

Receptor tyrosine and serine-threonine kinases

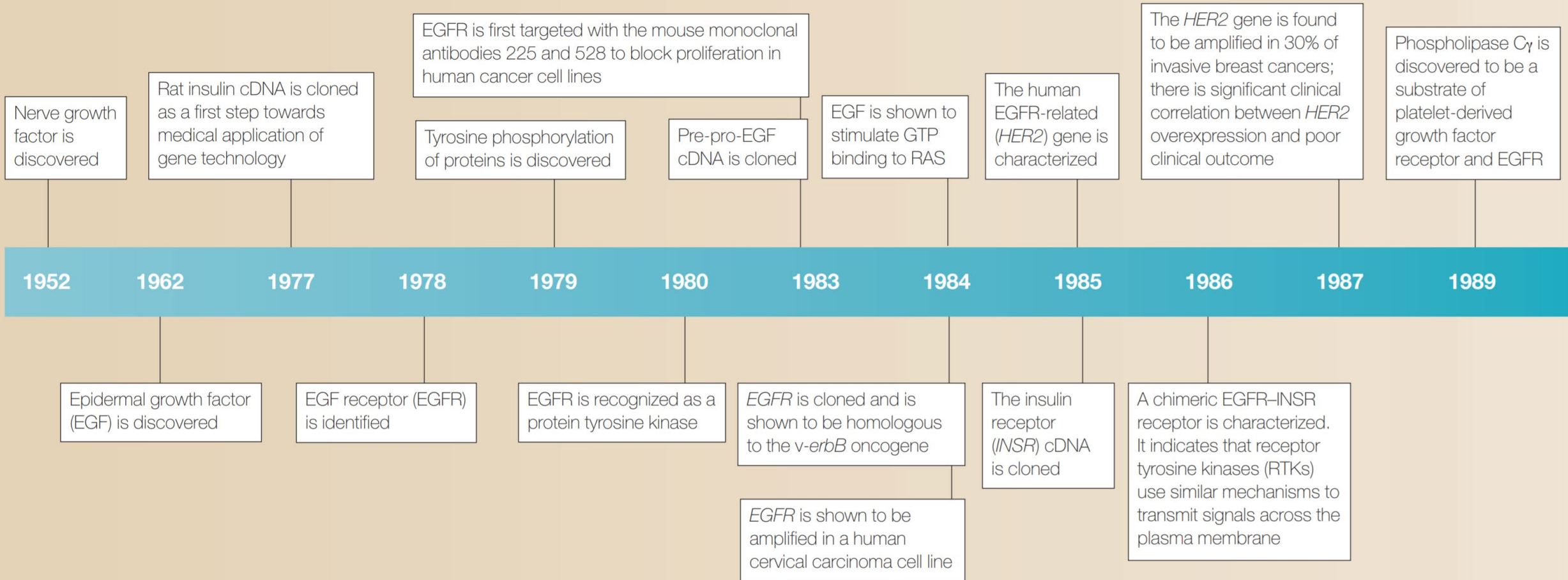
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Receptor Tyrosine Kinases

Timeline 1 | Breakthrough discoveries on RTK signal transduction and RTK-based cancer therapy



Levi-Montalcini: NGF

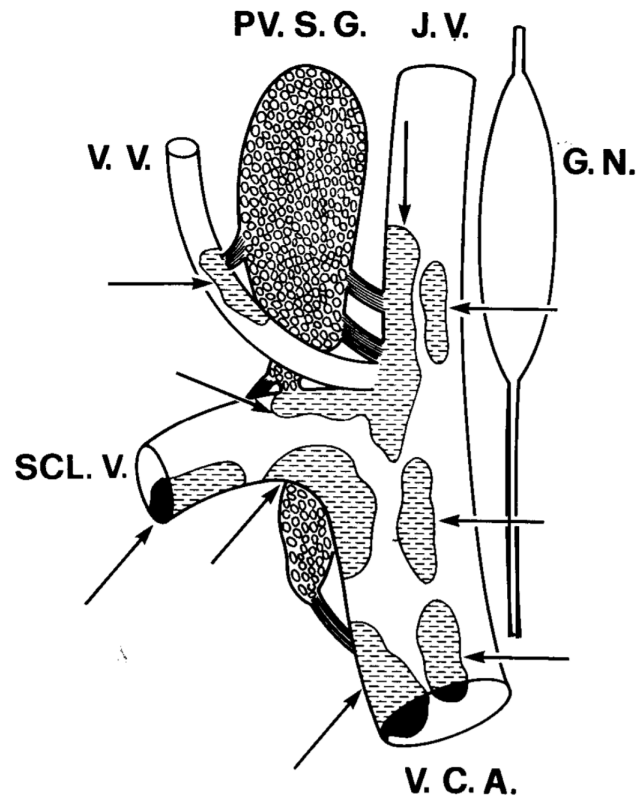
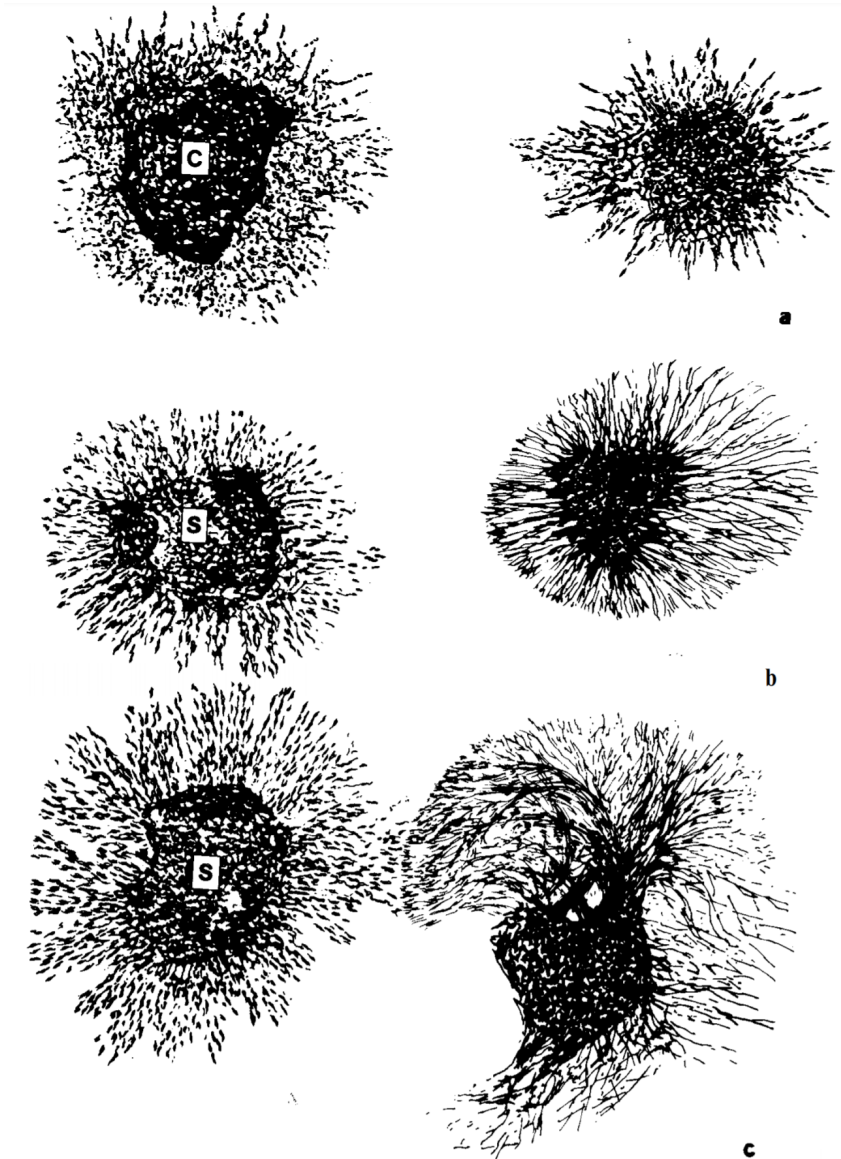


Fig 1. Sixteen-day chick embryo with intra-embryonic tumor (sarcoma 180). Ingrowth of sympathetic nerve fibers into the Jugular, Vertebral, Subclavian Anterior Caval Veins. GN, Ganglion Nodosum; JV, Jugular Vein; Pv.SG, Paravertebral Sympathetic Ganglion; SCL.V, Subclavian Vein; VCA, Anterior Caval Vein; VV, Vertebral Vein. Arrows point to nerve agglomerations. (from Ref. 12)



Cohen: EGF

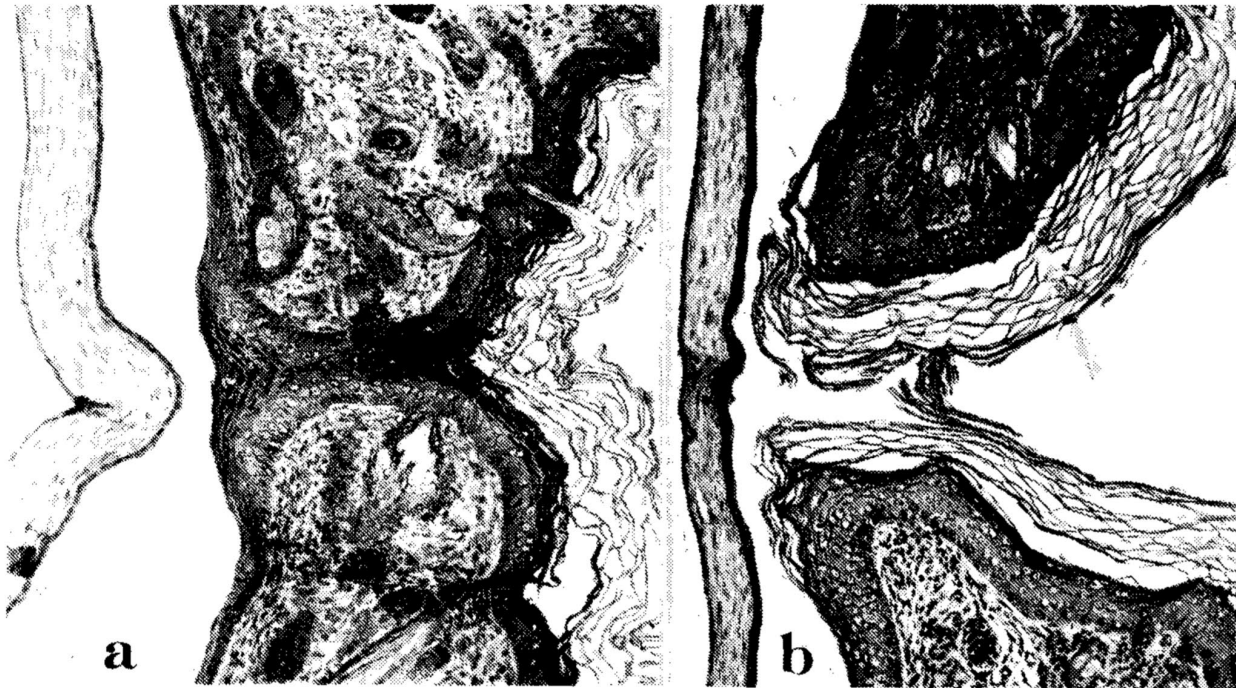


Figure 1a and b. Cross sections of the eyelid area from control, 1a, and experimental, 1b, 8-day-old rats. The experimental animal had received daily injections (1 μg per 1 gm body weight) of the active protein. $\times 100$. (reprinted from *Journal of Investigative Dermatology* (1963) 40, 1-5.)

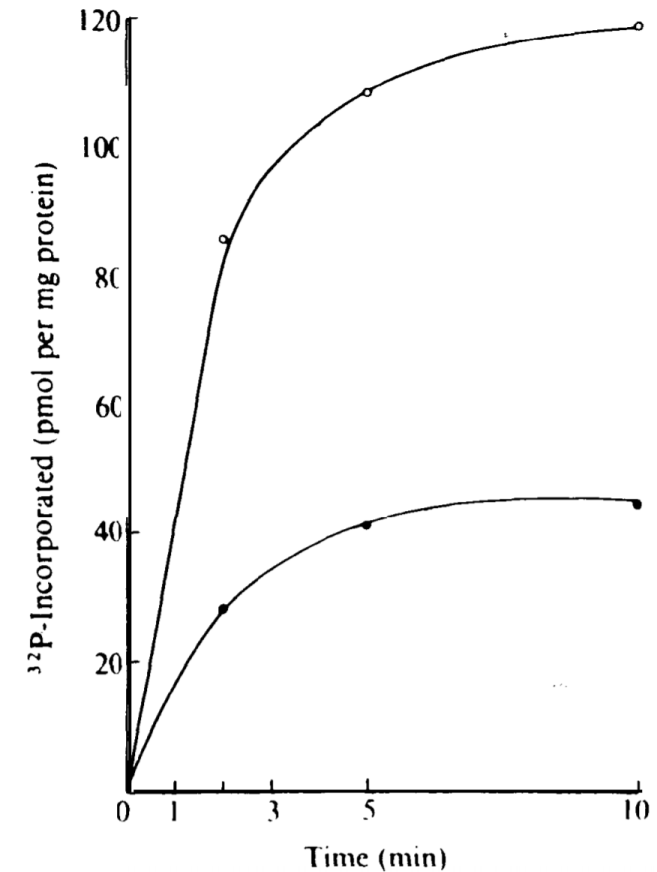
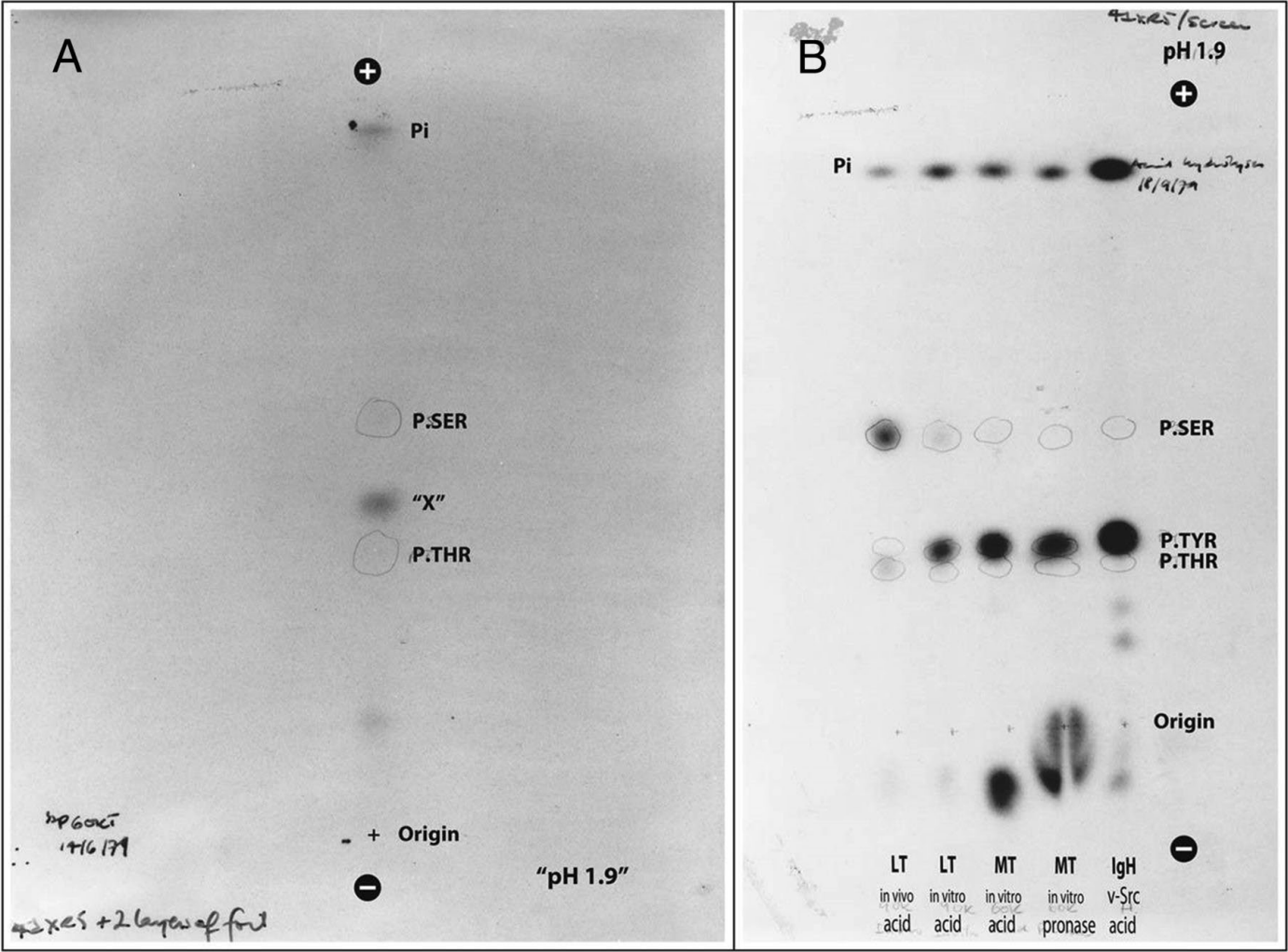


Figure 4. Stimulation of EGF of the incorporation of ^{32}P -hosphate from $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ into cell membranes. The reaction mixtures contained A-431 membrane (27 μg protein), HEPES buffer (20 mM, pH 7.4), MnCl_2 (1 mM), $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ (15 μM , $8 \times 10^5 \text{ c.p.m.}$), EGF (40 ng) and BSA (7.5 μg) in a final volume of 60 μl . The reaction tubes were placed on ice and preincubated for 10 min in the presence (○) or absence (●) of EGF. The reaction was initiated by the addition of labelled ATP and incubation at 0°C was continued for the indicated times. The reaction was terminated by pipetting 50- μl aliquots on to squares (2 cm) of Whatman 3 MM filter paper and immediately dropping the paper into a beaker of cold 10% TCA containing 0.01 M sodium pyrophosphate. The filter papers were washed extensively with pyrophosphate-containing 10% TCA at room temperature, extracted with alcohol and ether dried, and the radioactivity measured in a Nuclear Chicago gas-flow counter. (reprinted from *Nature* (1978) 276, 408-410.)

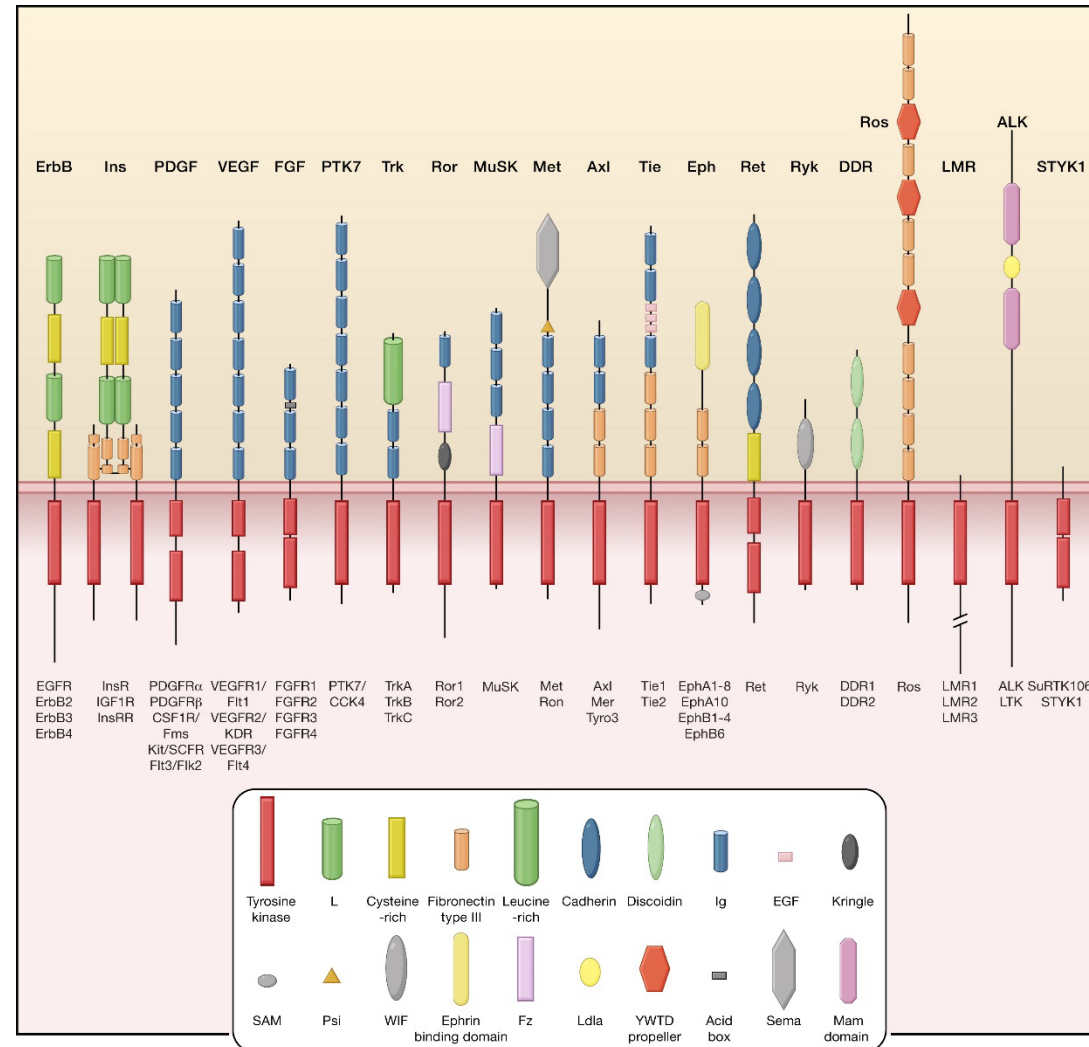
First experiments demonstrating polyoma middle T and v-Src tyrosine kinase activity



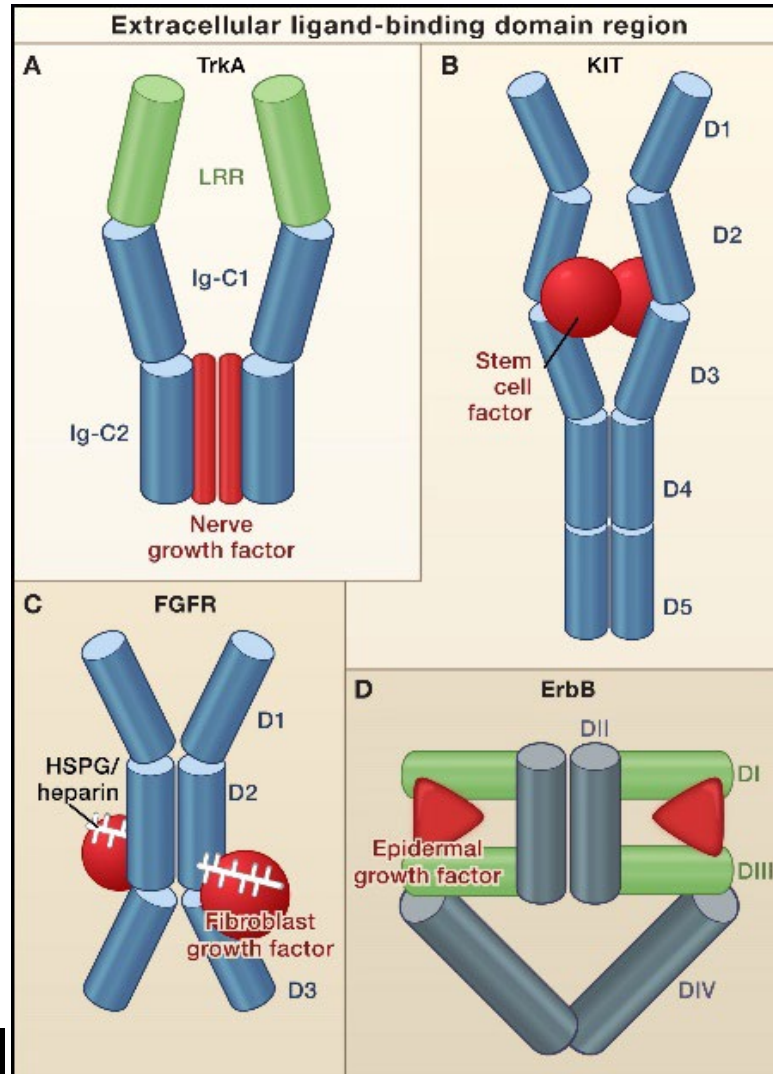
Tony Hunter PNAS 2015;112:26:7877-7882

PNAS

Humans have 58 known RTKs (20 subfamilies)



