivity threshold in independent experiments in 500 drug-naïve controls and confirmed that the recommend cutoff of 10 AU/mL corresponds to the 99th centile of the anti-drug antibody titer distribution. In support of this threshold, we have recently demonstrated that even modestly elevated anti-drug antibodies levels (10-30 AU/ mL) at weeks 14 and 54 of treatment are associated with lower drug levels at these time points, and non-remission at week 54.2 In addition, sensitivity analyses confirmed that the association and effect size between HLA-DQA1\*05 and immunogenicity remained at progressively higher diagnostic thresholds for immunogenicity, when we simulated a





**6.** (*A*) Figure Anti-drug antibody developmentinfliximab. (B) Anti-drua antibody developmentadalimumab. Kaplan-Meier estimator showing the rate of anti-drug antibody development (A, B) stratified by carriage of HLA-DQA1\*05 alleles and treatment regime. Dotted lines indicate patients undergoing anti-TNF monotherapy: solid lines indicate combination therapy with immunomodulators. Red indicates carriers of the HLA-DQA1\*05 allele (1 or 2 copies); blue indicates noncarriers. For both drugs and treatment regimes, immunogenicity rates are higher for HLA-DQA1\*05 carriers. The xaxis was truncated at 700 days due to the low number of observations.

free-assay, and when we removed patients with transient antibodies. Finally, HLA-DQA1\*05 was associated with the quantitative trait of maximal anti-drug antibody titer.

We acknowledge several important limitations of this study. Firstly, we may have underestimated the contribution of HLA-DQA1\*05 to immunogenicity because of the short duration of follow-up in patients who did not continue in the study beyond the first year. Secondly, because we designed a schedule of visits to minimize patients' inconvenience, there were fewer assessments for those treated with adalimumab than infliximab. As a result, we might have

underestimated rates of immunogenicity among adalimumab-treated patients.

Our genome-wide association study was limited to patients with luminal Crohn's disease of European descent. Given that HLA-DQA1\*05 is not associated with Crohn's disease risk,<sup>27</sup> the percentage of carriers among our patients (39%) was similar to that reported in an independent British population cohort (38%).<sup>28</sup> As such, we hypothesize that HLA-DQA1\*05 will make a similar contribution to anti-TNF immunogenicity in other patient populations where the allele is not associated with disease susceptibility

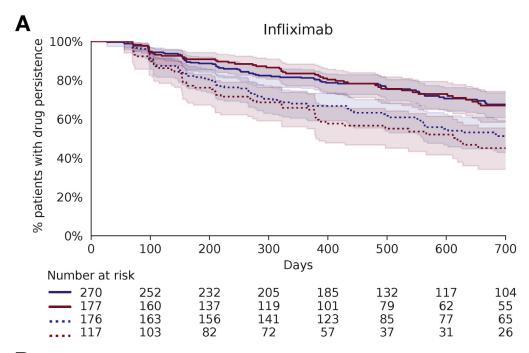
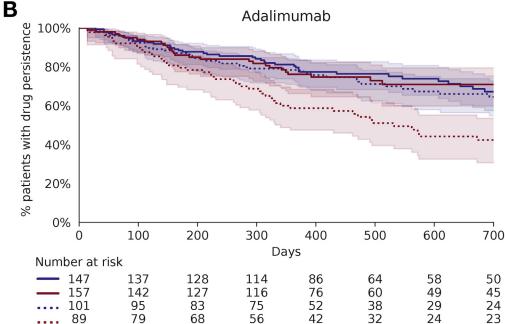


Figure 7. (A) Drug persistence-infliximab. (B) Drug persistence—adalimumab. Kaplan-Meier estimator showing the rate of drug persistence (A, B) stratified carriage HLAby of DQA1\*05 alleles and treatment regime. Dotted lines indicate patients undergoing anti-TNF monosolid therapy; lines indicate combination therapy with immunomodulators. Red indicates carriers of the HLA-DQA1\*05 allele (1 or 2 copies); blue indicates noncarriers. The xaxis was truncated at 700 days due to the low number of observations.