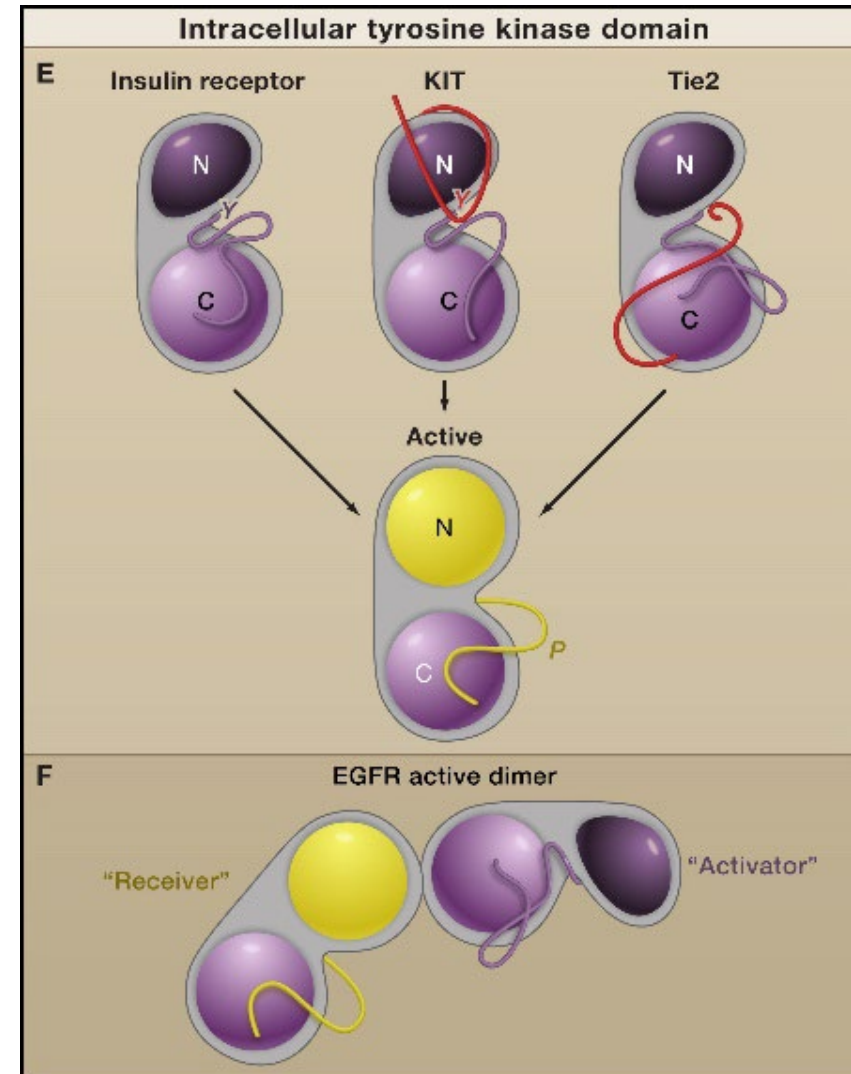
RTK Dimerization and Kinase Activation

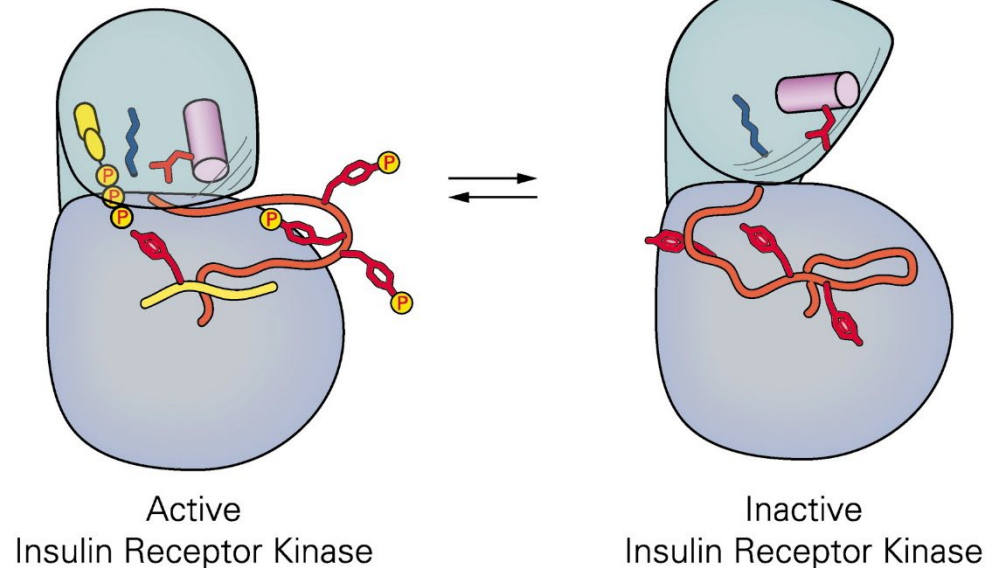
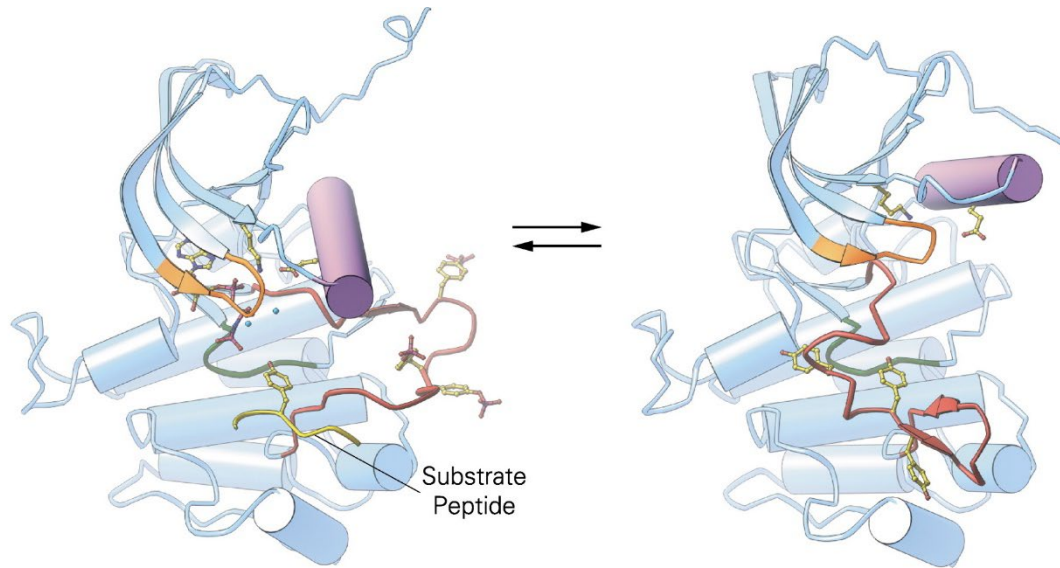


- Receptor activation requires dimerization/oligomerization
- Ligand-mediated dimerization
 - TrkA
- Ligand- and receptor-mediated dimerization
 - KIT
 - FGFR
- Receptor-mediated dimerization
 - ErbB

RTK Dimerization and Kinase Activation

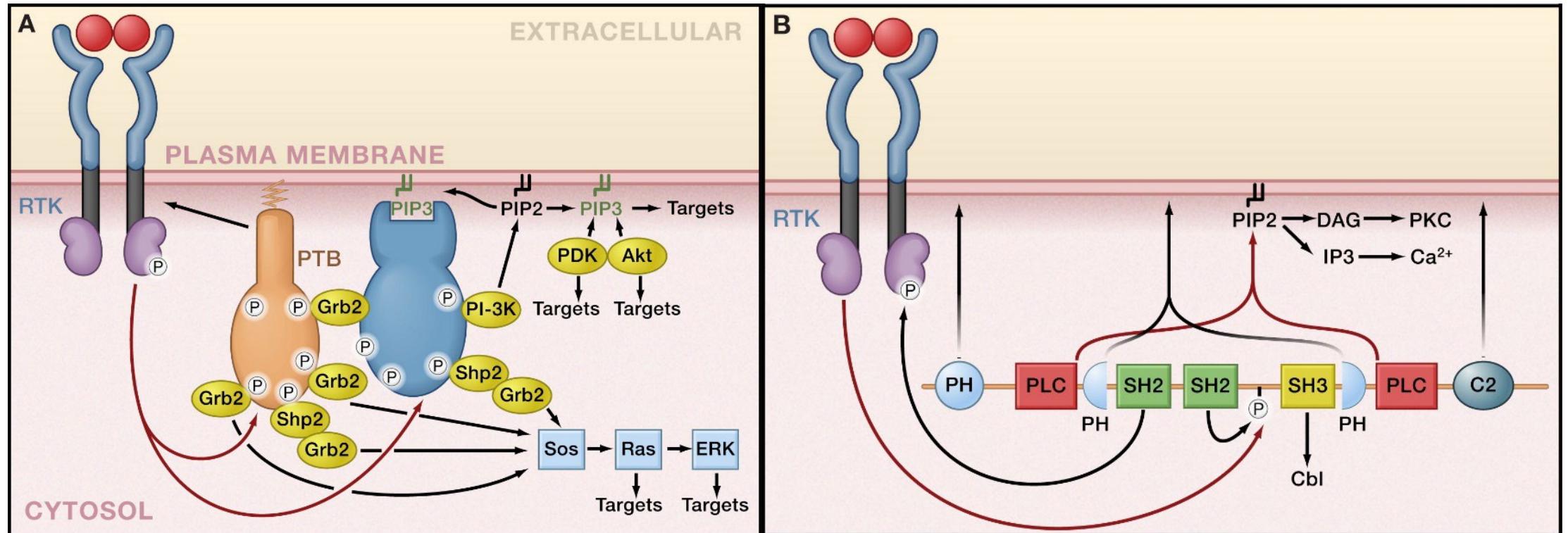
- Tyrosine kinase domain architecture: Each TKD is *cis*-autoinhibited and release of autoinhibition required for activity
 - N lobe
 - C lobe
 - Activation loop or other mechanism
- Mechanism for activation of kinase can differ





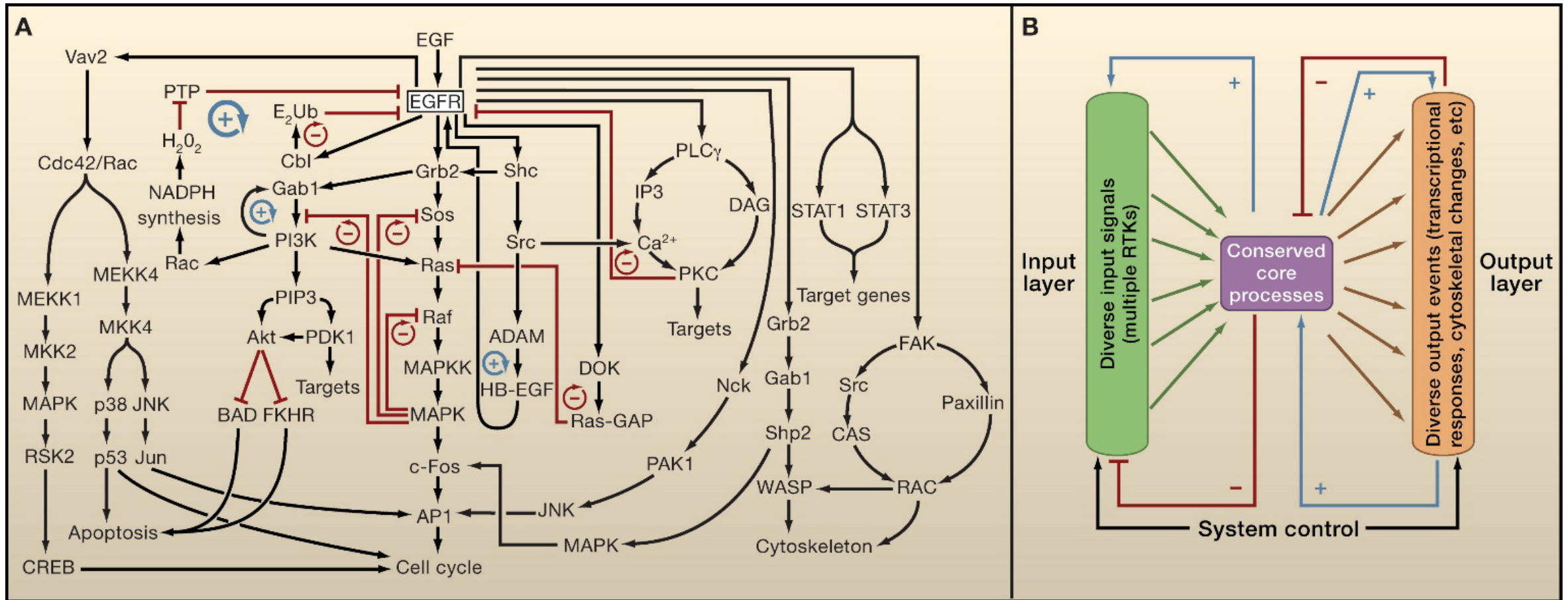
- N-lobe is composed of five-stranded β -sheet and one helix αC
- C-lobe predominantly helical
- ATP bound between the lobes with a GXGX ϕ G phosphate binding loop
- Activation loop provides platform for binding peptide substrate
- Catalytic lysine and glutamate are present in the N-lobe

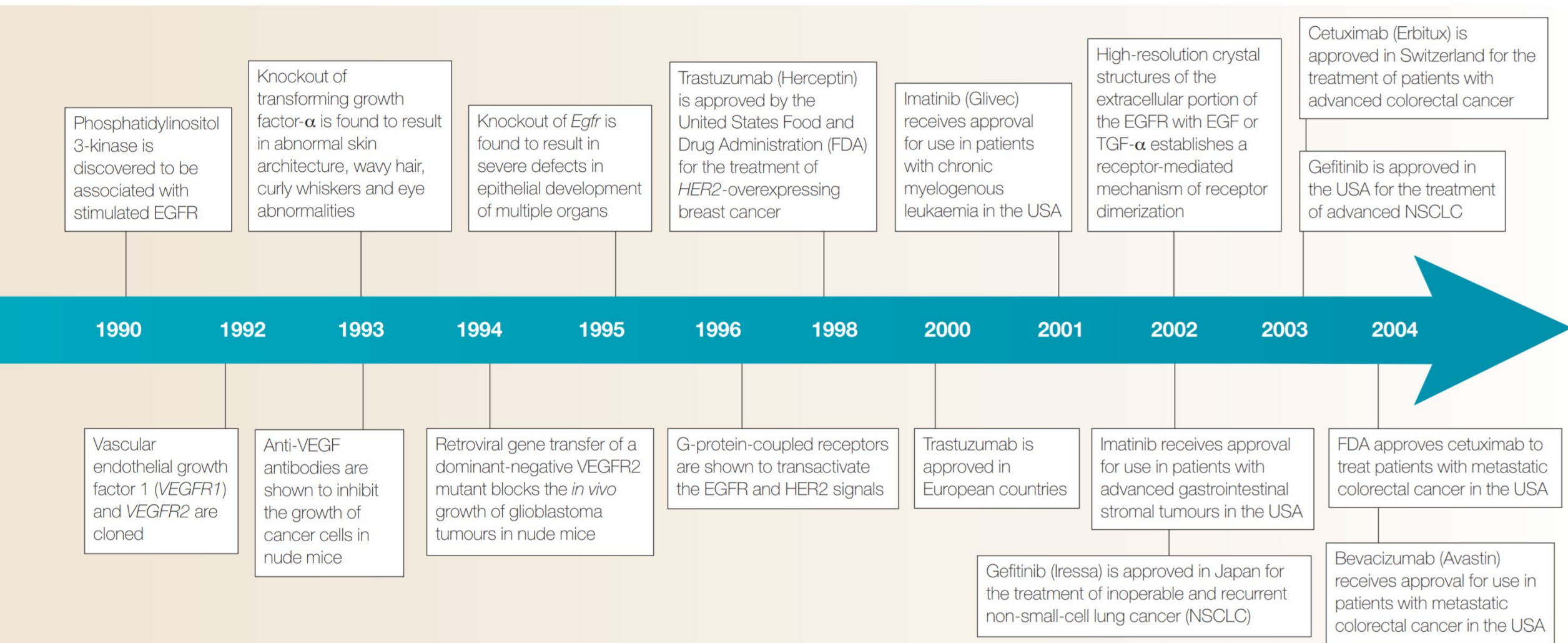
RTK Signaling through adapter proteins



EGF receptor signaling

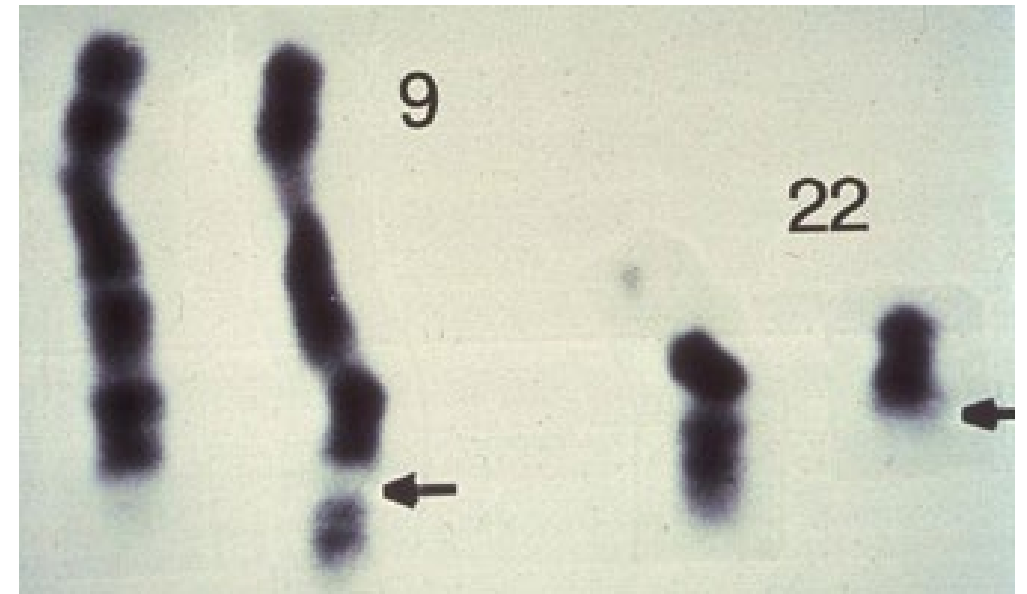
“Bow-tie” network for RTK integration





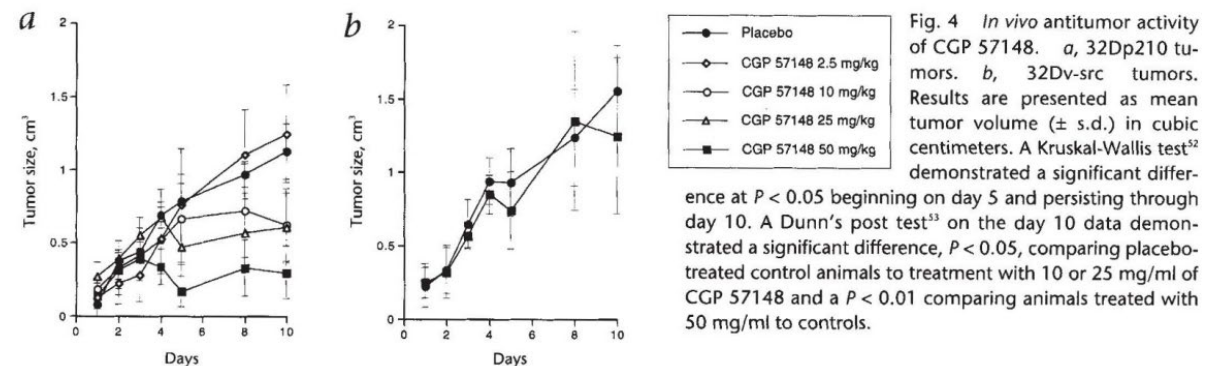
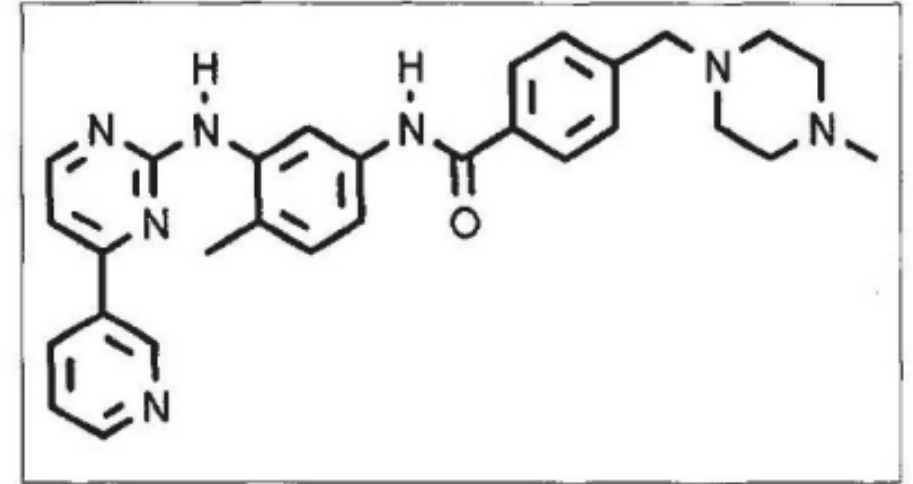
Imatinib (ok – it actually targets a non-receptor tyrosine kinase)

- Chronic myelogenous leukemia – Nowell and Hungerford show an abnormal chromosome (Philadelphia chromosome)
- Rowley shows it is due to translocation
- Later shown to result in a fusion protein BCR-ABL
- BCR-ABL protein promotes abnormal WBC proliferation



Imatinib (ok – it actually targets a non-receptor tyrosine kinase)

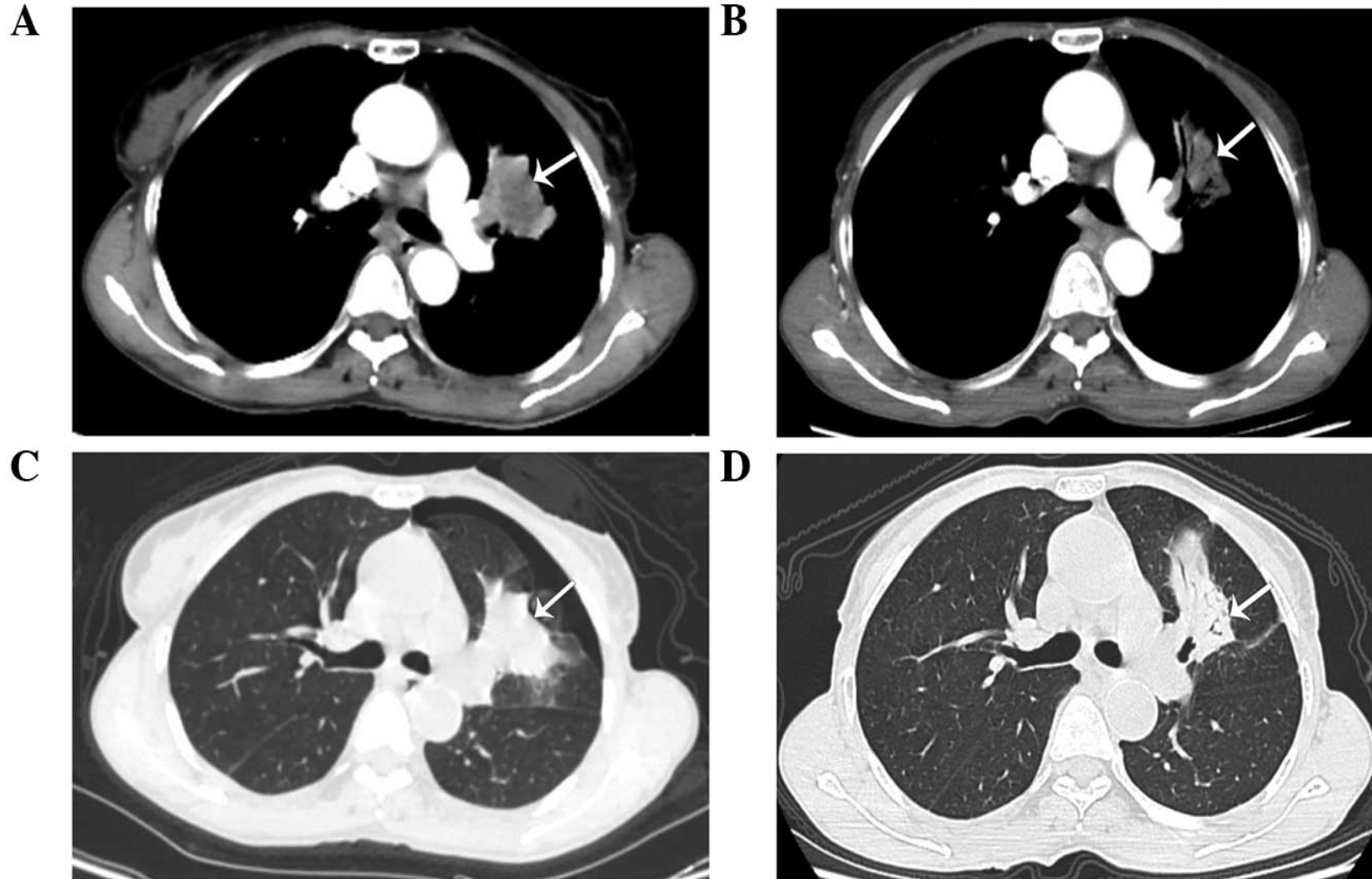
- Brian Druker collaborated with Ciba-Geigy (now Novartis) to screen their kinase inhibitors
- Identified a compound which inhibited BCR-ABL and tumor growth *in vitro*
- > 90% response *in vivo*, ushering in the era of targeted therapies



Cancer Therapies

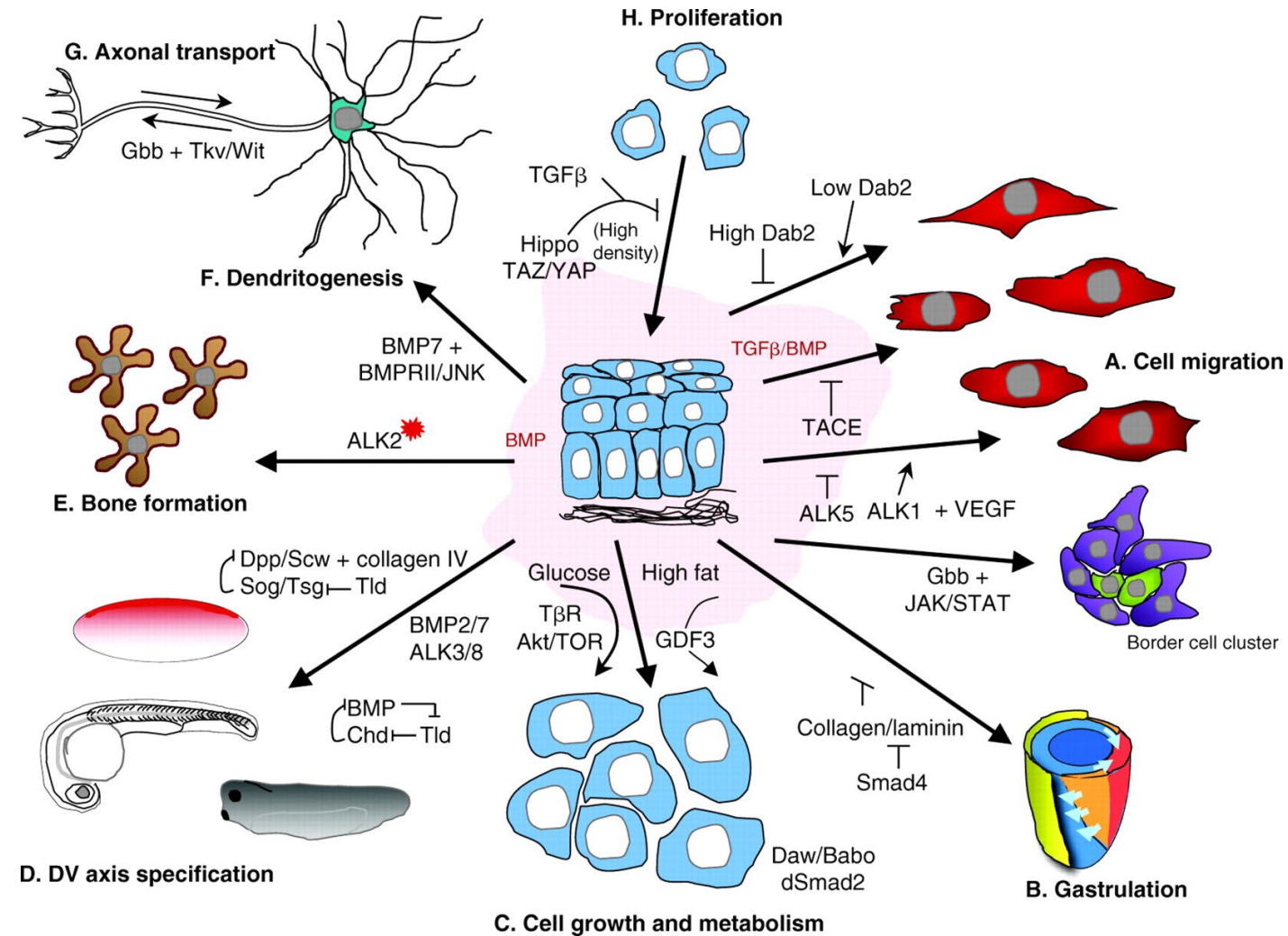
Target	Compound	Cancer	References
EGFR family			
HER2	Trastuzumab (Herceptin)	HER2-positive breast cancer	[7–9]
EGFR	Cetuximab (Erbix)	Metastatic colorectal cancer	[10]
	Panitumumab (Vectibix)	(RAS wild type)	[10–12]
	Gefitinib (Iressa)	Metastatic non-small-cell lung cancer	[13–15]
	Erlotinib (Tarceva)		[13,16]
EGFR and HER2	Lapatinib (Tykerb)	HER2-positive breast cancer (Trastuzumab-resistant)	[14,17]
	Afatinib	NSCLC HER2-positive breast cancer	[18–21]
VEGFR	Sorafenib (Nexavar)	Renal, liver and thyroid cancer	[22–24]
	Sunitinib (Sutent)	Renal cell cancer Gastrointestinal stromal tumor (GIST)	[25,26]
	Bevacizumab (Avastin)	Metastatic colorectal carcinoma	[27]
PDGFR	Imatinib (Gleevec)	GIST (KIT+)	[28]
PDGFR and VEGFR	Sunitinib	Angiogenesis	[29–32]
	Soratinib		
	Pazopanib		
	Nilotinib		
FGFR and VEGFR	Brivanib (BMS-540215)	Human hepatocellular carcinoma model	[33]
VEGFR, PDGFR, FLT-3, c-KIT and FGFR	CHIR-258 (TKI-258)	Multiple myelomas	[34,35]
MET	SGX523	MDCK and A549 cells and GTL16 xenograft models	[36]
C-KIT	Imatinib (Gleevec)	GIST	[37–39]

EGFR mutation+ response to TKIs



Receptor Serine/Threonine Kinases

A summary of biological responses to TGF β family signaling in development



Kristi Wharton, and Rik Derynck Development
2009;136:3691-3697

Isolation of Transforming Growth Factors

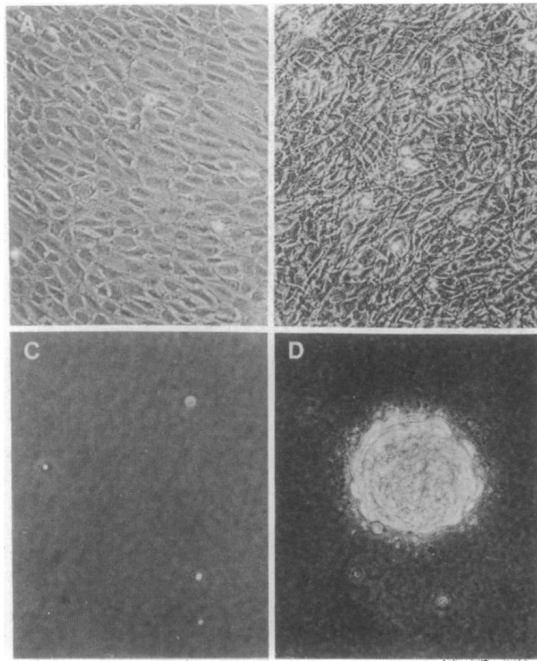


FIG. 2. Effect of SGFs on morphology and growth in soft agar. (A) Rat clone 49F seeded 6 days previously at 1×10^5 cells per 30-cm² tissue culture flask in Dulbecco's modified Eagle's medium containing 10% calf serum (the medium was changed once, 3 days after seeding). (B) Parallel culture treated with SGF (2.5 µg of protein per ml, fraction 43 from Fig. 1); the medium was changed 3 days after seeding and new SGF (fraction 43) was added. (C) Clone 49F cells at 2 weeks after seeding in soft agar without the addition of SGF. The cells were seeded at 1×10^4 cells per 20-cm² petri dish and were fed with a 2-ml overlay of 0.3% agar after 7 days. More than 10^7 rat fibroblast cells (clone 49F) have been plated in soft agar and, in the absence of SGFs, no spontaneous colony formation has been seen with this clone. (D) Cells in C except that 0.4 µg of protein from fraction 67 of Fig. 1 was incorporated per ml of the soft agar. After 1 week these cells were refed with 2 ml of 0.3% agar containing 0.4 µg of protein from the same fraction per ml. (A, $\times 45$; B, $\times 45$; C, $\times 90$; D, $\times 90$.)

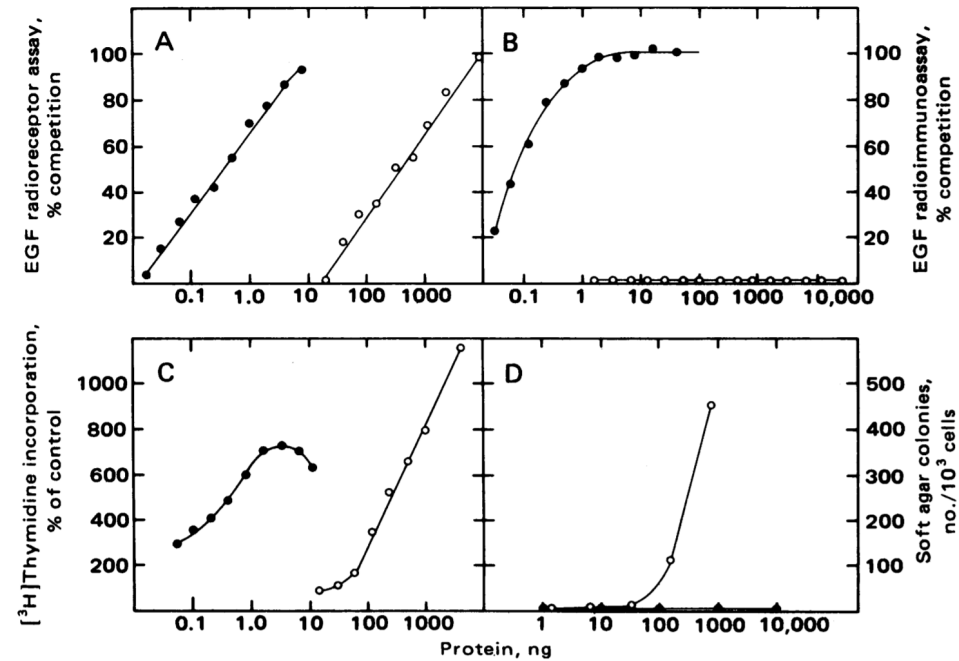


FIG. 3. Properties of EGF (●) and the 12,000 molecular weight peak of SGF (○) from Fig. 1. (A) EGF radioreceptor competition assays were performed with 2.5×10^4 mink lung cells and a sequential binding assay in which the competitor is allowed to preincubate with the cells at 22° for 1 hr. It is then removed, the cells are washed four times with binding buffer, and the ¹²⁵I-labeled EGF is incubated with the cells for 1 hr at 22°. (B) Radioimmunoassay using anti-EGF antibody and ¹²⁵I-labeled EGF. The antibody to mouse salivary gland EGF was produced in rabbits and the precipitating antibody was goat anti-rabbit IgG. (C) Thymidine incorporation studies using serum-depleted fibroblastic clone 49F. (D) Soft-agar growth-promoting activity. Rat clone 49F was seeded in the presence of varying concentrations of the peptides. The cells were fed 1 week after seeding and the colonies were counted at the end of 2 weeks.

Cloning of T β Rs

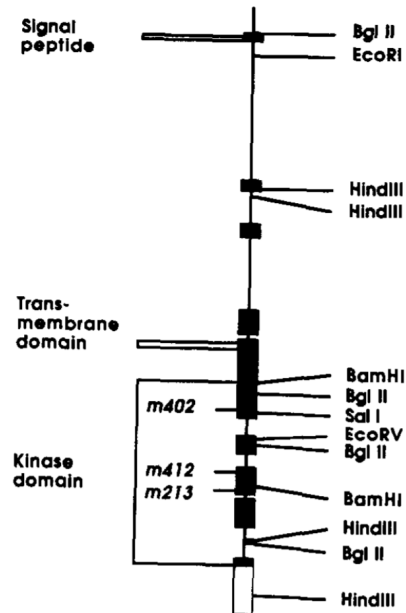


Figure 4. Physical Map of *daf-1*

A map based on restriction digests and sequence analysis of cDNA and genomic clones (see Figure 6) showing locations of 9 exons (solid boxes) and 8 introns. The open box represents the 3' untranslated region of the transcript. The sites of Tc1 insertion in the mutant alleles were determined from restriction digests (Figure 3a) and by sequence analysis of cloned mutant DNA. Also shown is the location of a C to T transition resulting in a novel BglIII site in the EMS-induced mutant allele *daf-1(m213)*. Evidence supporting the designation of protein functional domains (signal sequence, transmembrane and kinase domains) is provided in Figures 6–8.

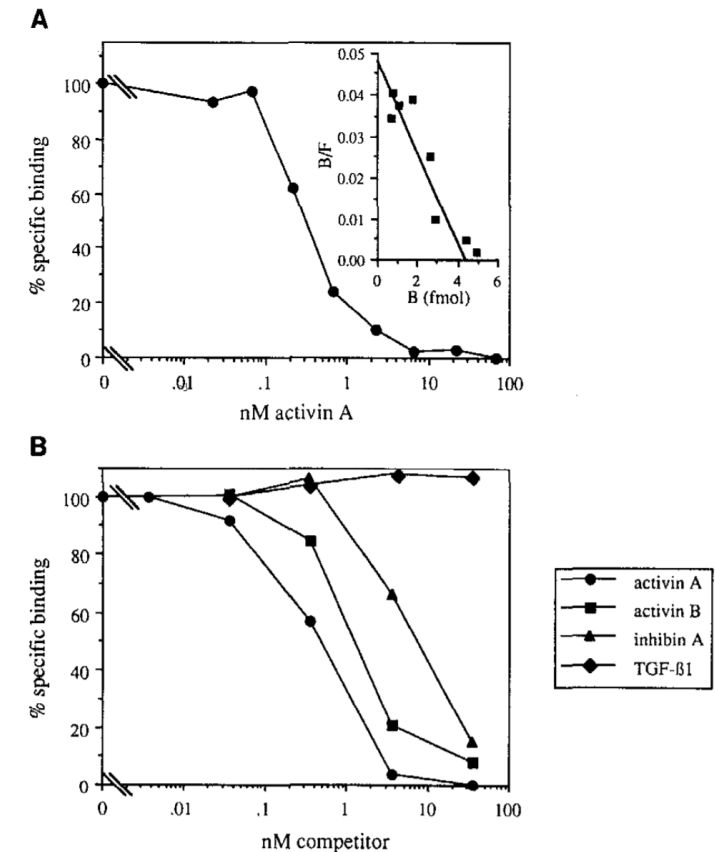
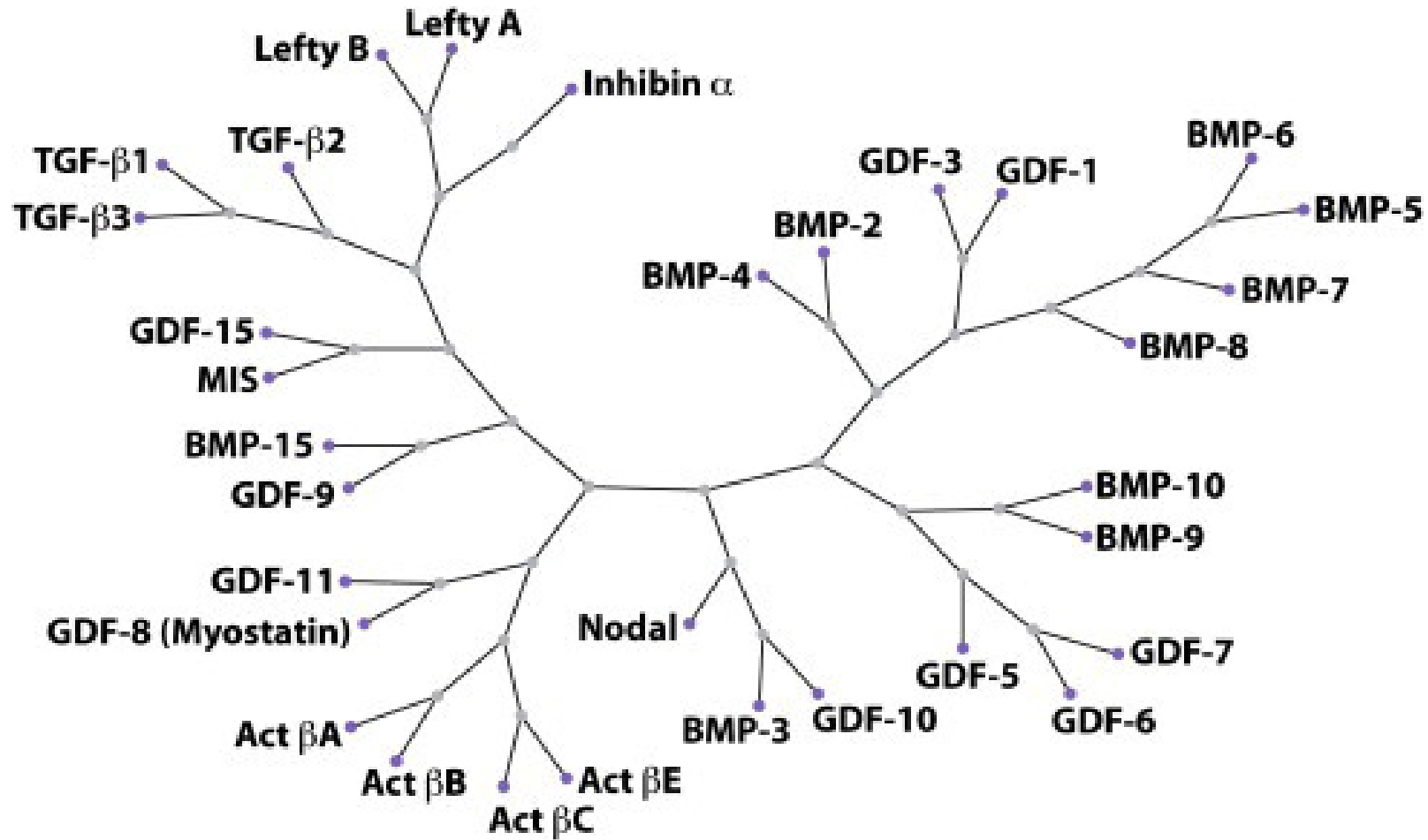


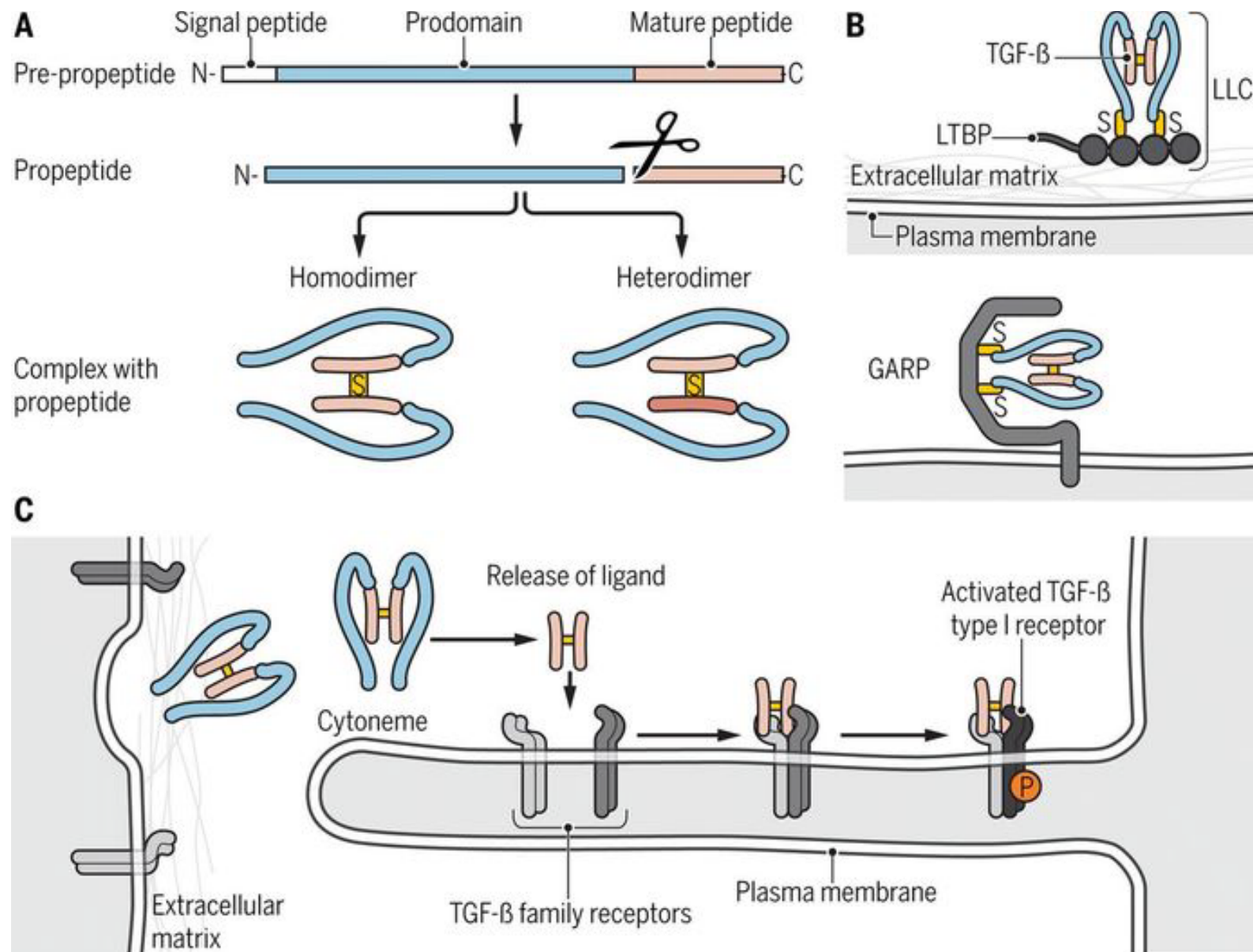
Figure 4. Competition Binding of ¹²⁵I-Activin A to COS Cells Transiently Transfected with pmActR1 or pmActR2

(A) ¹²⁵I-activin A binding to COS cells transfected with pmActR1. Binding was performed on cell monolayers as described in Experimental Procedures, and was competed with unlabeled activin A. Data are shown as percent specific binding, where 100% specific binding was 3.7% of input cpm and nonspecific binding was 0.9% of input cpm. Inset: Scatchard analysis of the data.

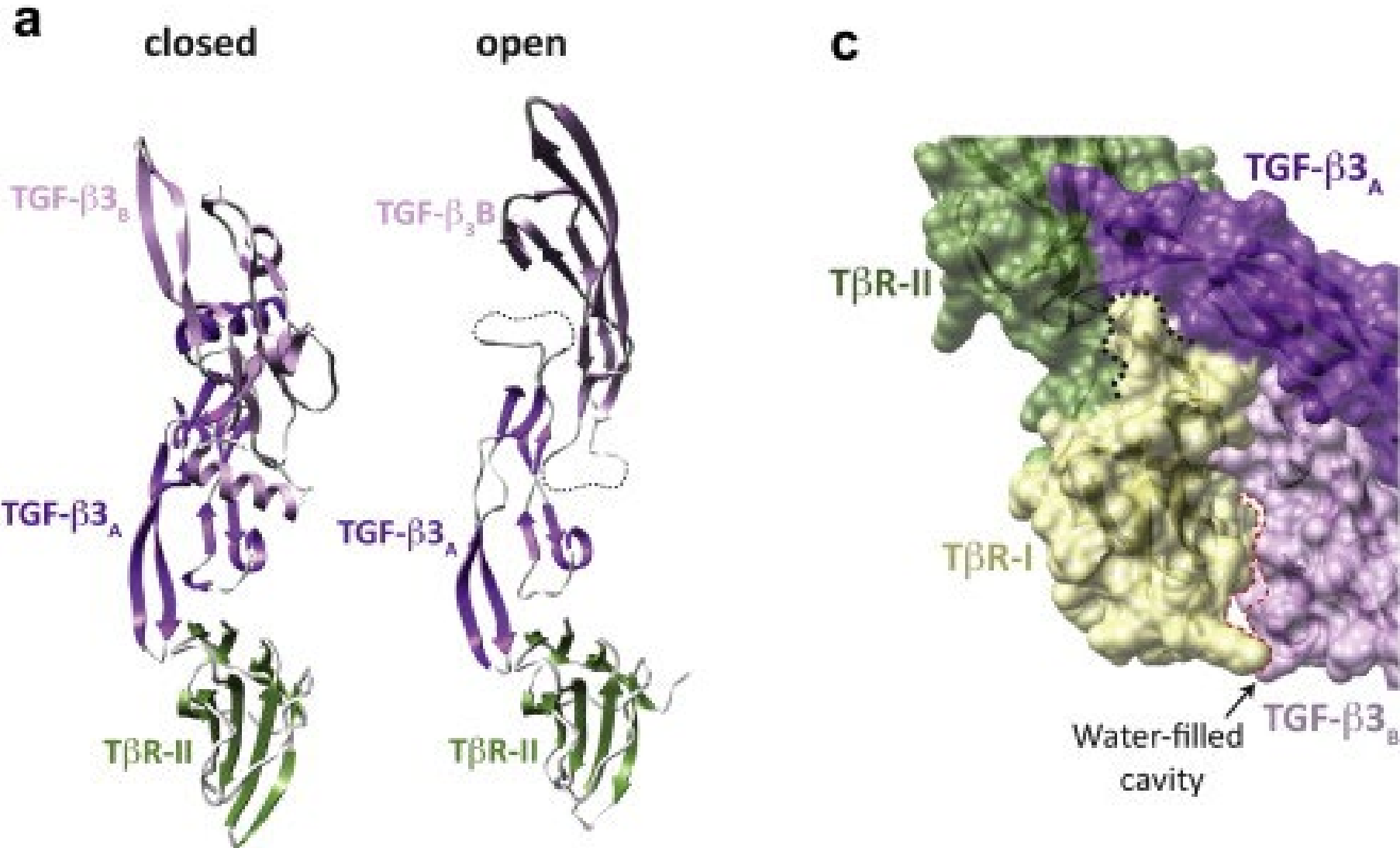
(B) ¹²⁵I-activin A binding to COS cells transfected with pmActR2. Binding was competed with unlabeled factors as indicated; 100% specific binding and nonspecific binding were 2.5% and 0.9%, respectively, of input cpm.