(eg, ankylosing spondylitis and perianal Crohn's disease). Due to the wide variation in the frequency of HLA-DQA1\*05 across ethnic groups, <sup>28</sup> further studies are required to assess the contribution of HLA-DQA1\*05 to immunogenicity across populations. Whether HLA-DQA1\*05 is also associated with immunogenicity to other biologic drugs also needs to be determined.

We report the first genome-wide significant association with immunogenicity to biologic drugs. Carriage of HLA-DQA1\*05 almost doubles the rate of anti-TNF anti-drug antibody development, independent of immunomodulator

use, for both infliximab and adalimumab. To minimize the risk of immunogenicity, pretreatment genetic testing for HLA-DQA1\*05 may help personalize the choice of anti-TNF and the need for combination therapy with an immunomodulator.

# **Supplementary Material**

Note: To access the supplementary material accompanying this article, visit the online version of *Gastroenterology* at www.gastrojournal.org, and at https://doi.org/10.1053/j.gastro.2019.09.041.

# References

- IQVIA Institute. The Global Use of Medicine in 2019 and Outlook to 2023. Available at: https://www.iqvia.com/-/media/iqvia/pdfs/institute-reports/the-global-use-of-medicine-in-2019-and-outlook-to-2023.pdf?\_ = 1557505804444. Accessed July 1, 2019.
- Kennedy NA, Heap GA, Green HD, et al. Predictors of anti-TNF treatment failure in anti-TNF-naive patients with active luminal Crohn's disease: a prospective, multicentre, cohort study. Lancet Gastroenterol Hepatol 2019;1253:1–13.
- Vermeire S, Gils A, Accossato P, et al. Immunogenicity of biologics in inflammatory bowel disease. Therap Adv Gastroenterol 2018;11:1756283X17750355.
- Colombel JF, Sandborn WJ, Reinisch W, et al. Infliximab, azathioprine, or combination therapy for Crohn's disease. N Engl J Med 2010;362:1383–1395.
- Panaccione R, Ghosh S, Middleton S, et al. Combination therapy with infliximab and azathioprine is superior to monotherapy with either agent in ulcerative colitis. Gastroenterology 2014;146:392–400.e3.
- Matsumoto T, Motoya S, Watanabe K, et al. Adalimumab monotherapy and a combination with azathioprine for Crohn's disease: a prospective, randomized trial. J Crohns Colitis 2016;10:1259–1266.
- Lichtenstein GR, Feagan BG, Cohen RD, et al. Drug therapies and the risk of malignancy in Crohn's disease: results from the TREAT<sup>TM</sup> Registry. Am J Gastroenterol 2014;109:212–223.
- 8. Osterman MT, Sandborn WJ, Colombel J-F, et al. Increased risk of malignancy with adalimumab combination therapy, compared with monotherapy, for Crohn's disease. Gastroenterology 2014;146:941–949.
- Lemaitre M, Kirchgesner J, Rudnichi A, et al. Association between use of thiopurines or tumor necrosis factor antagonists alone or in combination and risk of lymphoma in patients with inflammatory bowel disease. JAMA 2017;318:1679–1686.
- Romero-Cara P, Torres-Moreno D, Pedregosa J, et al. A FCGR3A polymorphism predicts anti-drug antibodies in chronic inflammatory bowel disease patients treated with anti-TNF. Int J Med Sci 2018;15:10–15.
- Billiet T, Casteele N Vande, Stappen T Van, et al. Immunogenicity to infliximab is associated with HLA-DRB1. Gut 2015;64:1344–1345.
- 12. Liu M, Degner J, Davis JW, et al. Identification of HLA-DRB1 association to adalimumab immunogenicity. PLoS One 2018;13:e0195325.
- 13. Perry M, Bewshea C, Brown R, et al. Infliximab and adalimumab are stable in whole blood clotted samples for seven days at room temperature. Ann Clin Biochem 2015;52:672–674.
- Johnson JL, Abecasis GR. GAS Power Calculator: webbased power calculator for genetic association studies. bioRxiv 2017:164343.
- McCarthy S, Das S, Kretzschmar W, et al. A reference panel of 64,976 haplotypes for genotype imputation. Nat Genet 2016;48:1279–1283.
- Lange KM de, Moutsianas L, Lee JC, et al. Genomewide association study implicates immune activation of