TGF- β superfamily

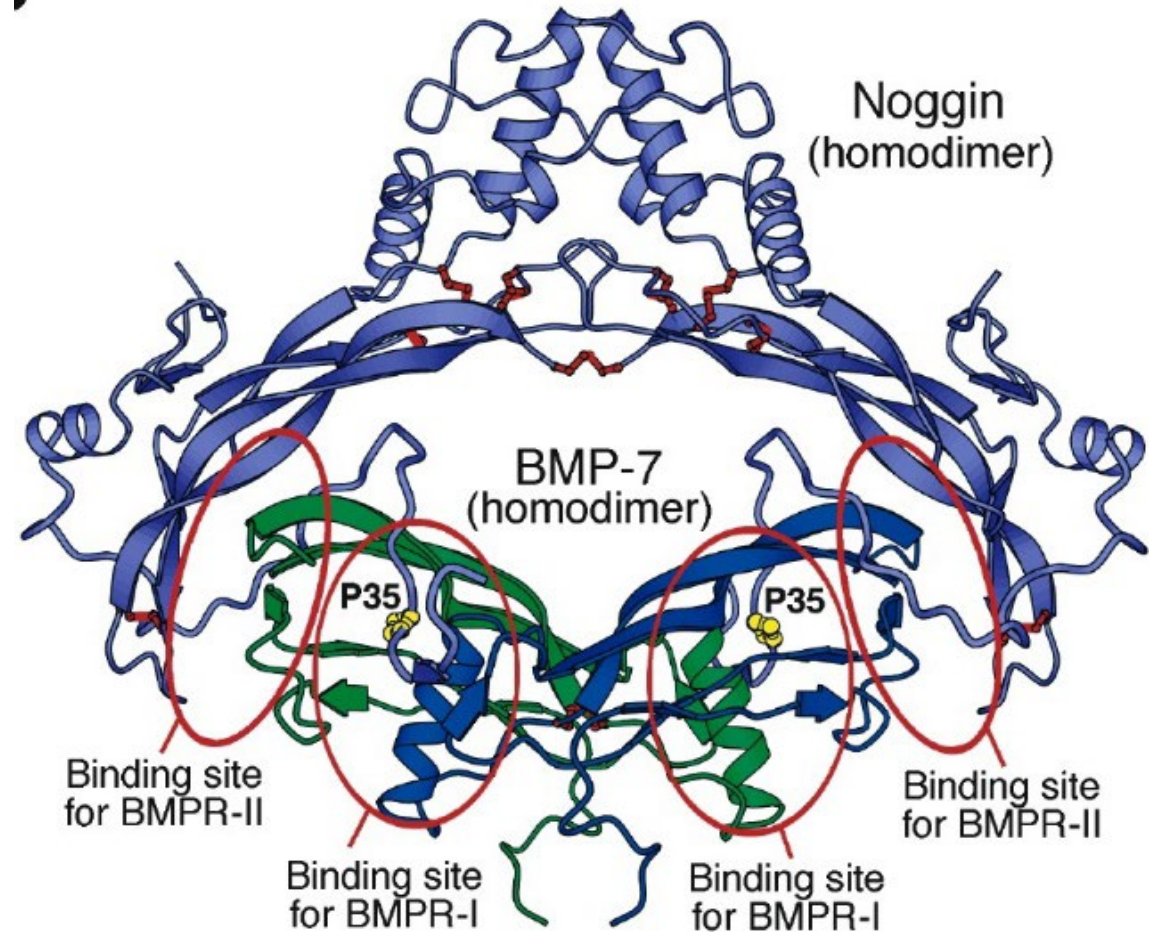
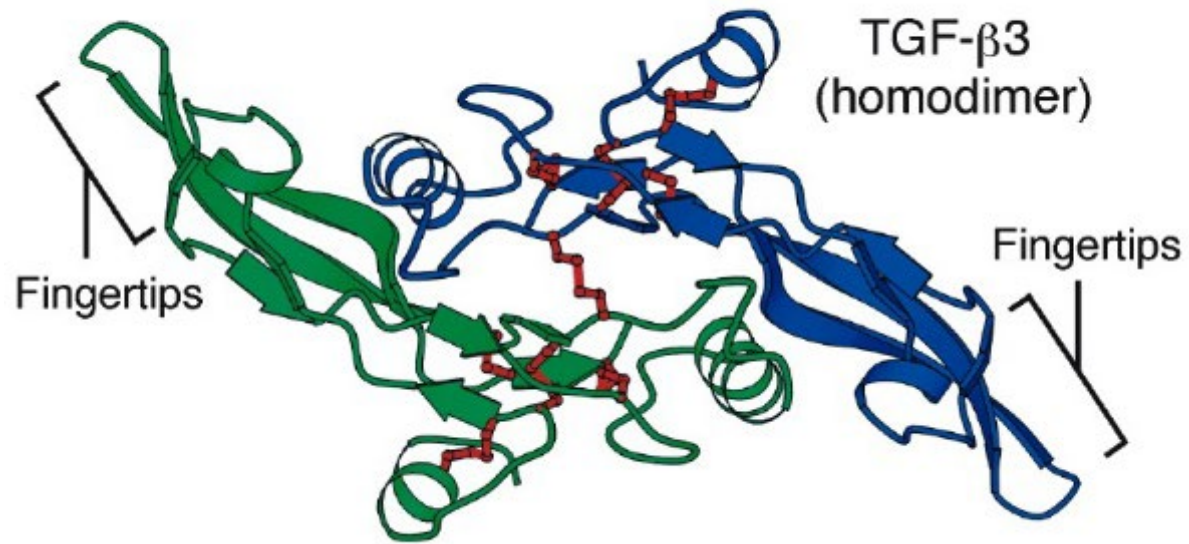




Alternative modes of binding

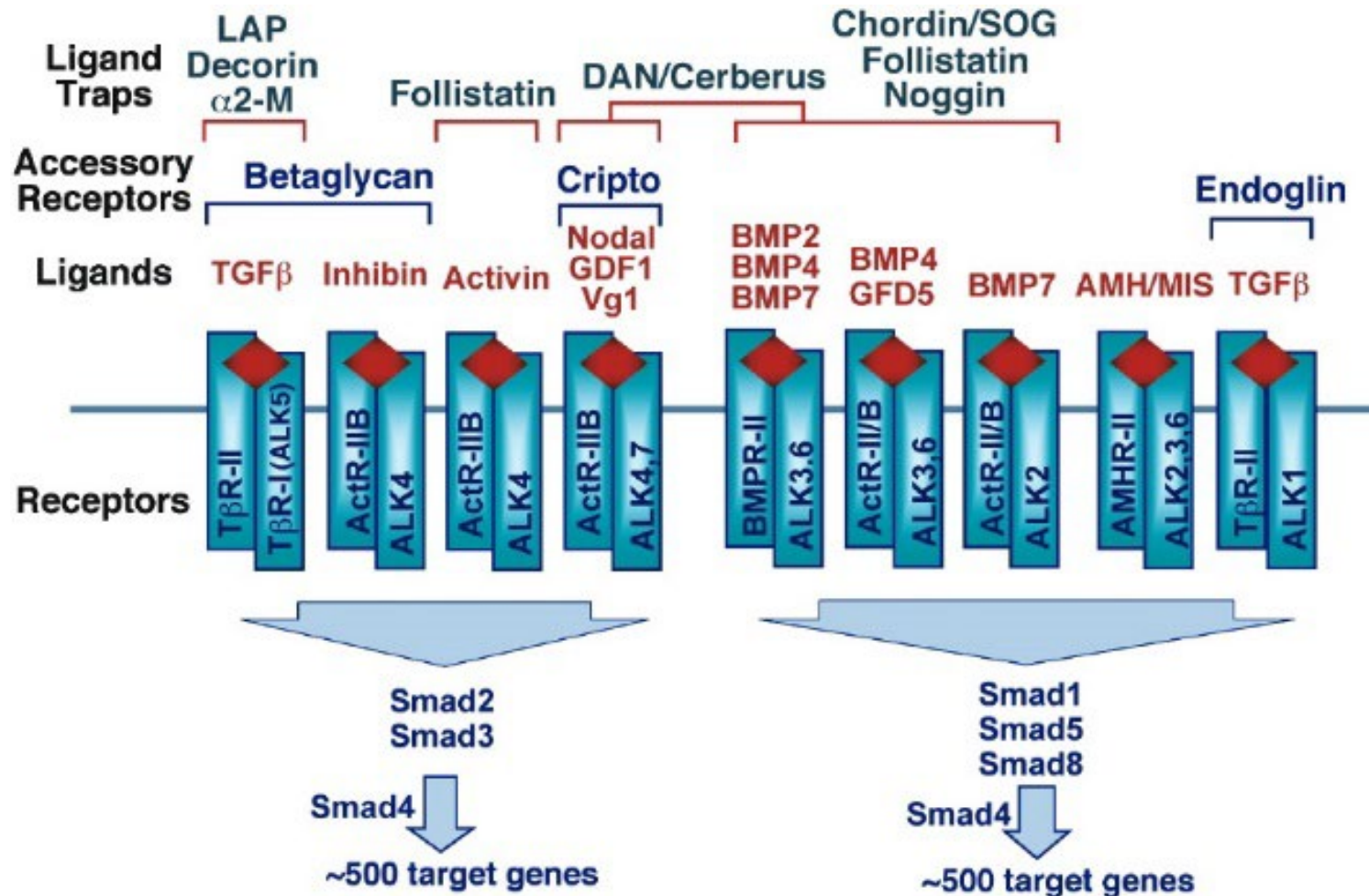


TGF- β dimerization and ligand traps

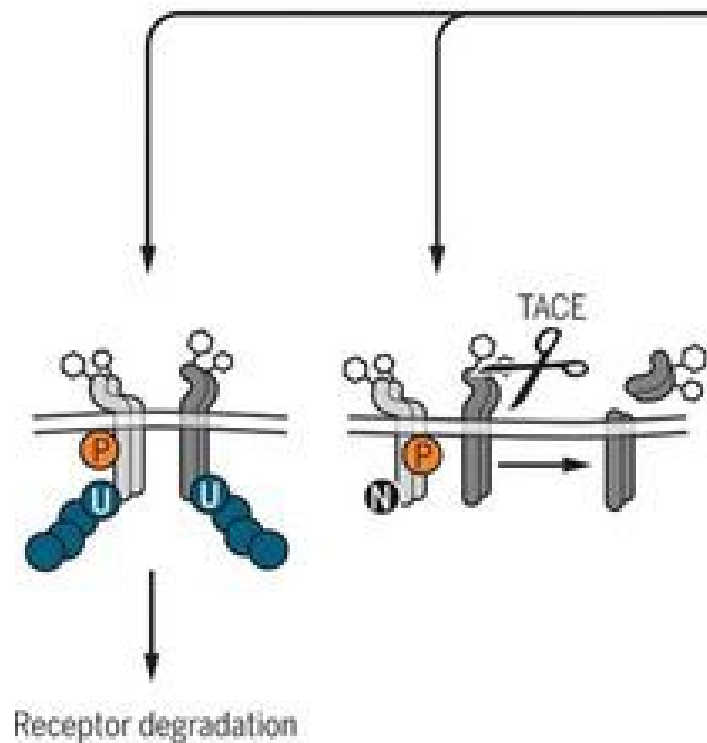


TGF- β superfamily, receptors and effectors

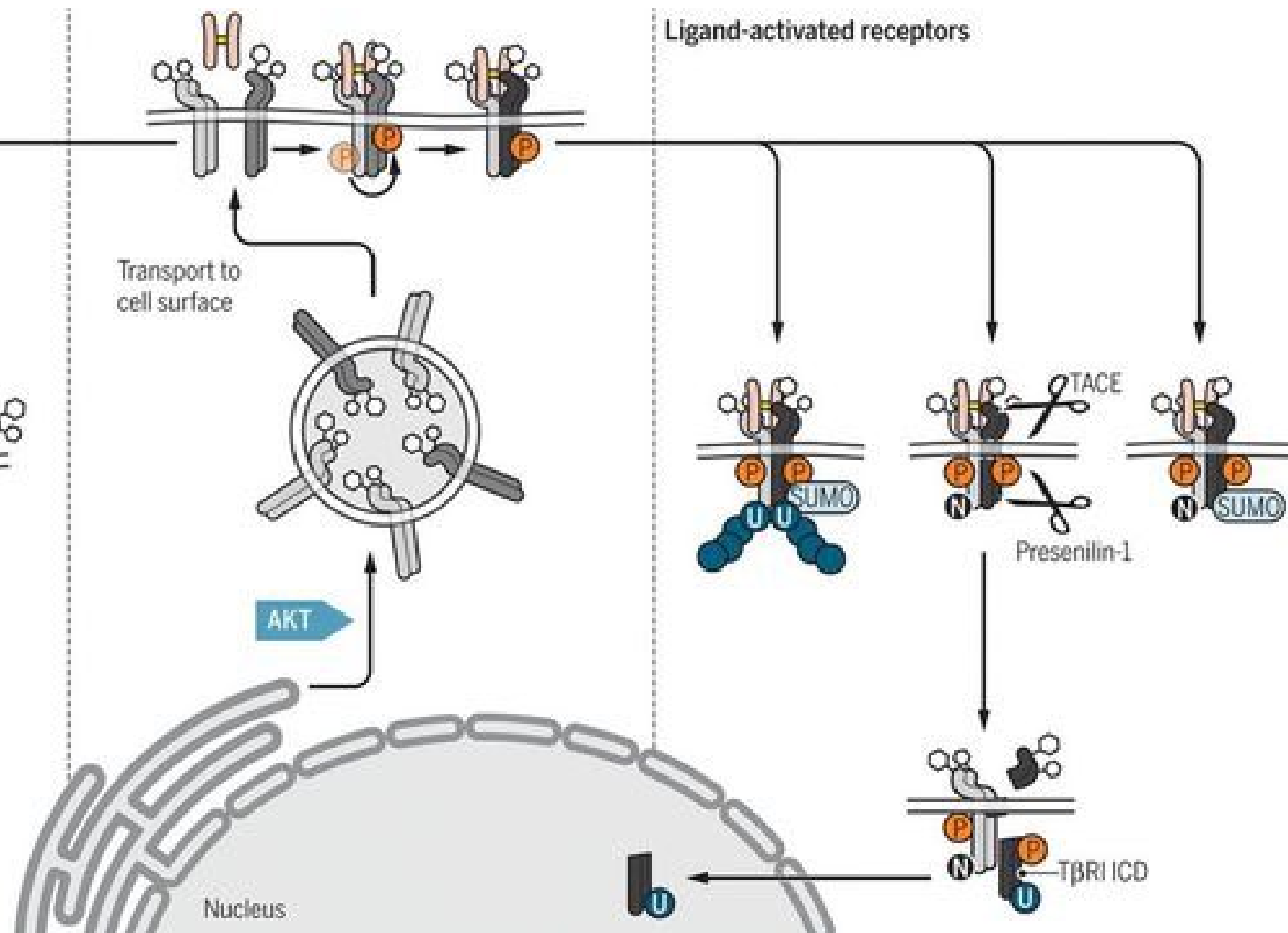
7 type I receptors
5 type II receptors



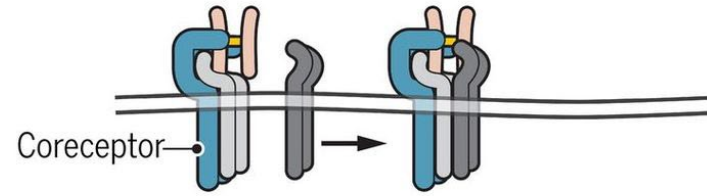
Ligand-free receptors



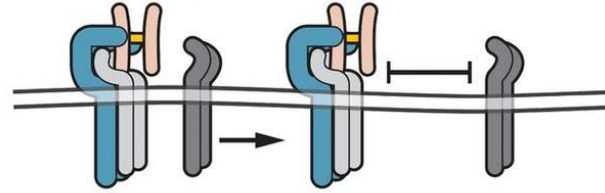
Ligand-activated receptors



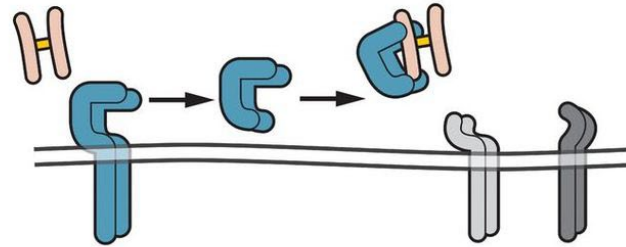
Promotes complex formation



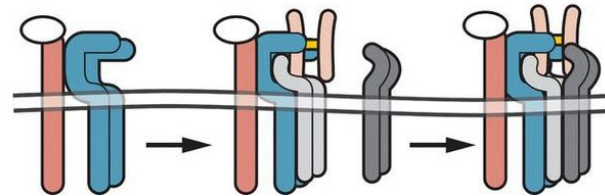
Interferes with complex formation



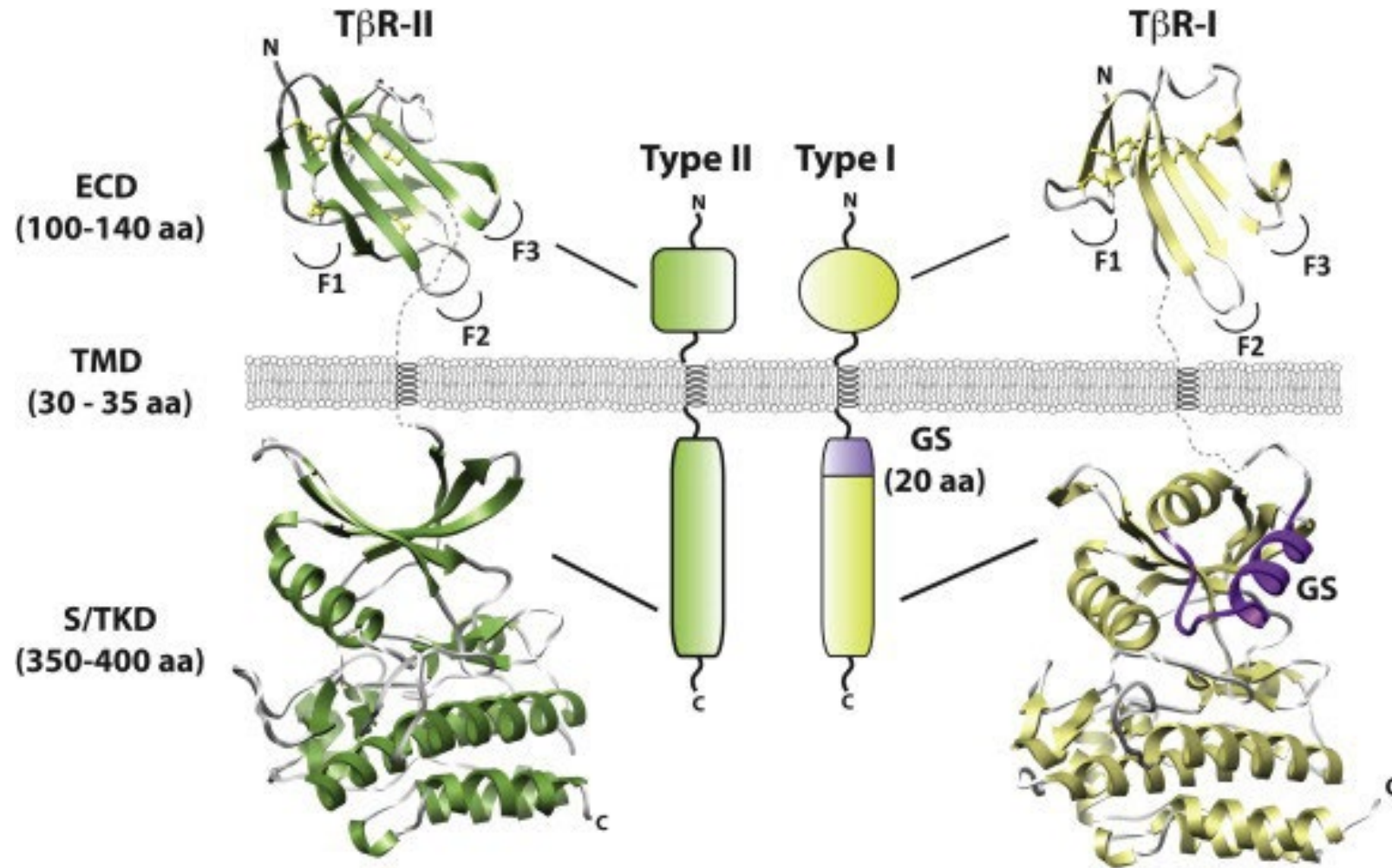
Sequesters soluble ligand



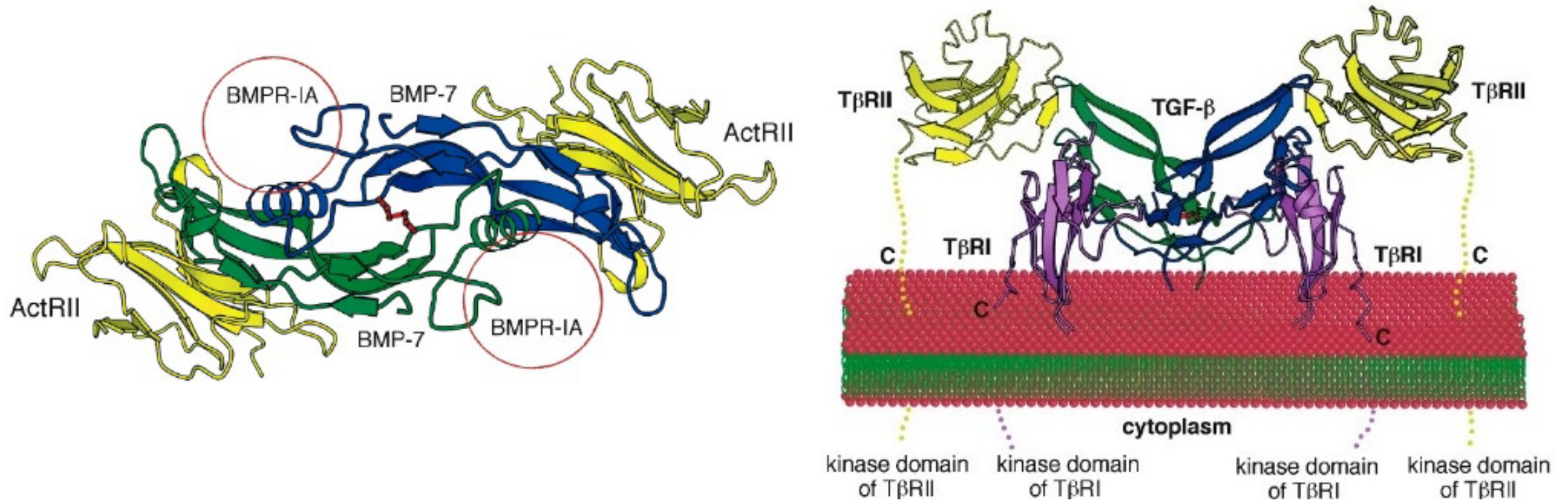
Engages with other signaling receptor



TGF- β type I and type II receptors form a heterotetramer – T β RII activates T β RI kinase

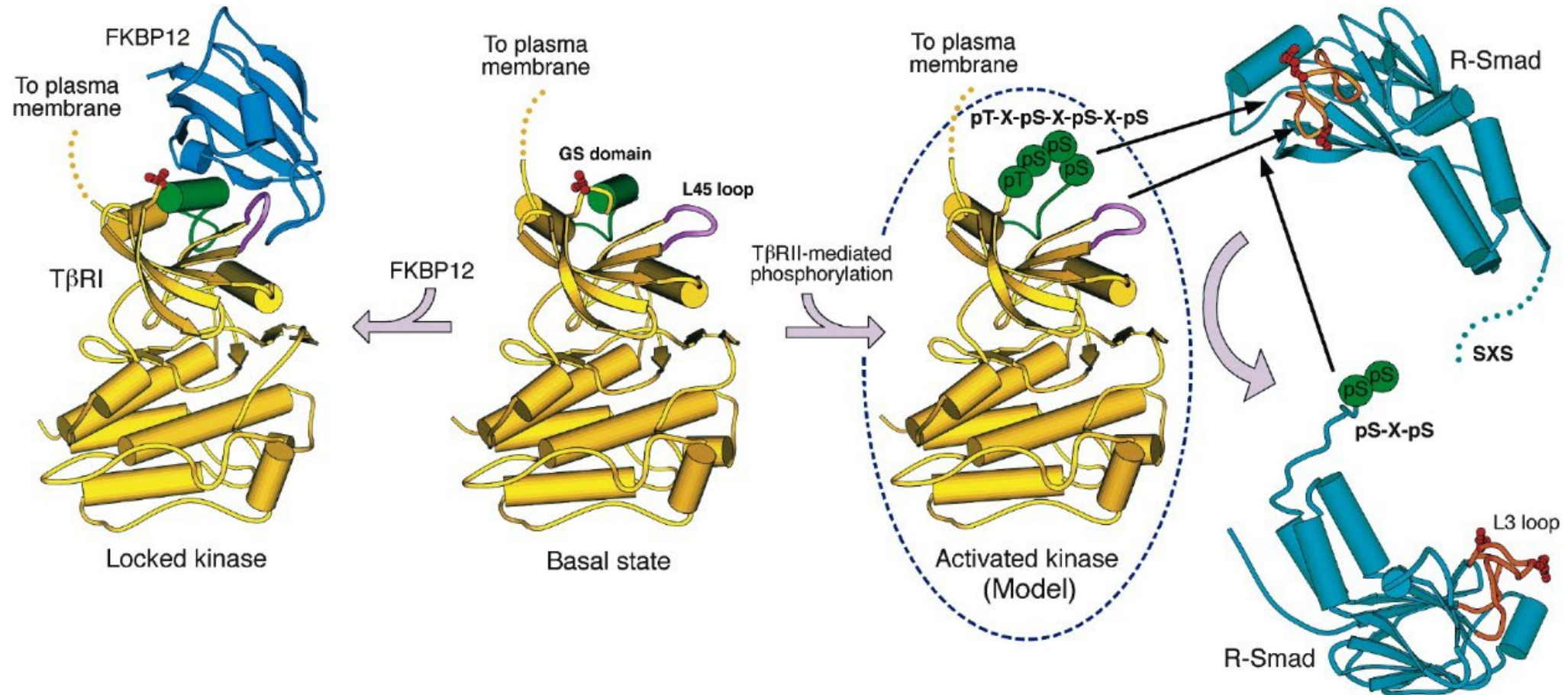


Assembly of Ligand-Receptor Complexes

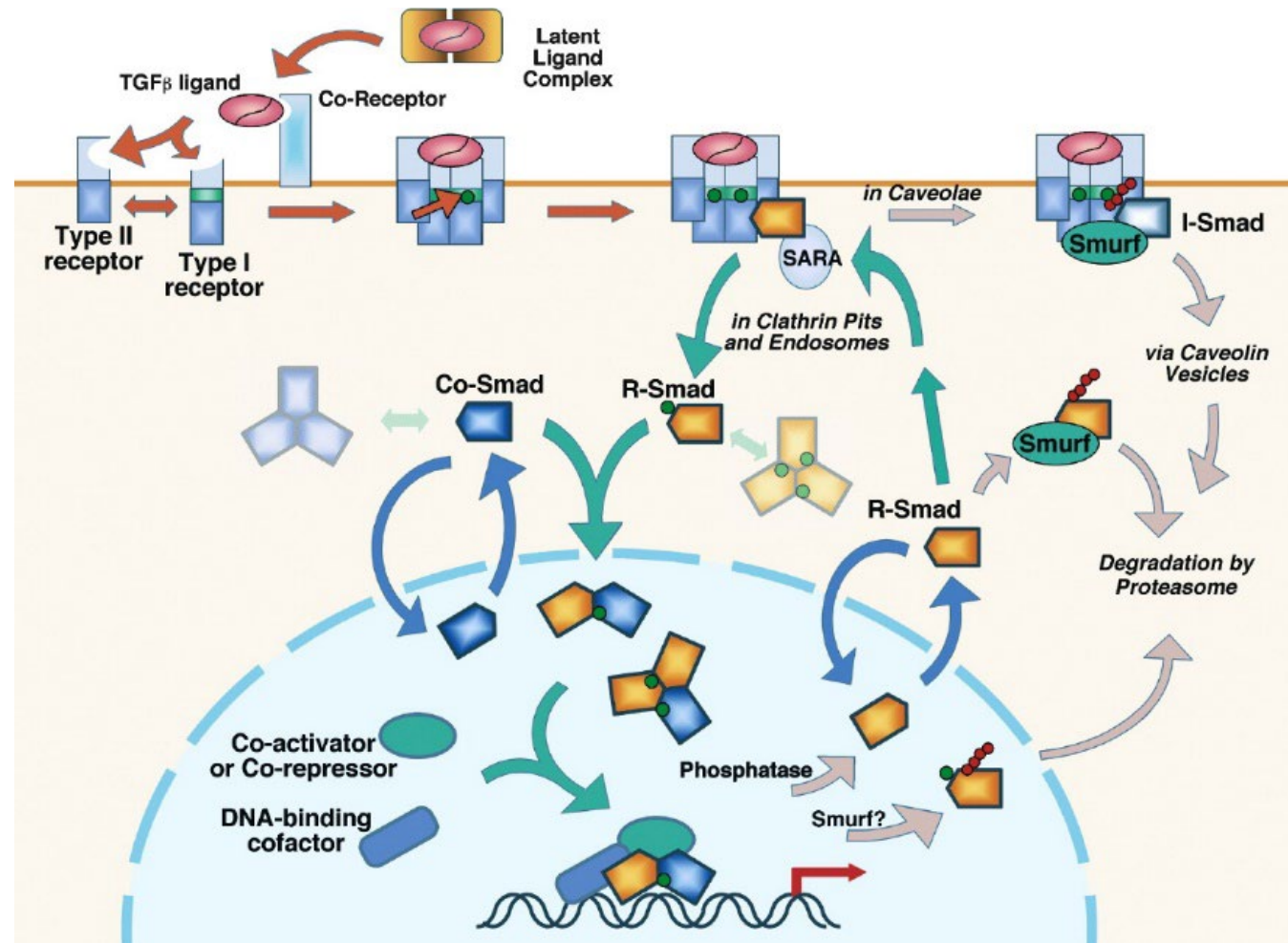


BMPs – bind to type I receptor first and then bind to type II receptor
TGF- β /Activins – bind to type II receptor first and then to type I

Regulation of S/T kinase activity



Signaling by TGF- β receptors

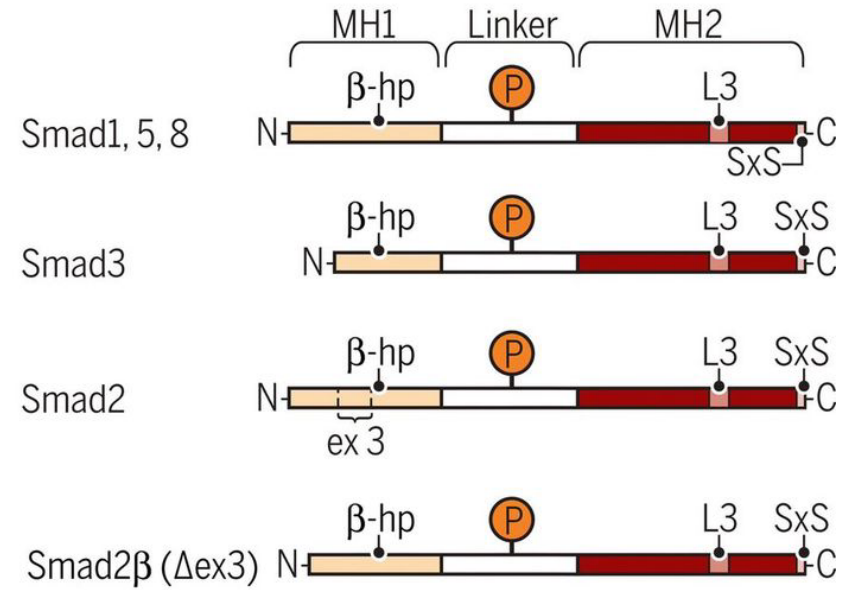


R-smad: 1,2,3,5,8

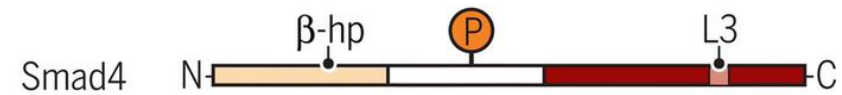
Co-Smad: 4

I-Smad: 6,7

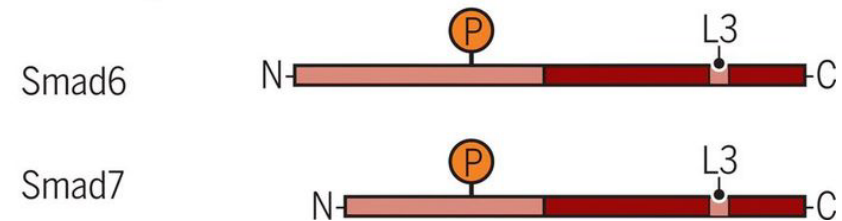
Receptor-activated Smads

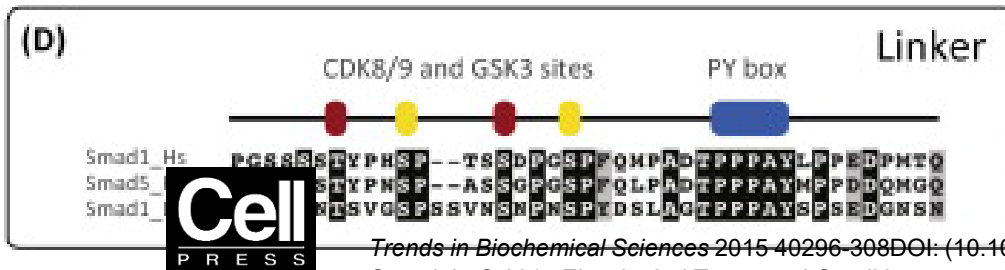
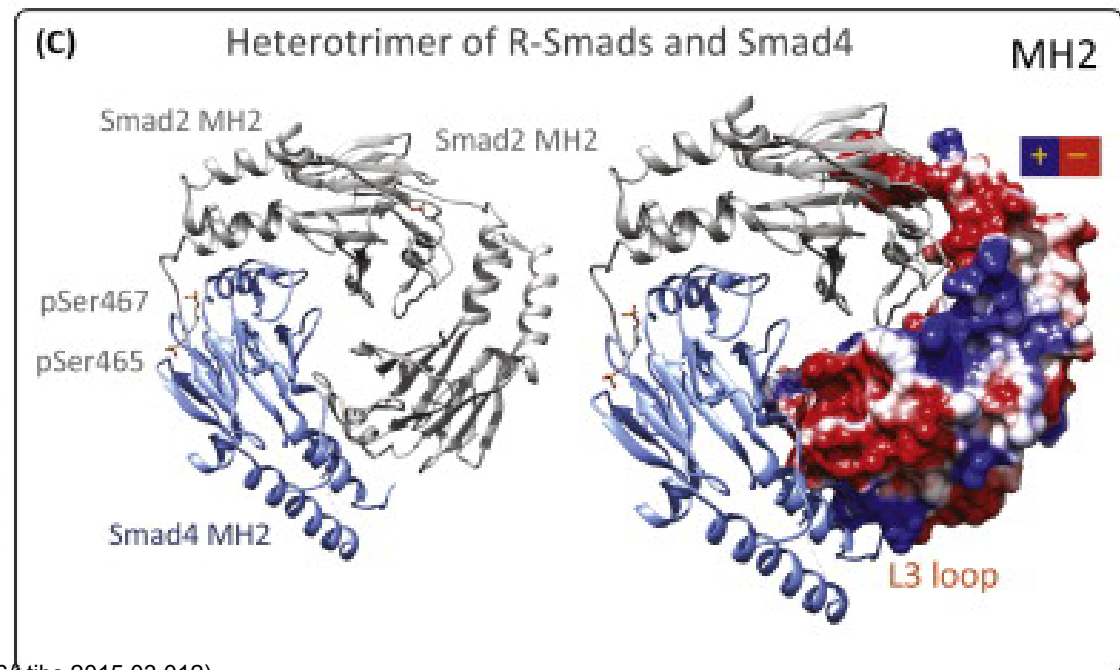
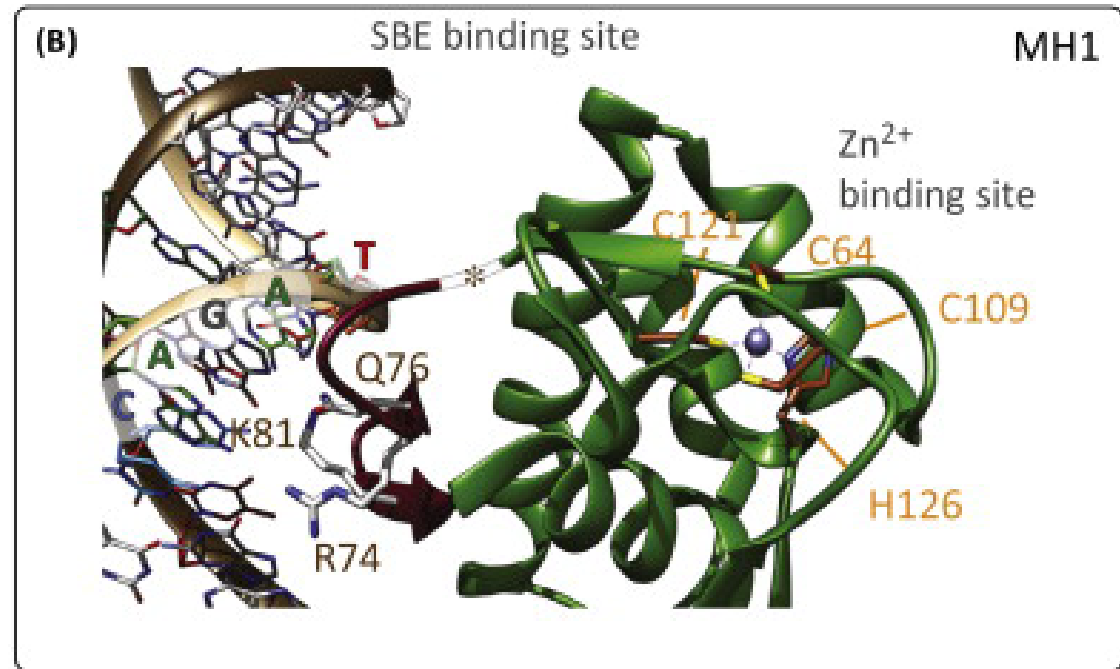
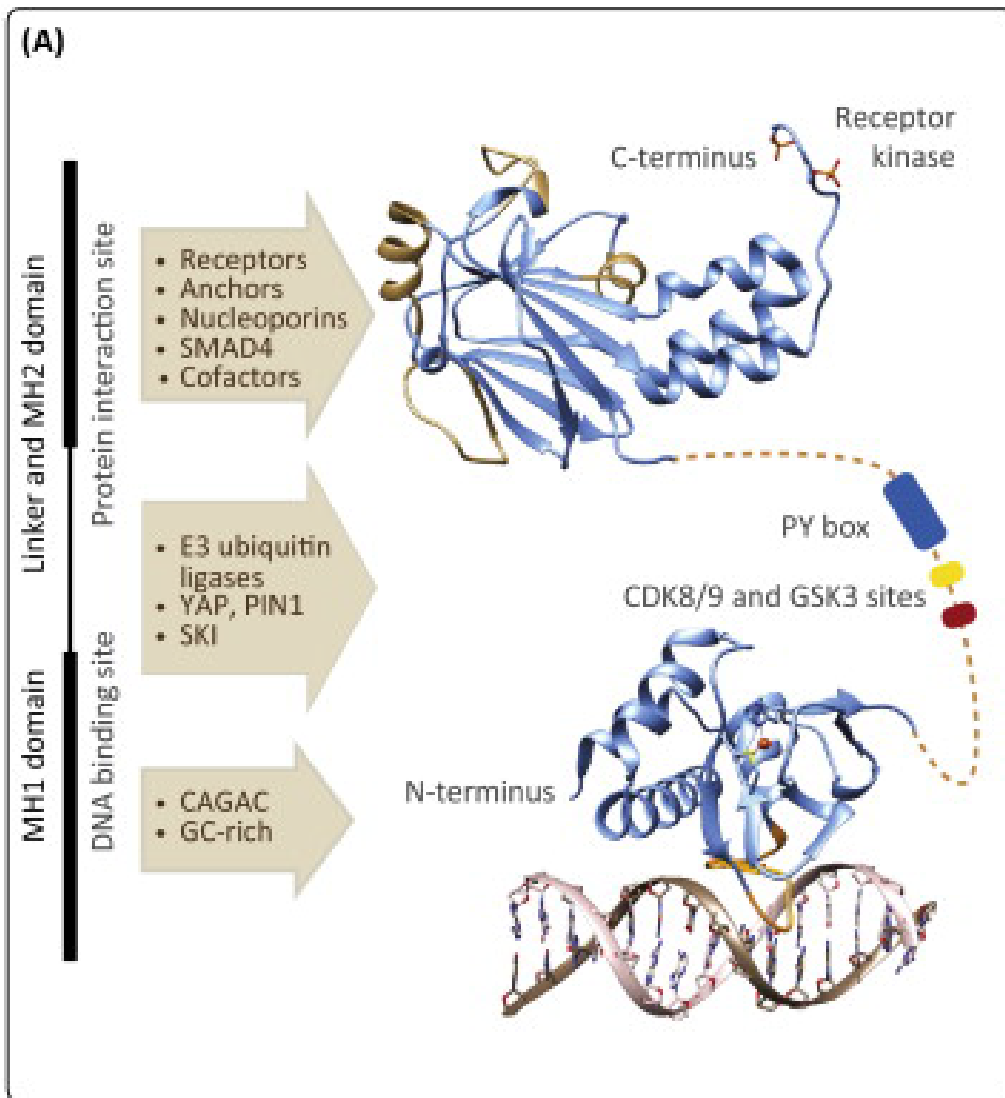


Common-mediator Smad

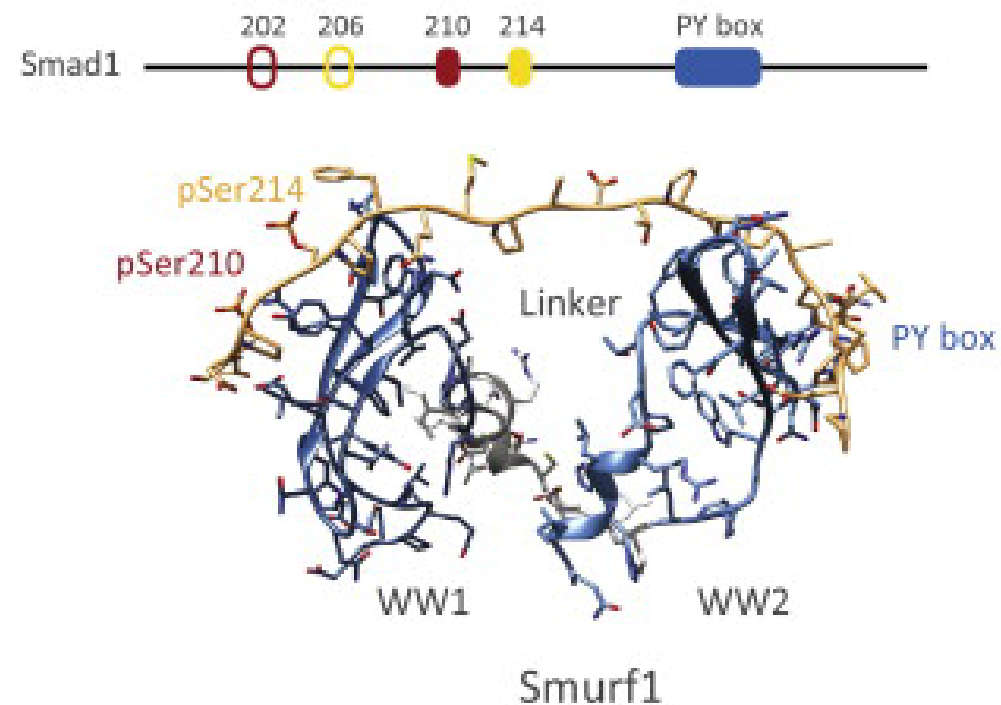
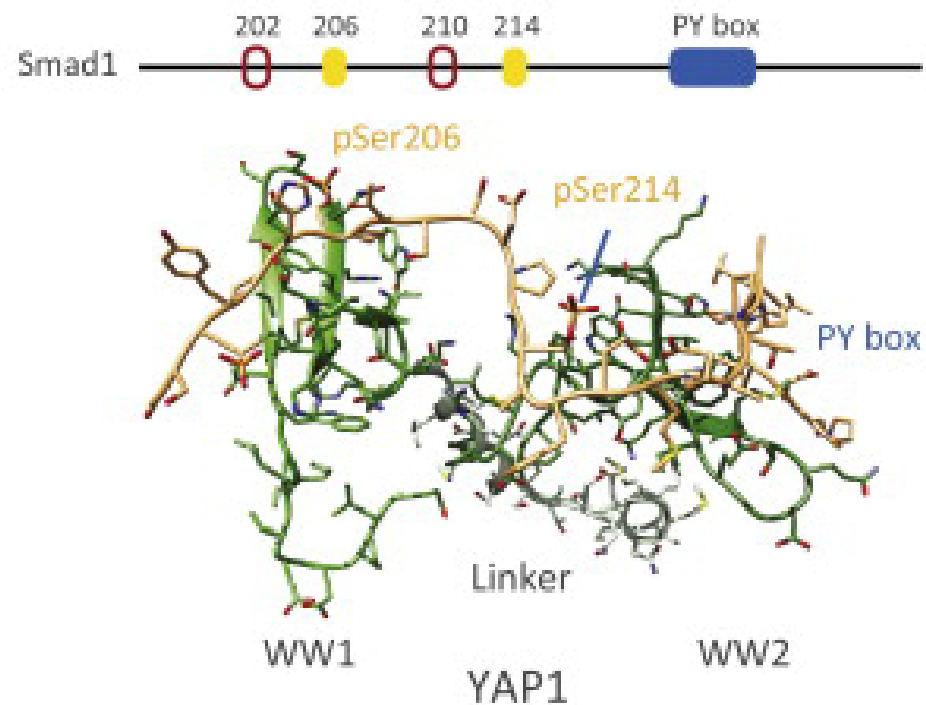