- multiple integrin genes in inflammatory bowel disease. Nat Genet 2017;49:256–261.
- Wellcome Trust Case Control Consortium. Genomewide association study of 14,000 cases of seven common diseases and 3,000 shared controls. Nature 2007; 447:661–678.
- Panaccione R, Loftus EV, Binion D, et al. Efficacy and safety of adalimumab in Canadian patients with moderate to severe Crohn's disease: results of the Adalimumab in Canadian SubjeCts with ModErate to Severe Crohn's DiseaSe (ACCESS) trial. Can J Gastroenterol 2011; 25:419–425.
- Sandborn WJ, Rutgeerts P, Enns R, et al. Adalimumab induction therapy for Crohn disease previously treated with infliximab. Ann Intern Med 2007;146:829.
- Megiorni F, Pizzuti A. HLA-DQA1 and HLA-DQB1 in Celiac disease predisposition: practical implications of the HLA molecular typing. J Biomed Sci 2012; 19:88.
- Oliveira-Cortez A, Melo AC, Chaves VE, et al. Do HLA class II genes protect against pulmonary tuberculosis? A systematic review and meta-analysis. Eur J Clin Microbiol Infect Dis 2016;35:1567–1580.
- Reddy MPL, Wang H, Liu S, et al. Association between type 1 diabetes and GWAS SNPs in the southeast US Caucasian population. Genes Immun 2011;12:208–212.
- 23. Eleftherohorinou H, Hoggart CJ, Wright VJ, et al. Pathway-driven gene stability selection of two rheumatoid arthritis GWAS identifies and validates new susceptibility genes in receptor mediated signalling pathways. Hum Mol Genet 2011;20:3494–3506.
- 24. Ursum J, Weijden MAC van der, Schaardenburg D van, et al. IL10 GGC haplotype is positively and HLA-DQA1\*05-DQB1\*02 is negatively associated with radiographic progression in undifferentiated arthritis. J Rheumatol 2010;37:1431–1438.
- De Groot AS, Knopp PM, Martin W. De-immunization of therapeutic proteins by T-cell epitope modification. Dev Biol (Basel) 2005;122:171–194.
- Sathish JG, Sethu S, Bielsky M-C, et al. Challenges and approaches for the development of safer immunomodulatory biologics. Nat Rev Drug Discov 2013;12:306–324.
- 27. **Goyette P, Boucher G**, Mallon D, et al. High-density mapping of the MHC identifies a shared role for HLA-DRB1\*01:03 in inflammatory bowel diseases and heterozygous advantage in ulcerative colitis. Nat Genet 2015;47:172–179.
- González-Galarza FF, Takeshita LYC, Santos EJM, et al. Allele frequency net 2015 update: new features for HLA epitopes, KIR and disease and HLA adverse drug reaction associations. Nucleic Acids Res 2015;43:D784– D788.

Author names in bold designate shared co-first authorship.

Received August 30, 2019. Accepted September 29, 2019.

### Reprint requests

Address requests for reprints to: Carl A. Anderson, PhD, Genomics of Inflammation and Immunity Research Group, Morgan Building, Wellcome Sanger Institute, Wellcome Genome Campus, Hinxton CB10 1SA, UK.

e-mail: carl.anderson@sanger.ac.uk; or Tariq Ahmad, DPhil, Exeter IBD and Pharmacogenetics Research Group, RILD Building, Barrack Road, Exeter EX2 5DW, UK. e-mail: tariq.ahmad1@nhs.net.