Inhibitory Smads

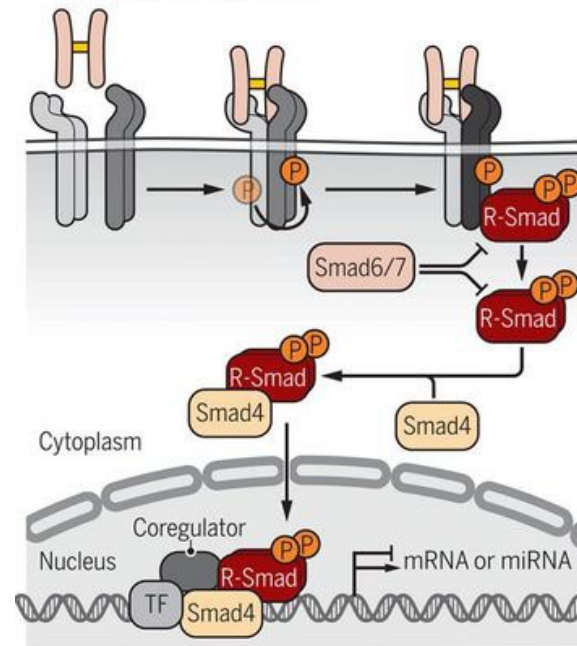
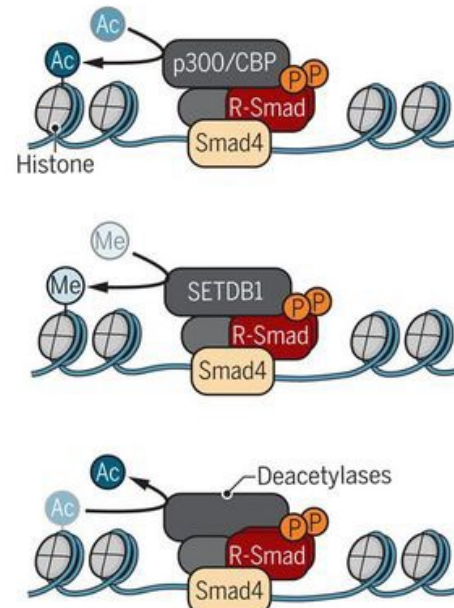
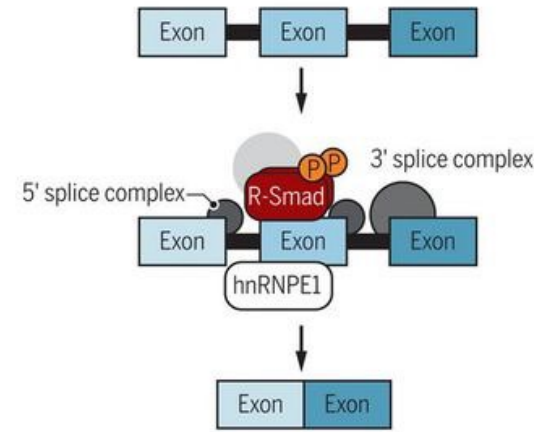
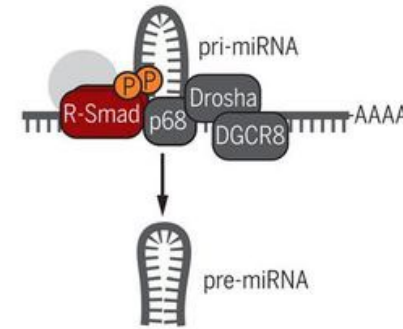
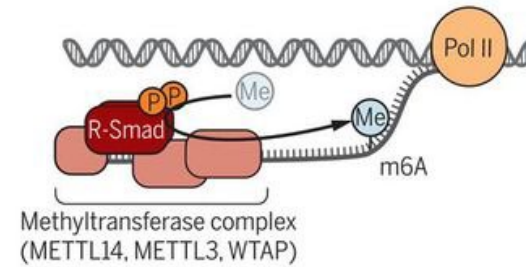


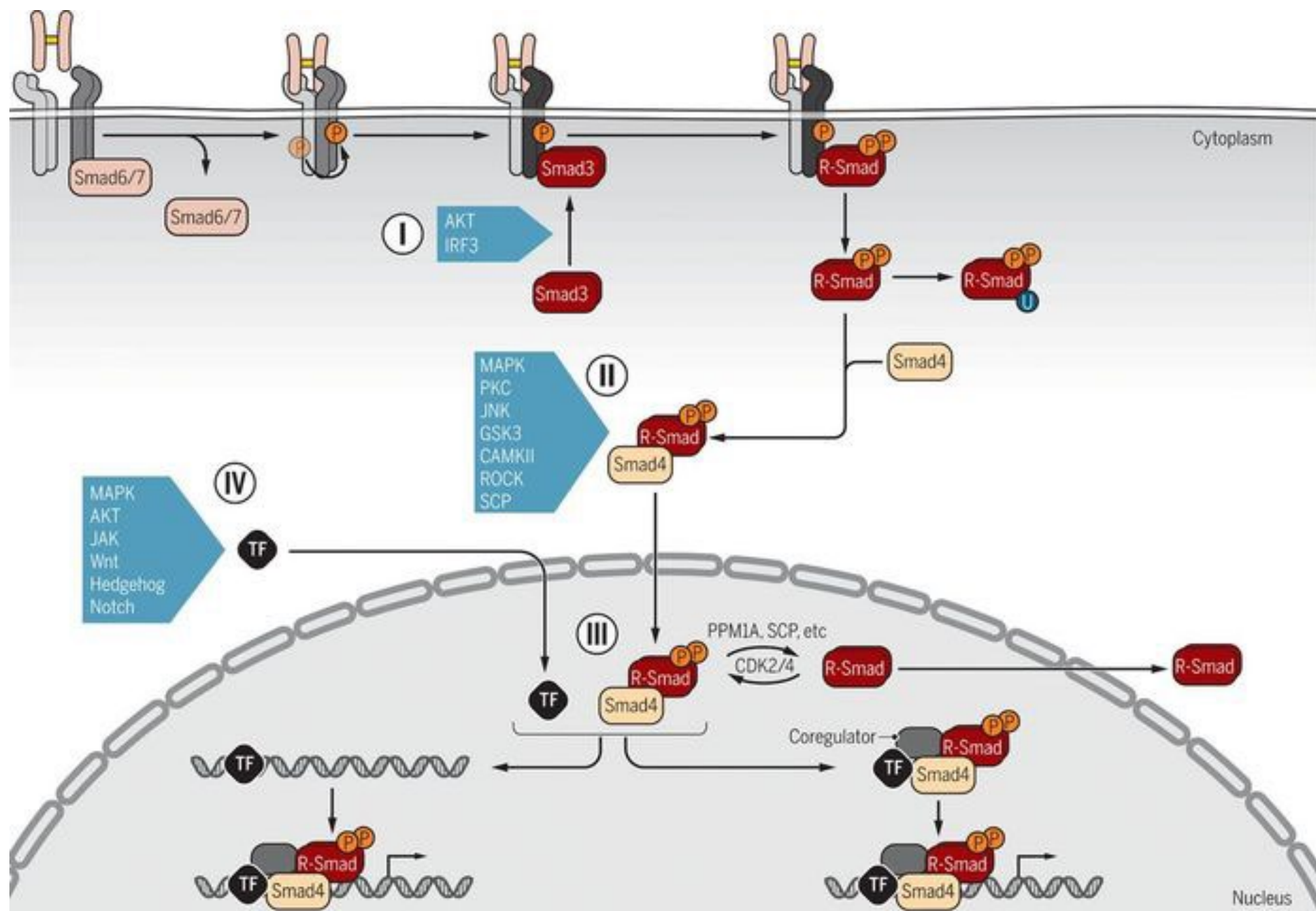


(E)



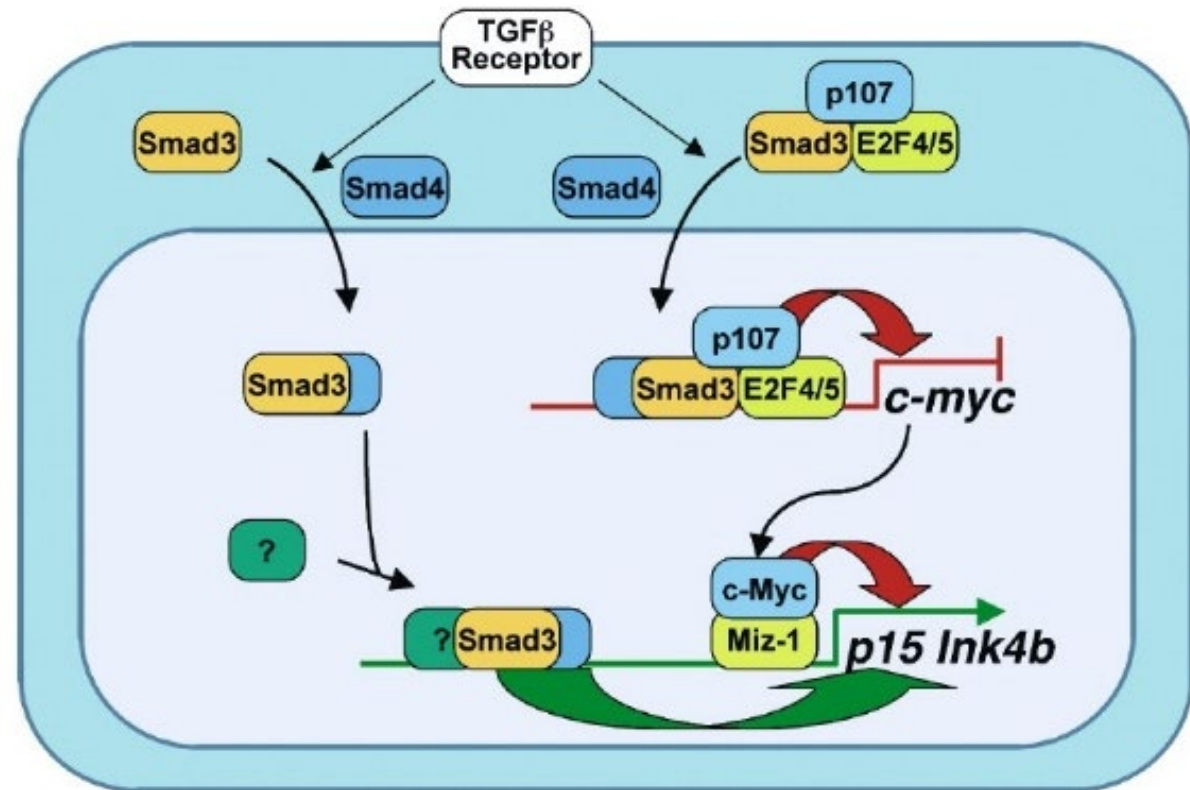
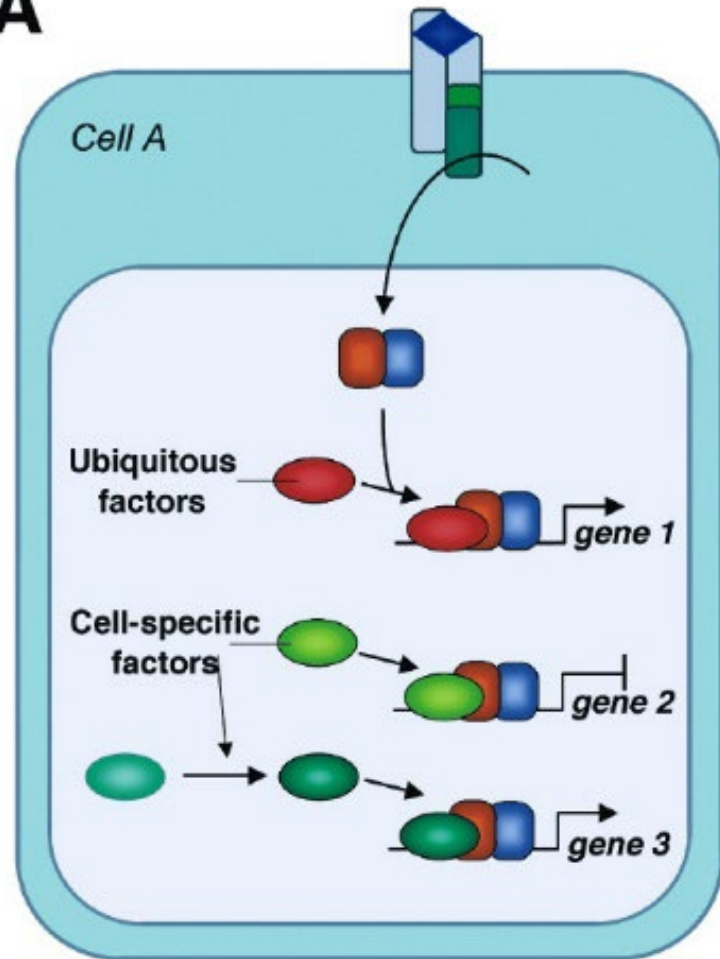
T/BS

A Transcription regulation**B Epigenetic remodeling****C RNA splicing****D miRNA processing****E m6A mRNA methylation**



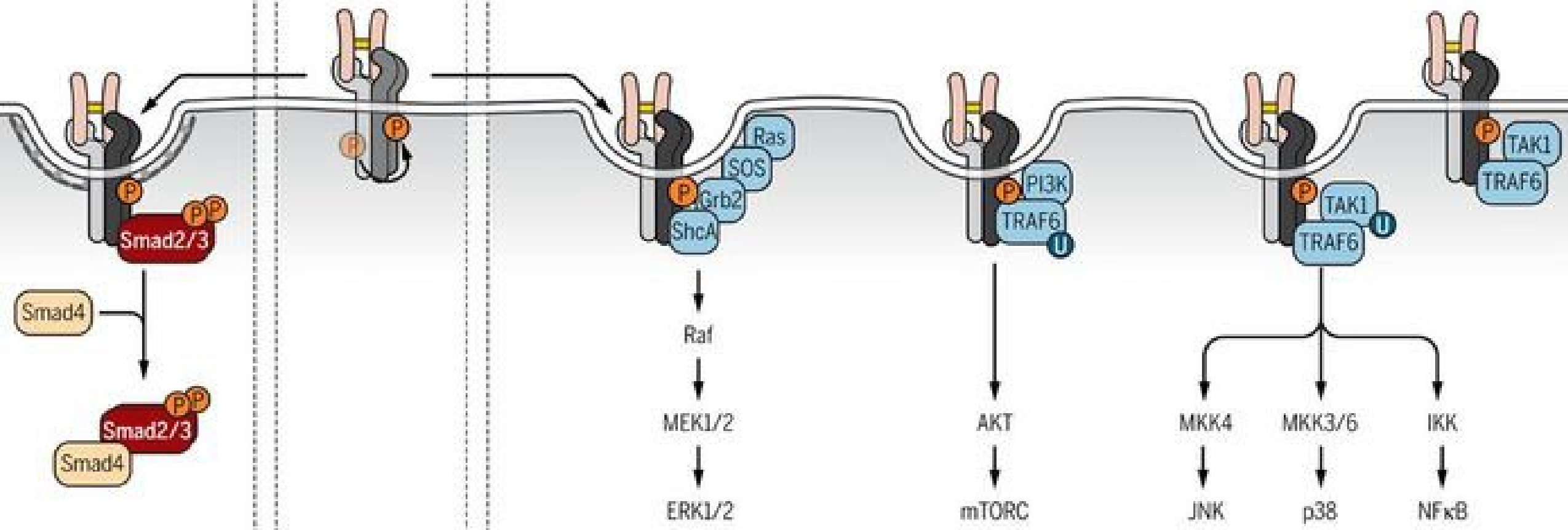
Context-dependent transcription by Smads

A



Clathrin-bound receptors

Caveolar receptors



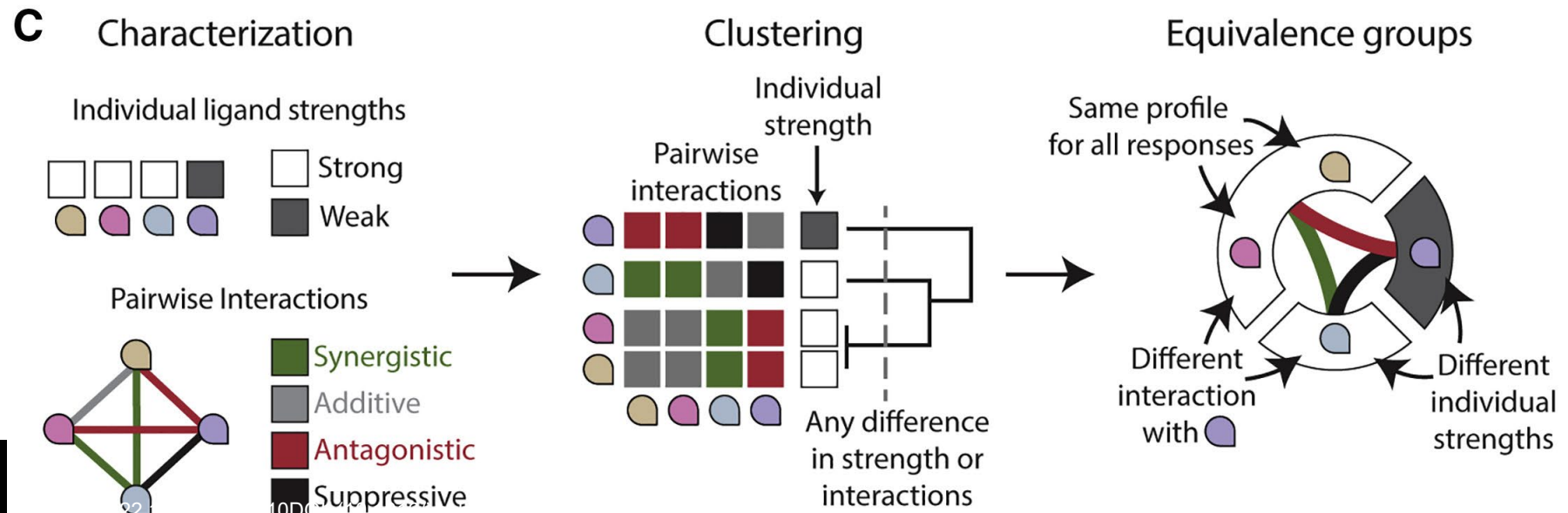
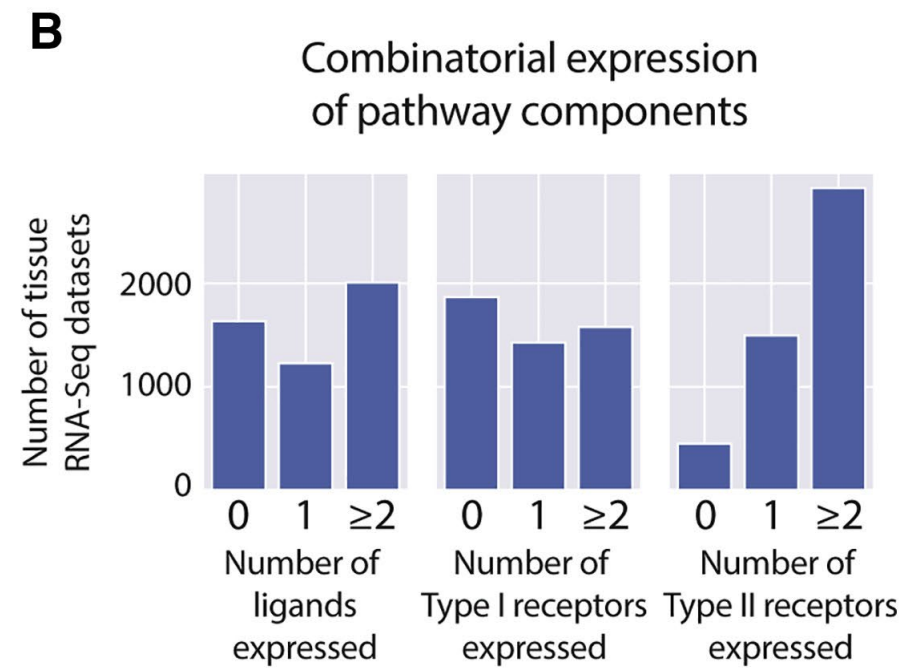
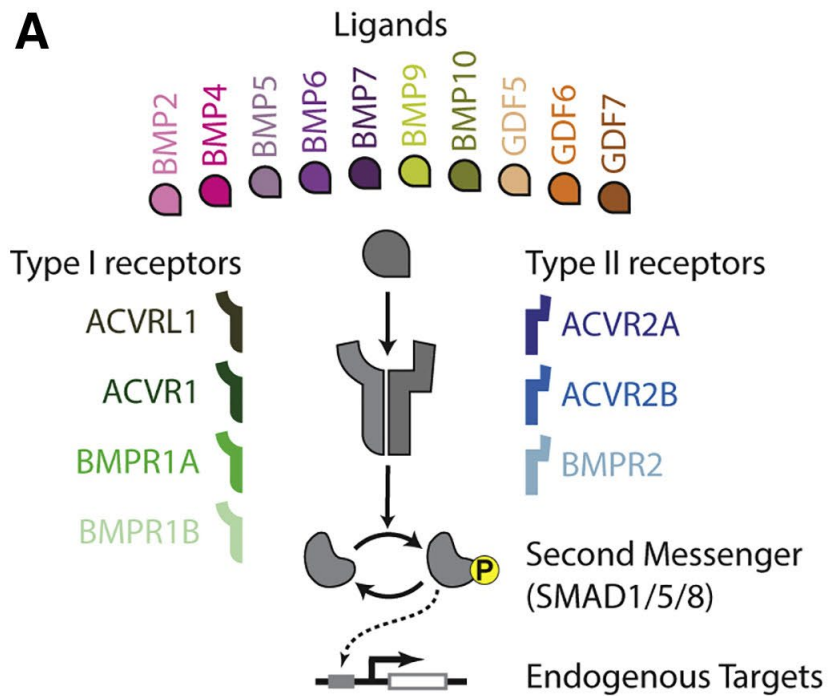
The context-dependent, combinatorial logic of BMP signaling

Heidi E. Klumpe, Matthew A. Langley, James M. Linton, Christina J. Su, Yaron E. Antebi,
Michael B. Elowitz

Cell Systems

Volume 13 Issue 5 Pages 388-407.e10 (May 2022)

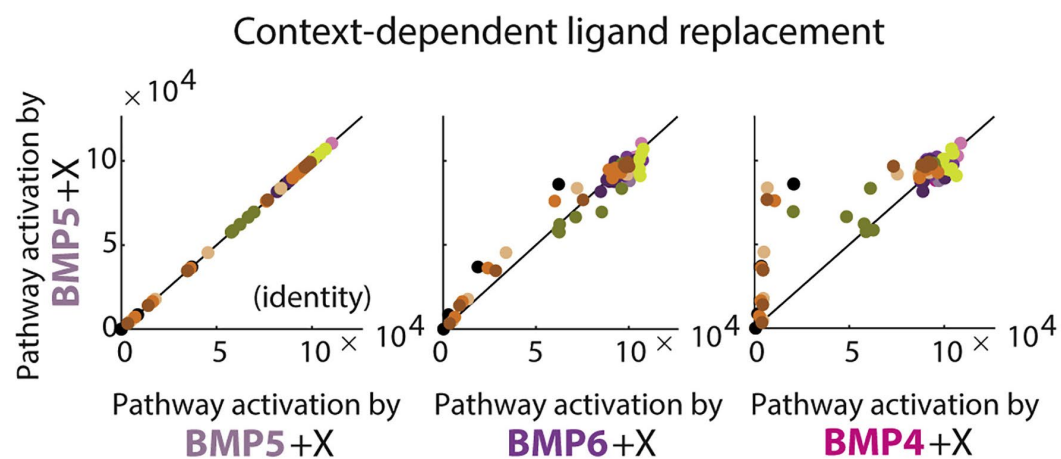
DOI: [10.1016/j.cels.2022.03.002](https://doi.org/10.1016/j.cels.2022.03.002)



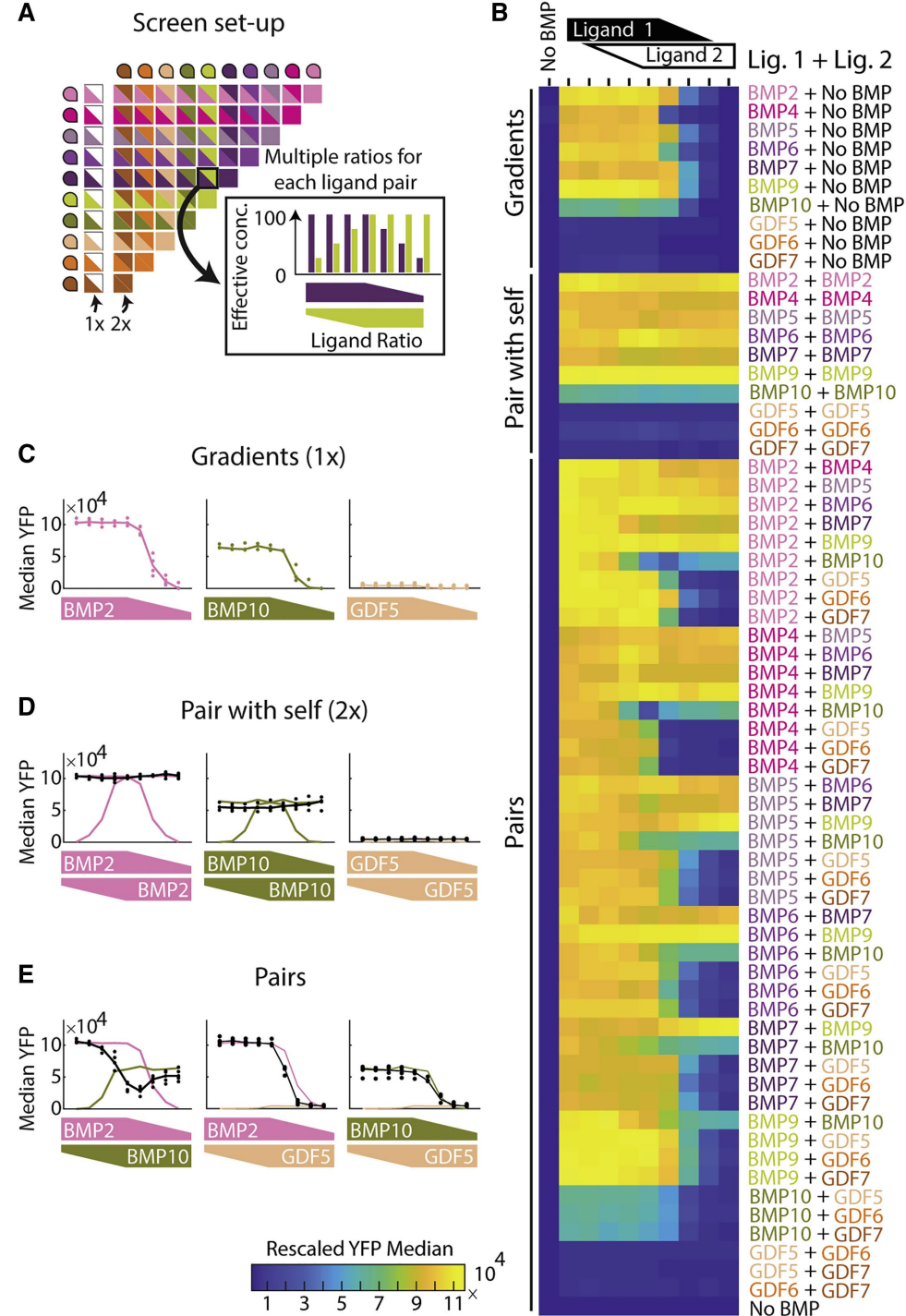
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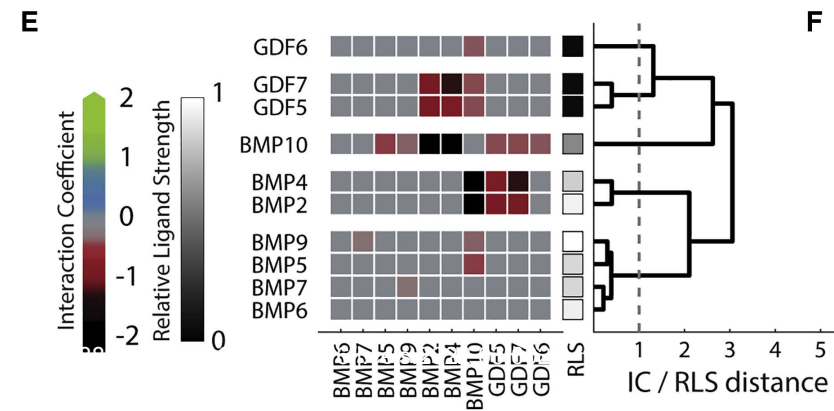
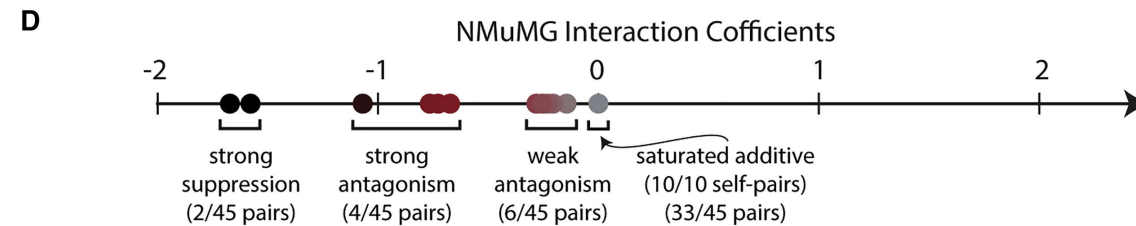
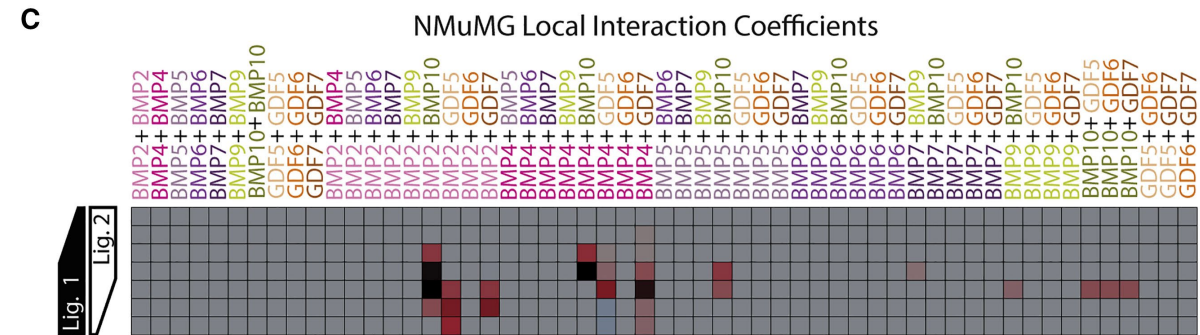
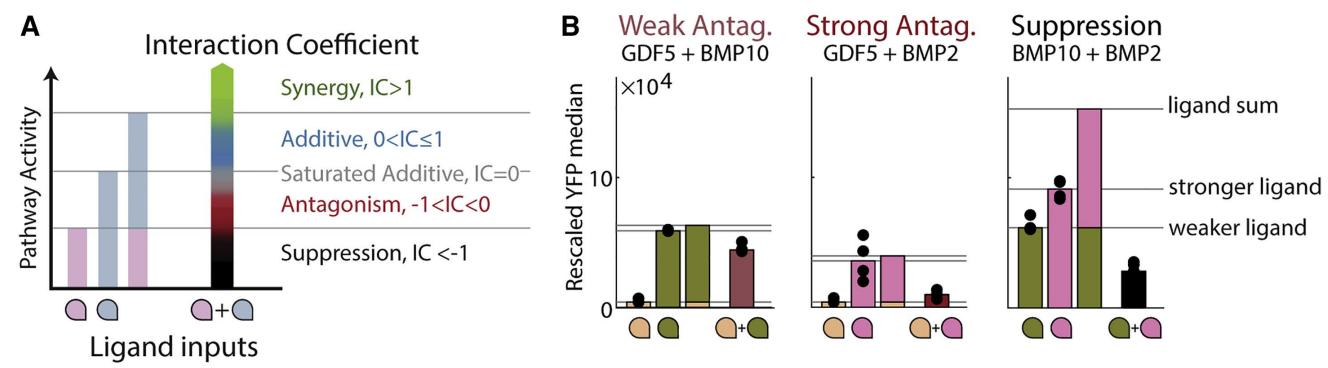
X =

- No BMP
- BMP2
- BMP4
- BMP5
- BMP6
- BMP7
- BMP9
- BMP10
- GDF5
- GDF6
- GDF7

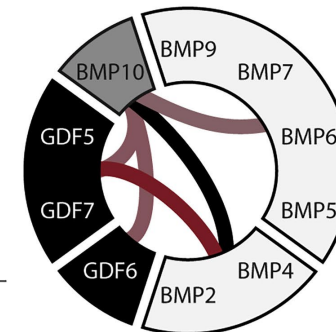


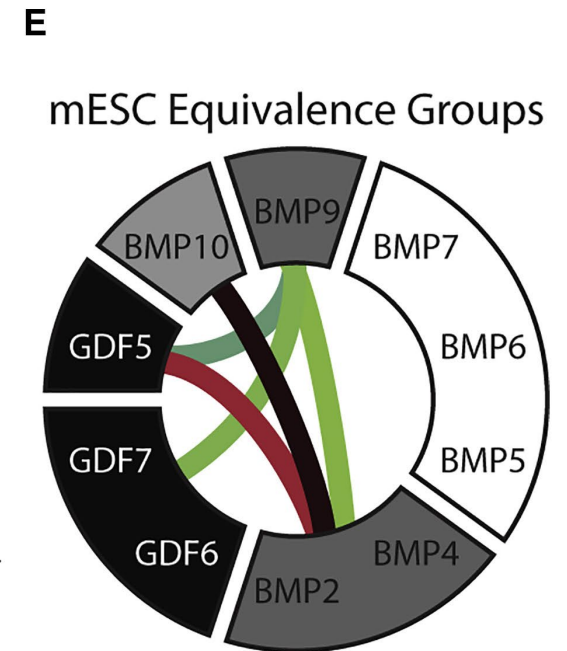
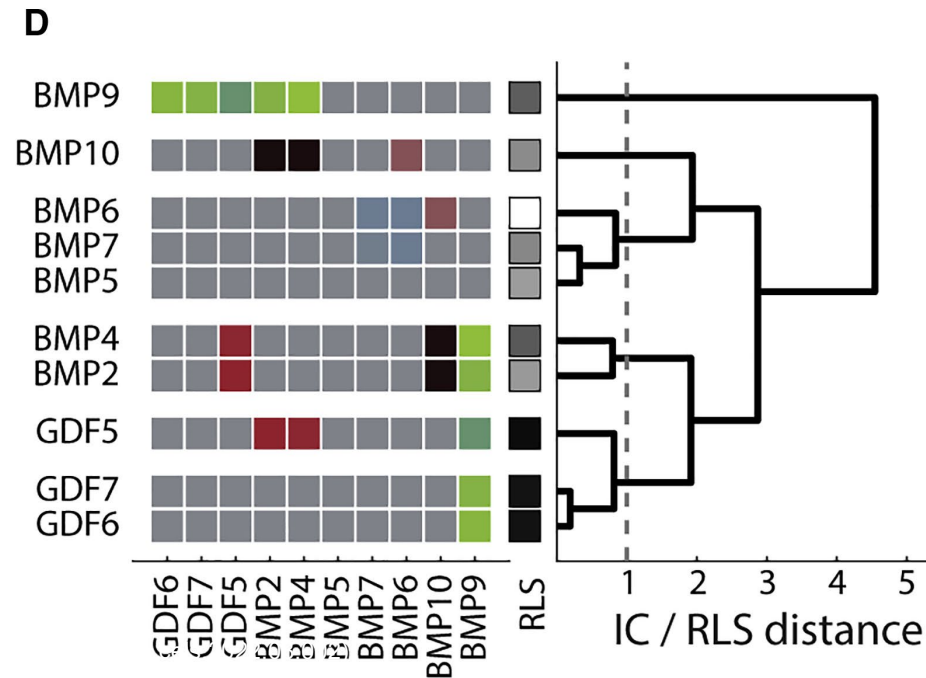
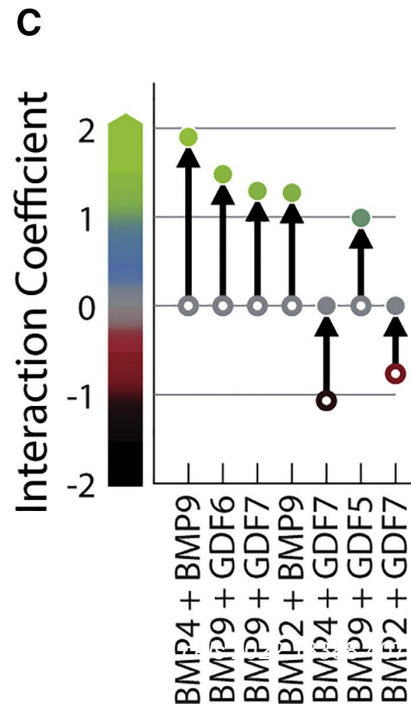
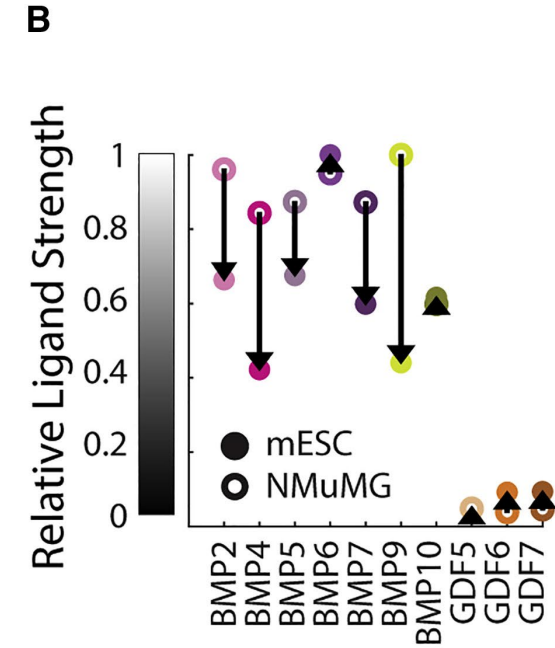
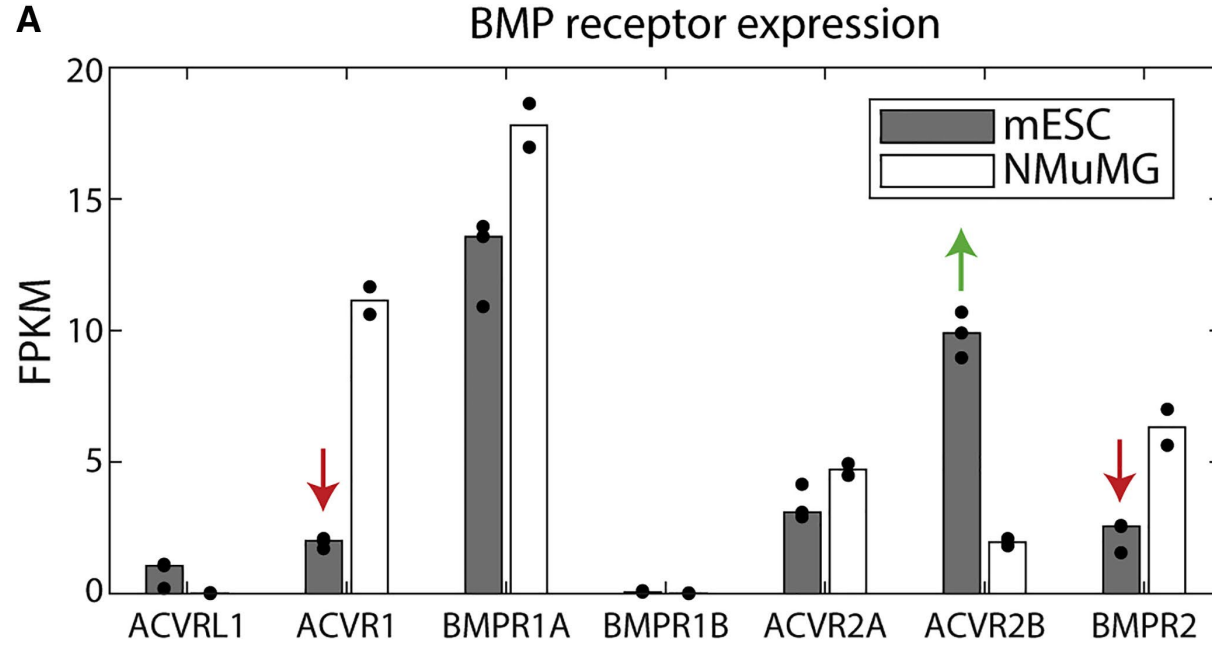
Terms and Conditions

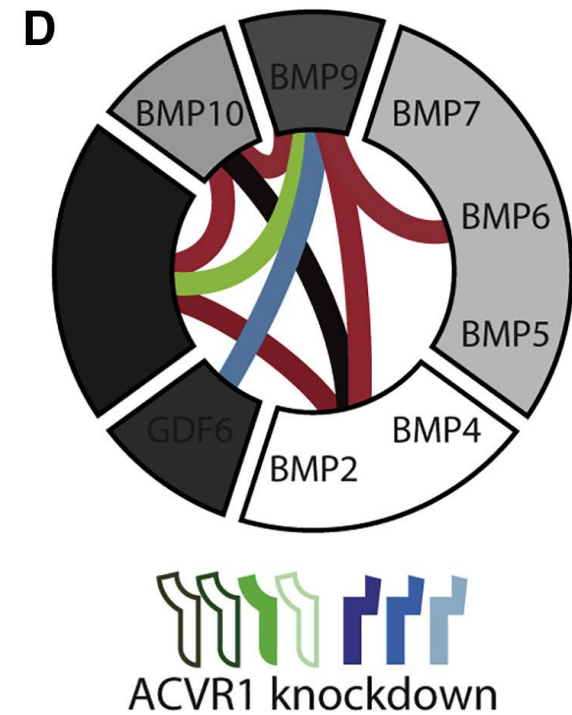
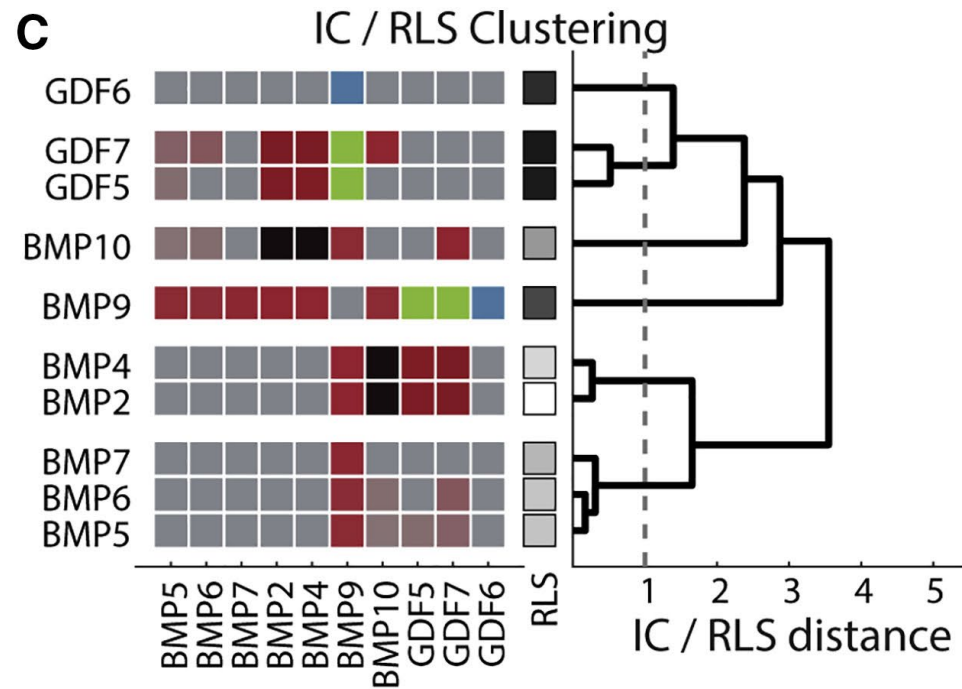
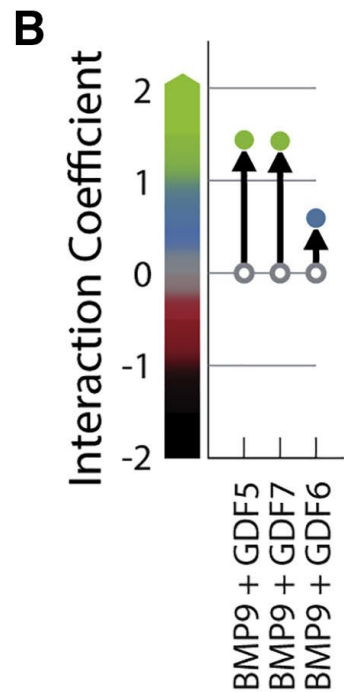
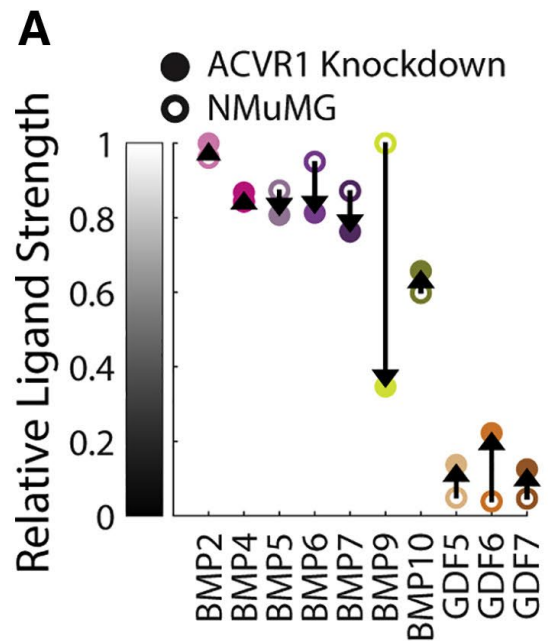