### Acknowledgments

The authors would like to acknowledge the study coordinators of the Exeter IBD Research Group, United Kingdom, Marian Parkinson and Helen Gardner-Thorpe, for their ongoing administrative support to the study. The authors would also like to thank the National Institute for Health Research UK IBD BioResourse for help establishing the replication cohort.

Transcript profiling: Summary statistics will be deposited on the NHGRI-EBI GWAS Catalog upon publication. Genotype data that supports this study has been deposited in the European Genome-phenome Archive under the accession code EGAS00001000924.

Author contributions: T.A., C.A.A., J.C.B., G.A.H., C.M.B., T.J.M., M.H.P., P.M.I., and N.A.K. participated in the conception and design of the work: C.M.B was the project manager and coordinated recruitment. A.S., L.M., D.L.R., J.C.B., C.A.A, M.R, G.J.W, and N.A.K. were involved in the acquisition, analysis or interpretation of data. C.W.L., J.R.F.C., M.P., J.C.M., D.L.R., P.M.I., and N.A.K. were involved in the acquisition of the replication cohort. The statistical analysis was performed by A.S., N.A.K., L.M., J.C.B., and C.A.A. Drafting of the manuscript was conducted by A.S., N.A.K., L.M., G.A.H., C.M.B., N.C., D.M., J.R.G., C.A.A, and T.A. All the authors contributed to the critical review and final approval of the manuscript. T.A. and C.A.A. obtained the funding for the study. All remaining authors contributed by submitting a substantial number of samples in line with International Committee of Medical Journal Editors criteria. Carl A. Anderson and Tariq Ahmad contributed equally to this work.

#### Conflicts of interest

These authors disclose the following: Graham A. Heap reports non-financial support from AbbVie, outside the submitted work; and he is now an employee of AbbVie and owns stock in the company. Mark Reppell is an employee of AbbVie and owns stock in the company. Gareth J. Walker has received honoraria from AbbVie. Charlie W. Lees reports grants and personal

fees from AbbVie, Amgen, Pfizer, and Samsung Bioepis. J.R. Fraser Cummings reports personal fees from AbbVie, grants and personal fees from Biogen, personal fees from MSD, Celltrion, NAPP, and Sandoz. Peter M. Irving reports personal fees from AbbVie, Pfizer, Sandoz, Samsung and Bioepis, grants from MSD; outside the submitted work. Dermot McGovern reports grants and personal fees from Pfizer outside the submitted work. James R. Goodhand received honoraria from AbbVie, for unrelated topics. Tariq Ahmad has received unrestricted research grants, advisory board fees, speaker honorariums, and support to attend international meetings from AbbVie, Merck, Pfizer, NAPP, Celltrion, and Hospira. The remaining authors disclose no conflicts.

### **Funding**

PANTS is an investigator-led study funded by CORE (renamed Guts UK in 2018), the research charity of the British Society of Gastroenterology, and by unrestricted educational grants from AbbVie Inc, USA, Merck Sharp & Dohme Ltd, UK, NAPP Pharmaceuticals Ltd, UK, Pfizer Ltd, USA, Celltrion Healthcare, South Korea, and Cure Crohn's Colitis (Scottish IBD Charity). HLA long-read sequencing was supported through a sponsored research agreement with AbbVie Inc. The drug and anti-drug antibody assays were provided at reduced cost by Immundiagnostik AG. Laboratory tests were undertaken by the Exeter Blood Sciences Laboratory at the Royal Devon & Exeter National Health Service Trust (https://www.exeterlaboratory.com). The sponsor of the PANTS study is the Royal Devon and Exeter National Health Service Foundation Trust. Alekseis Sazonovs, Loukas Moutsianas, Daniel L. Rice, Jeffrey C. Barrett, and Carl A. Anderson are funded by the Wellcome Trust, who also funded the genotyping of all samples (206194). Timothy J. McDonald is funded by a National Institute for Health Research Health Education England Senior Lectureship. The views expressed in this publication are those of the author(s) and not necessarily those of the National Health Service, Health Education England, National Institute for Health Research, or the Department of Health. None of the listed funders had a role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, and decision to submit the manuscript for publication.