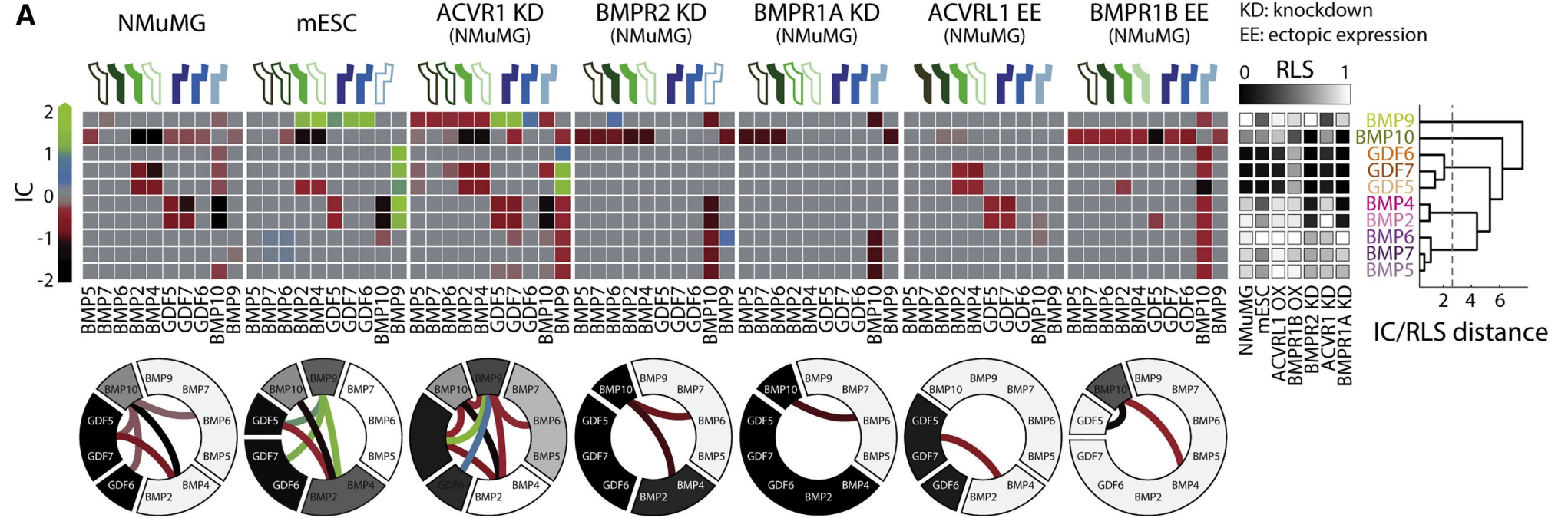


F NMuMG Equivalence Groups

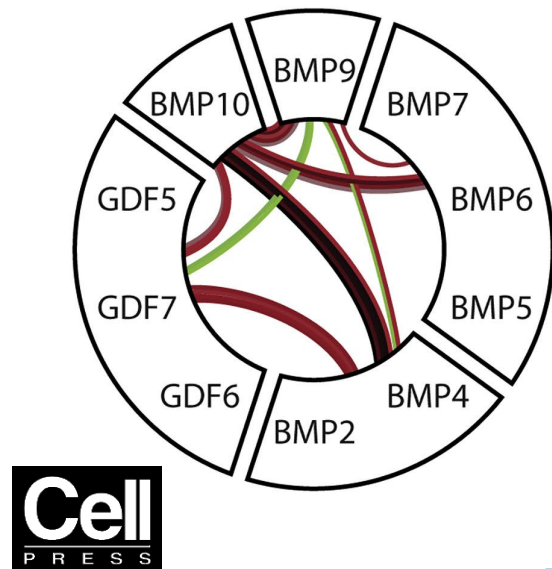




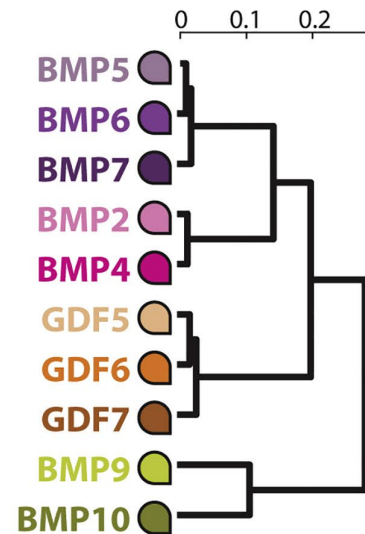




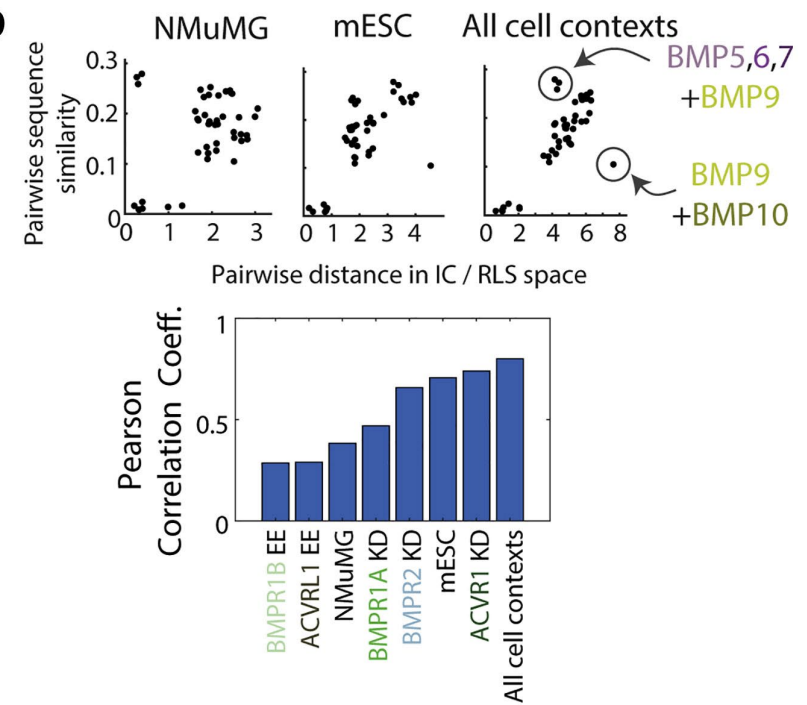
B Global ligand equivalence



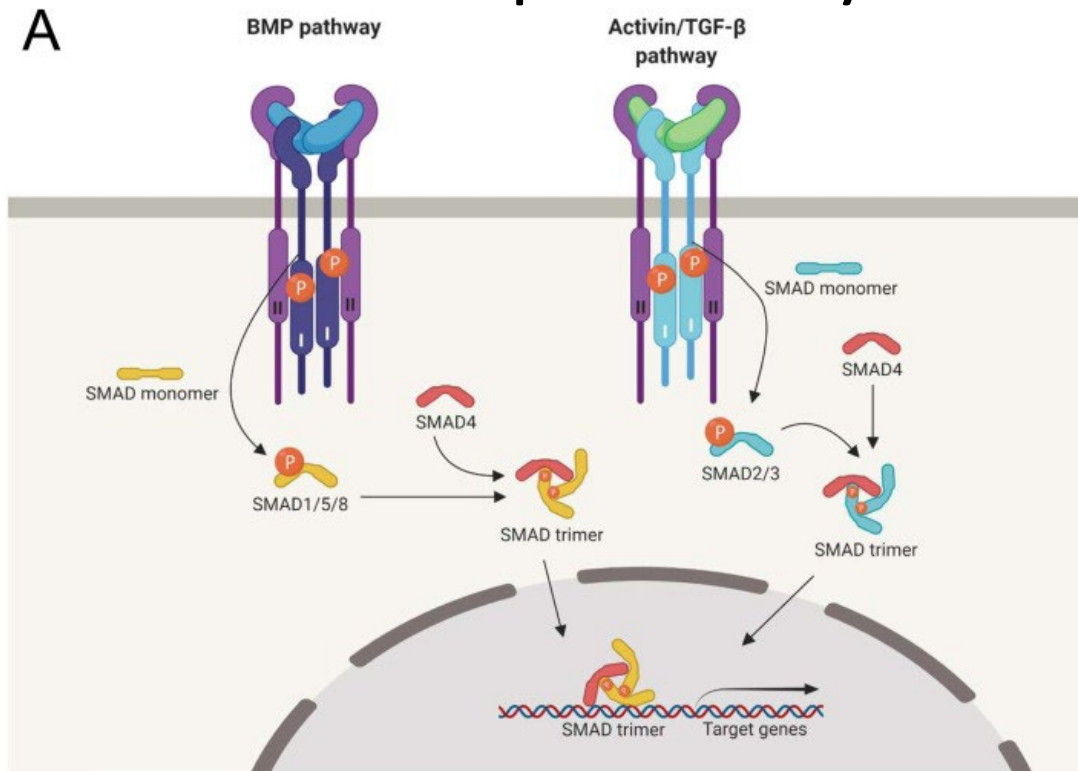
C Sequence similarity



D



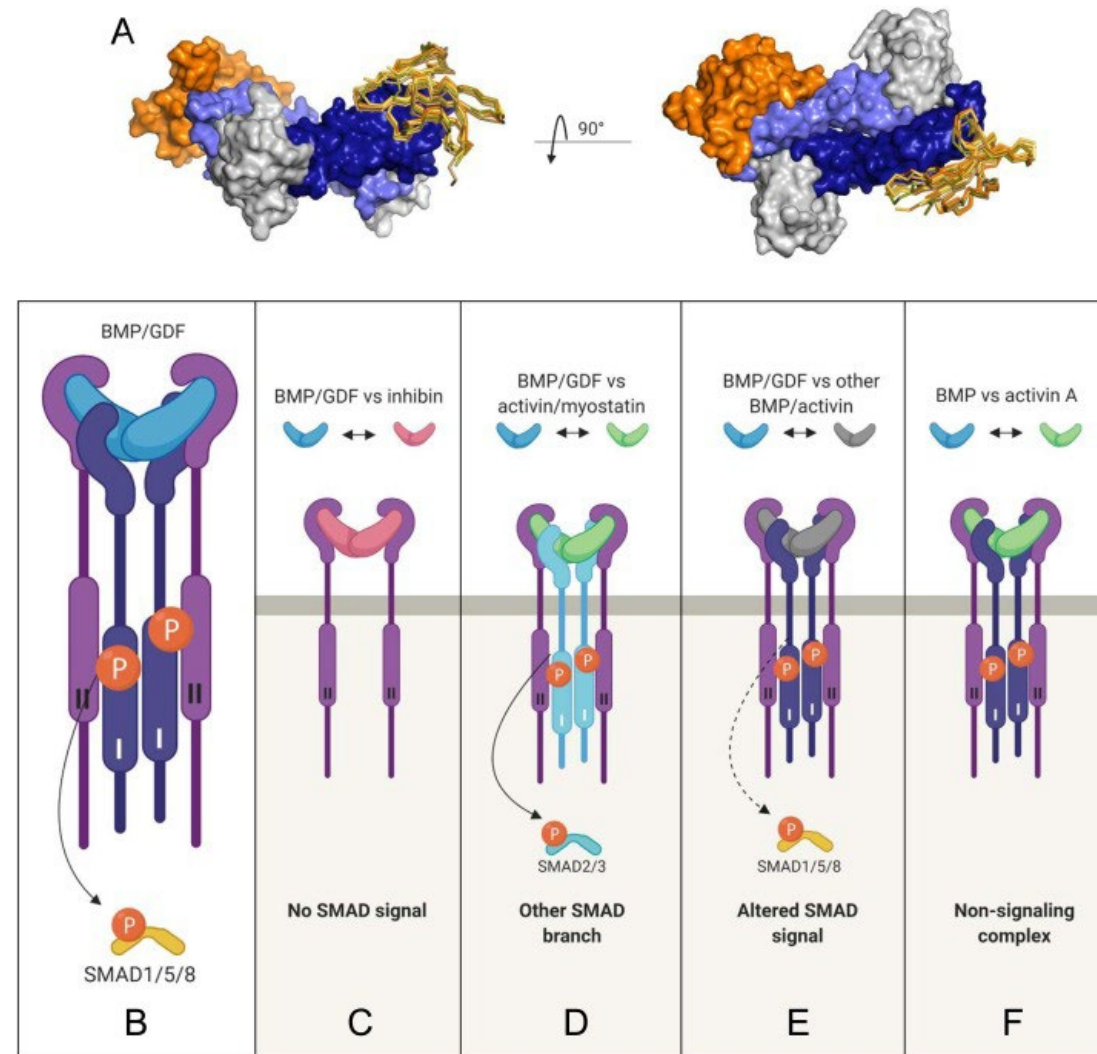
Receptor binding competition as a paradigm for TGF- β family action



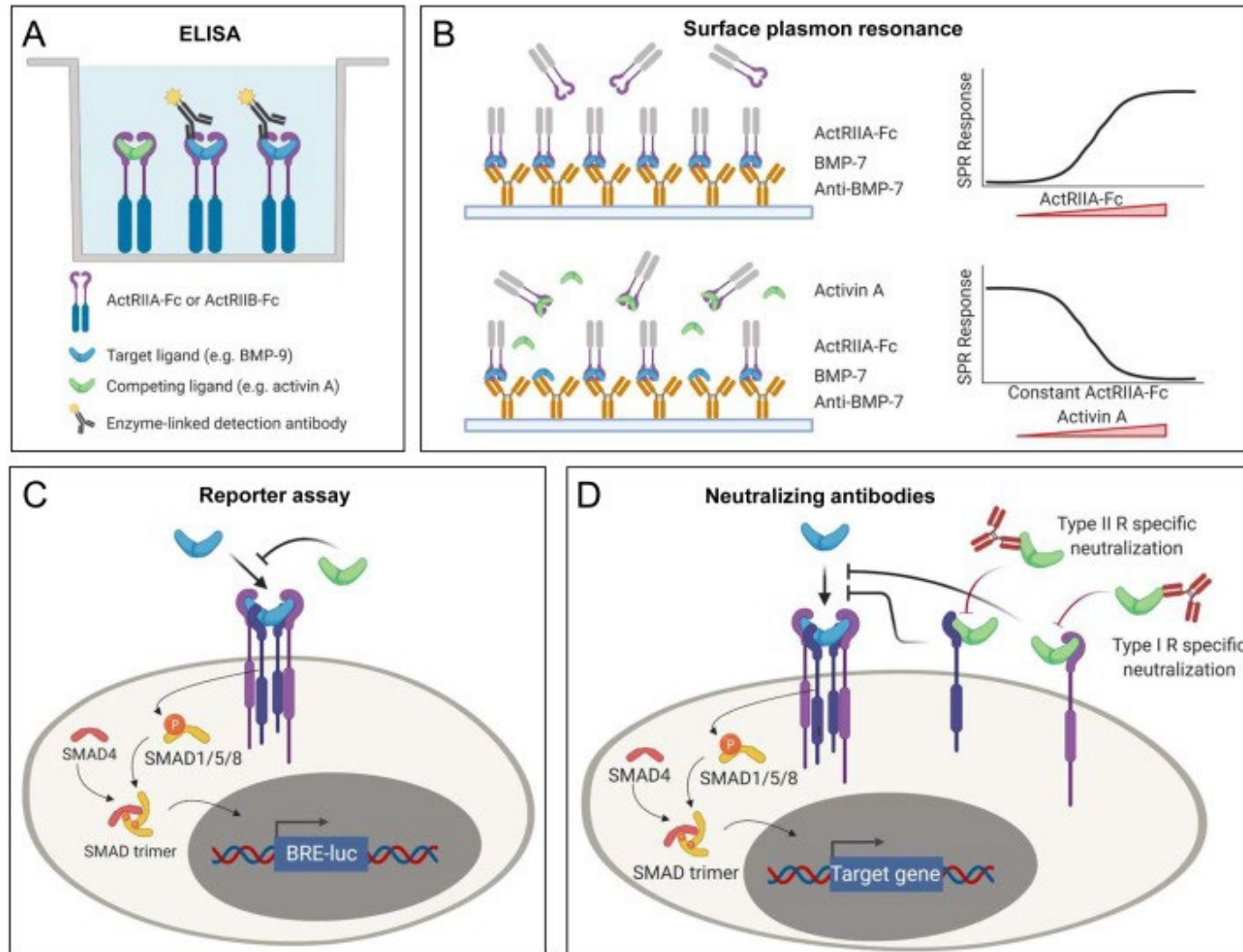
B

Ligands	Type II receptor	Type I receptor	R-SMAD
BMP pathway > 20 ligands	BMPRII (BMPRII) ActRIIA (ACVR2A) ActRIIB (ACVR2B) AMHRII (AMHR2)	ALK1 (ACVRL1) ALK2 (ACVR1) ALK3 (BMPRI1A) ALK6 (BMPRI1B)	SMAD1 SMAD5 SMAD8
Activin/TGF-β pathway > 15 ligands	ActRIIA (ACVR2A) ActRIIB (ACVR2B) TGF β RII (TGFBRI2)	ALK4 (ACVR1B) ALK7 (ACVR1C) ALK5 (TGFBRI1)	SMAD2 SMAD3

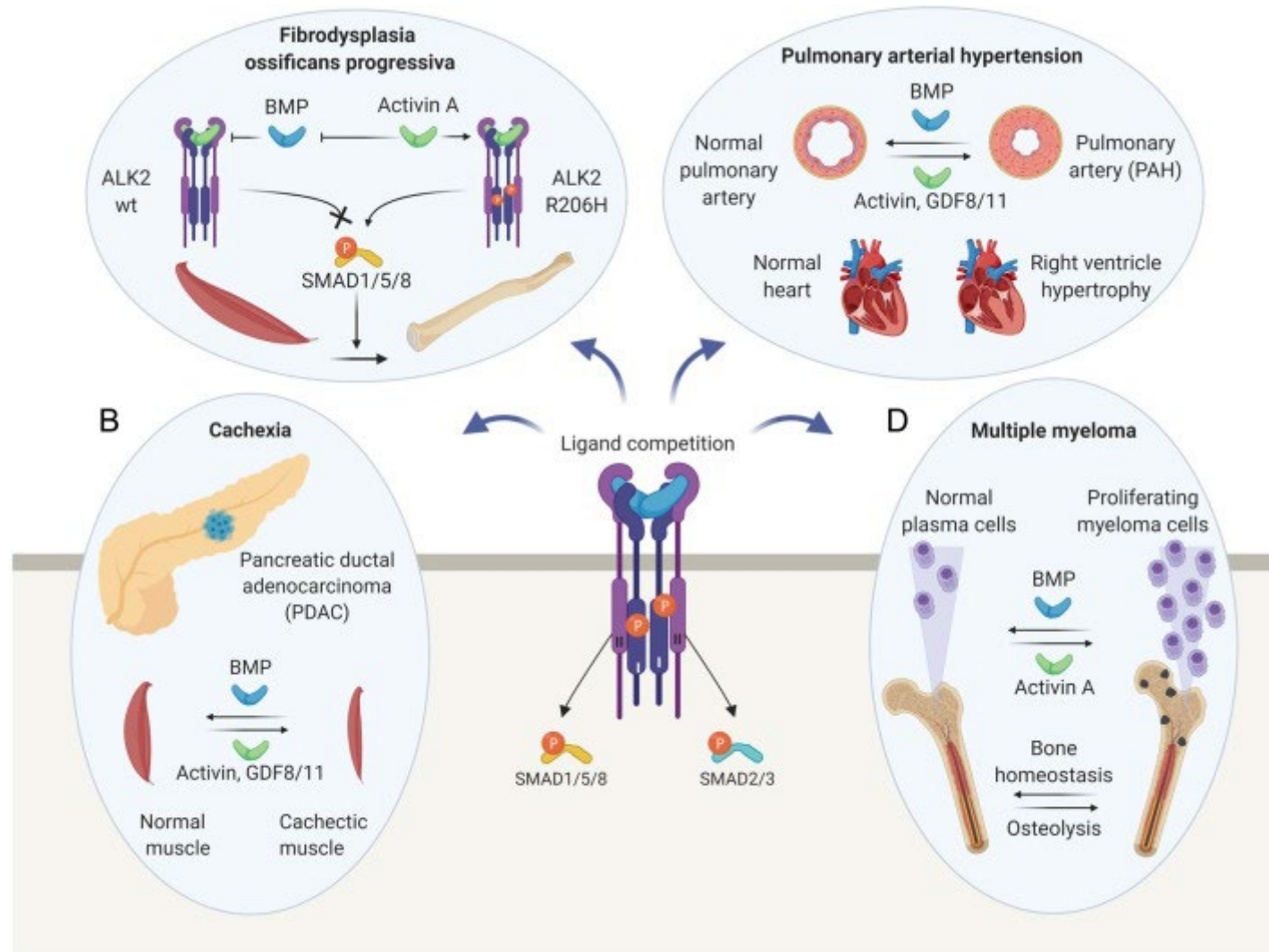
Promiscuous binding by TGF- β ligands



Measuring ligand competition

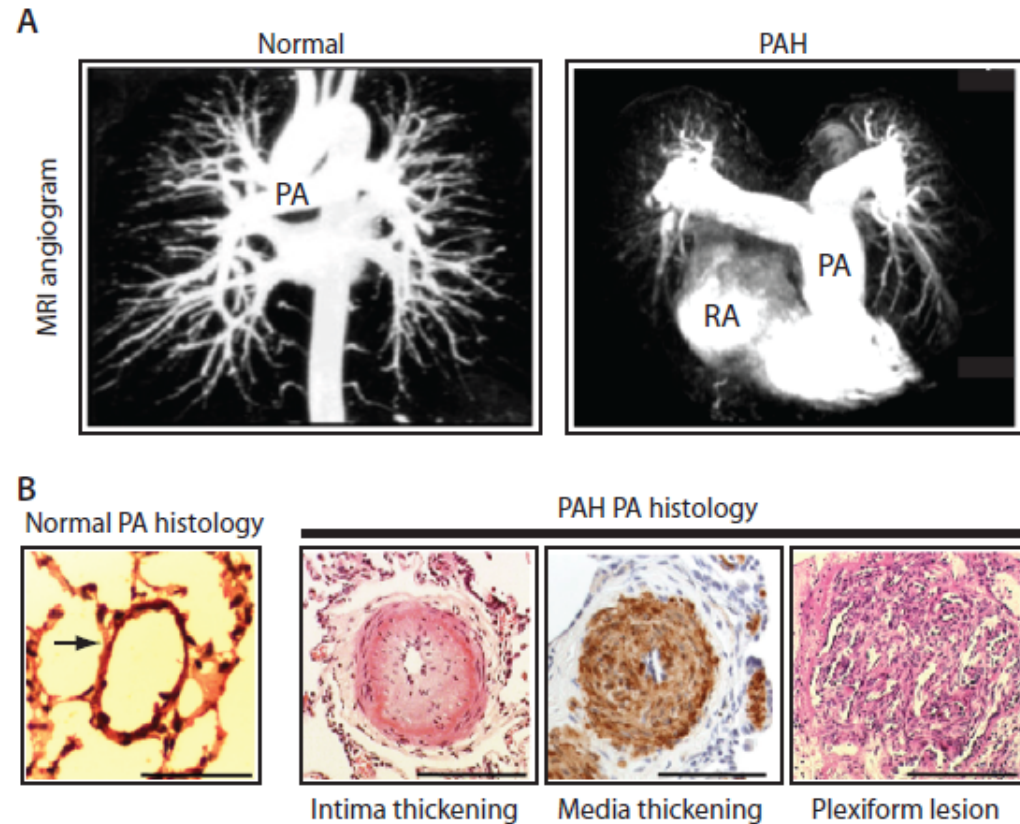


Pathological roles of ligand competition

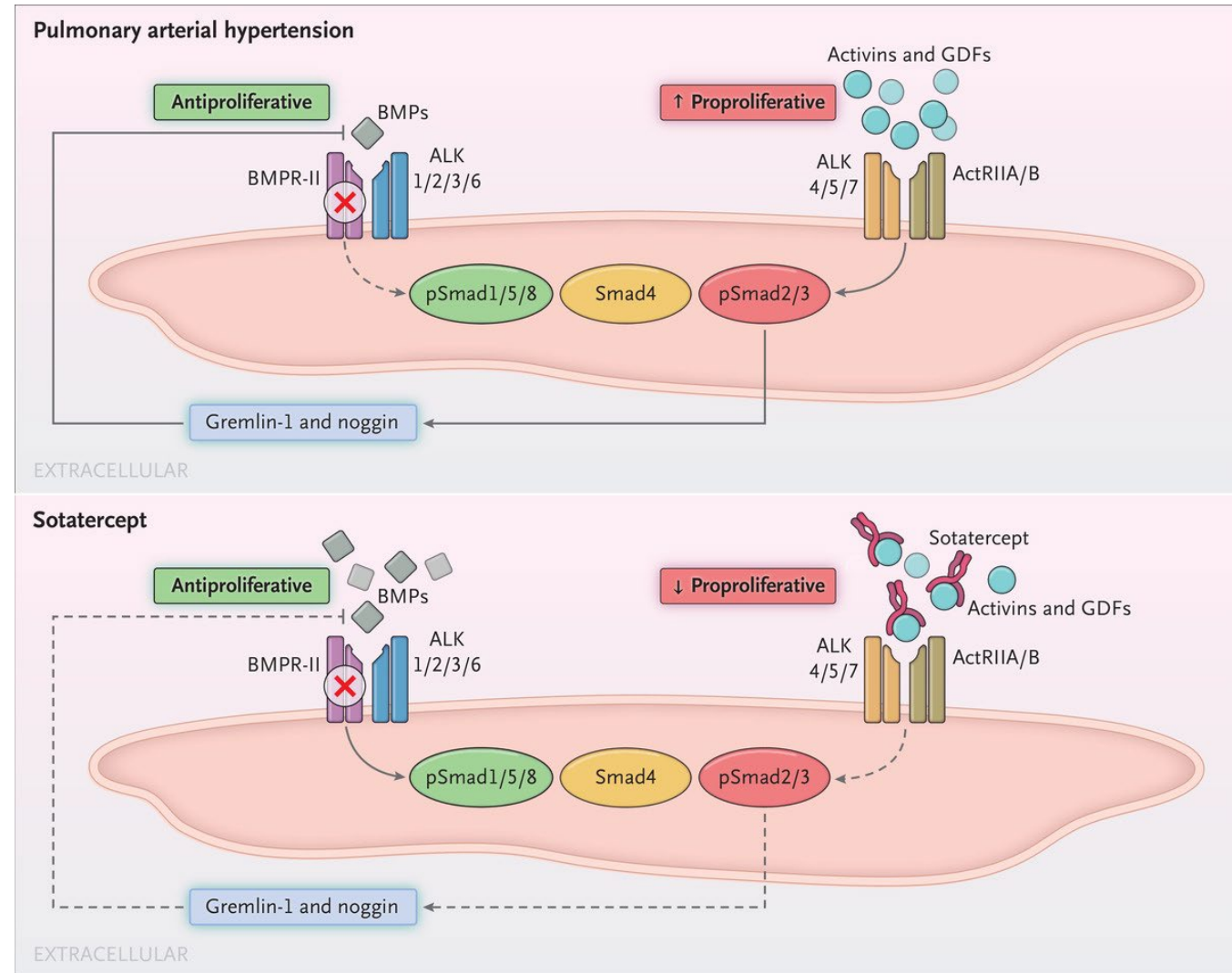


Pulmonary Arterial Hypertension

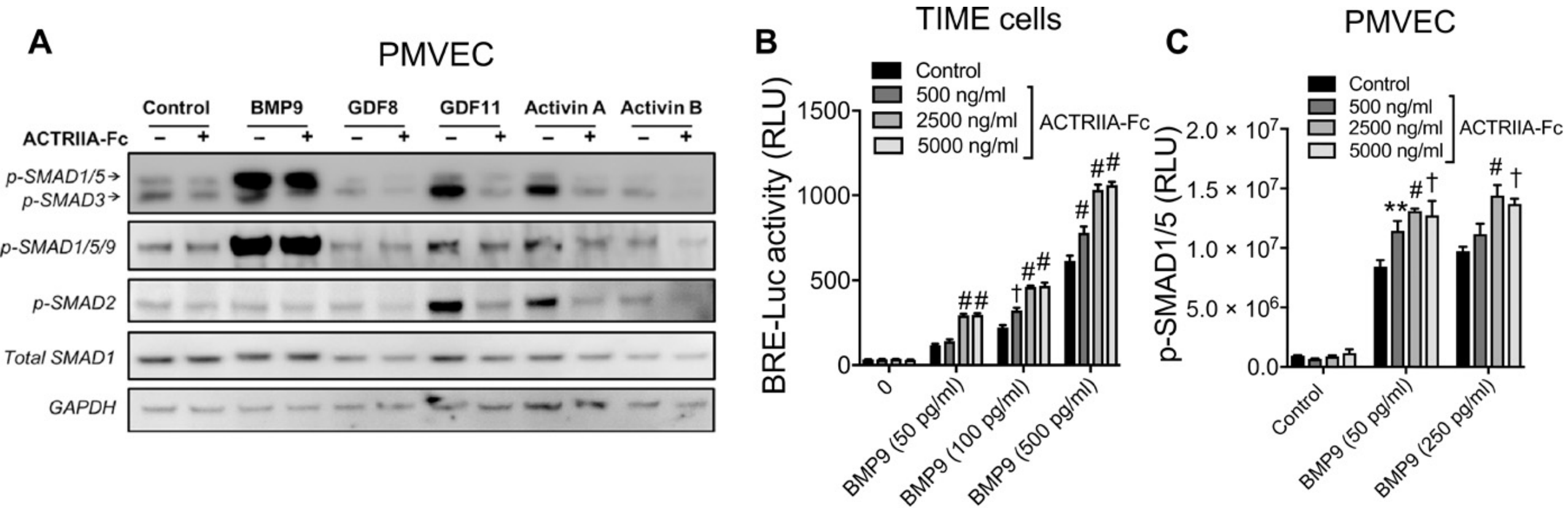
- Disease of the pulmonary vasculature
- Insidious onset - Frequently only symptom is shortness of breath
- Leads to progressive RV dysfunction, failure and ultimately death



Proposed Mechanism of Action for Sotatercept in Pulmonary Arterial Hypertension.



ACTRIIA-Fc modulates signaling, gene regulation, and proliferation of human pulmonary vascular cells.



Lai-Ming Yung et al., *Sci Transl Med* 2020;12:eaaz5660

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Change in Pulmonary Vascular Resistance from Baseline to Week 24 in the Intention-to-Treat Population.

